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Calibrated kallikrein generation in human plasma

D. Biltoft a,b,⁎, J.J. Sidelmann a,b, L.F. Olsen c, Y. Palarasah a,b, J. Gram a,b

a Unit for Thrombosis Research, Department of Public Health, University of Southern Denmark, Esbjerg, Denmark
b Department of Clinical Biochemistry, Hospital of South West Denmark, Esbjerg, Denmark
c Department of Biochemistry and Molecular Biology, University of Southern Denmark, Odense, Denmark

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Objectives: The physiological role of the contact system remains inconclusive. No obvious clinical complications have been observed for factor XII (FXII), prekallikrein (PK), or high molecular weight kininogen deficiencies even though the contact system in vitro is associated with coagulation, fibrinolysis, and inflammation. A global generation assay measuring the initial phase of the contact system could be a valuable tool for studies of its physiological role.

Design and methods: We investigated whether such a method could be developed using the principle of the Calibrated Automated Thrombin generation method as a template.

Results: A suitable kallikrein specific fluorogenic substrate was identified (K_M = 0.91 mM, k_cat = 19 s⁻¹), and kallikrein generation could be measured in undiluted plasma when silica was added as activator. Disturbing effects, including substrate depletion and the inner-filter effect, however, affected the signal. These problems were corrected by external calibration with α₂-macroglobulin-kallikrein complexes. Selectivity studies of the substrate, experiments with FXII and PK depleted plasmas, and plasma with high or low complement C1-esterase inhibitor activity indicated that the obtained and calibrated signal predominantly was related to FXII-dependent kallikrein activity.

Conclusions: The findings described show that establishment of a kallikrein generation method is possible. Potentially, this setup could be used for clinical studies of the contact system.

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1. Introduction

Four plasma proteins comprise the contact activation system [1]. In plasma, prekallikrein (PK) and factor XI (FXI) are mostly found in complex with the non-enzymatic co-factor high molecular weight kininogen (HMWK) [2,3]. Factor XII (FXII), on the other hand, circulates unbound. The contact system is initiated when encountered with activating surfaces. While the exact mechanism is unknown, the surface causes a conformational change in FXII rendering it susceptible for cleavage [4]. Once cleaved, activated FXII (FXIIa) can activate neighboring PK and FXI bound to the surface through HMWK. Amplification of the system follows from the ability of kallikrein to generate FXIIa [5]. Several protease inhibitors regulate the contact system. Complement C1-esterase inhibitor (C1-inhibitor) is predominating [6], but contributions from α₂-macroglobulin (α₂M) and other inhibitors have been observed [7,8]. In vitro experiments reveal connections between the contact system and coagulation, fibrinolysis, and inflammation. FXI serves as the link between contact activation and coagulation as its activation initiates the intrinsic pathway of coagulation and the formation of thrombin [9]. In vivo, FXII, PK, and HMWK deficiencies are not associated with bleeding tendencies, questioning the role of the contact system in normal haemostasis [1]. Recent studies, however, have shown that FXII deficient mice are protected from occlusive thrombi in experimental thrombosis models [10]. Other studies suggest inhibition of FXII and FXI as alternative anti-coagulant approaches [11]. Decreased or absent C1-inhibitor activity is associated with symptoms related to activation of inflammation [1].

A sensitive global activity assay measuring contact activation is currently not available, and in general, there is only a limited assortment of biomarker assays available to measure the contact system [12–17]. Studies of the connection between the contact system and clinical
conditions are therefore often hampered. A global generation test that measures the contact system may, potentially, facilitate such investigations.

The Calibrated Automated Thrombin generation method (CAT-method) is a global generation test that utilizes a fluorogenic substrate to measure the thrombin generation potential in human plasma after initiation of the extrinsic pathway of coagulation [18]. The CAT-method has proven to be a valuable tool to study disturbances of the coagulation system in vitro, and it shows potential for routine clinical application [19]. The use of a synthetic fluorogenic substrate to measure thrombin generation in undiluted plasma is, however, associated with several complications such as substrate depletion, the inner-filter effect, and cleavage of substrate by αM-thrombin [20]. In the CAT-method, these complications have been circumvented by unique approaches developed by Hemker and colleagues [18,21]. We investigated if the CAT-method could be used as a template for a method to measure contact activation through kallikrein generation after initiation of the contact system.

2. Materials and methods

2.1. Buffers

“BSA60 buffer” (20 mM Heps, 0.02% NaN₃, 60 g/L bovine serum albumin (BSA), pH = 7.4). “BSA5 buffer” (20 mM Heps, 0.02% NaN₃, 5 g/L BSA, pH = 7.4). “BSA-blank buffer” (20 mM Heps, 150 mM NaCl, 0.02% NaN₃, pH = 7.4). “Gel filtration buffer” (20 mM Heps, 300 mM NaCl, 0.02% NaN₃, pH = 7.4). “SDS sample buffer” (2 × Laemmli Sample Buffer (Bio-Rad, Hercules, California, U.S.)). “PBS-TW buffer” (Phosphate-buffered-saline (PBS), 0.05% Tween-20, pH = 7.4).

2.2. Plasma

Sodium citrate (0.109 M) and ethylenediaminetetraacetic acid (EDTA) stabilized plasma pools were obtained from 12 healthy individuals, who did not take hormone supplements. Plasma was frozen at –80 °C in 300 μL amouple. FXII depleted plasma (FXII DEP) was obtained from Haematologic Technologies Inc. (Essex Junction, Vermont, USA). Prekallikrein depleted plasma (PK DEP) was obtained from Affinity Biologicals (Ancaster, Ontario, Canada). EDTA stabilized plasma was obtained from two hereditary angioedema (HAE) patients.

2.3. Reagents and materials

Corn trypsin inhibitor (CTI), 2 mg/mL, was obtained from HTI. Soybean trypsin inhibitor (SBTI) was obtained from Sigma-Aldrich Denmark ApS (Sigma, Brøndby, Denmark). Twenty-five milligram of lyophilized SBTI was dissolved in 5 ml distilled water. Human complement C1-esterase inhibitor (C1-inhibitor) was obtained from CSL Behring (Lyngby, Denmark), dissolved in 10 mL H₂O, and dialyzed towards 5 L PBS at 4 °C for 72 h in a Spectra/Por® Dialysis Membrane molecular weight cut-off (MWCO) 6–8000 (Spectrumlabs®, Rancho Dominguez, Ca, U.S.).

Affinity purified kallikrein and FXIIa from human plasma were obtained from Enzyme Research Laboratories (Swansea, UK). One milligram of lyophilized protein was dissolved in 1 mL distilled water. Plasmin from human plasma was obtained from Sigma. One milligram of lyophilized protein was dissolved in 1 mL distilled water. Plasmin from human plasma was obtained from Sigma. One milligram of lyophilized protein was dissolved in 1 mL distilled water. Human FXIIa, 2.0 mg/mL in 50% glycerol, and human α-thrombin, 7.8 mg/mL in 50% glycerol, were obtained from HTI. BSA was from Sigma.

Kallikrein sensitive chromogenic substrate, S-2302, was obtained from Chromogenix (Mölndal, Sweden). Twenty-five milligram of substrate was dissolved to a concentration of 4 mM in distilled water. Fluorogenic substrate, OmniCathepsin fluorogenic substrate (Cbz-Phe-Arg-AMC), was obtained from Enzo Life Science (Exeter, UK). Fluorogenic substrate, Cbz-Pro-Phe-Arg-AMC, was synthesized on request by Chempeptide Limited (Shanghai, China). Fluorogenic substrate, Boc-Leu-Lys-Arg-AMC, was obtained from Bachem (Bubendorf, Switzerland). All fluorogenic substrates were dissolved in DMSO to a concentration of 50 mM (stock solutions).


APTT reagent, STA®-PTT Automate 5, was obtained from Triohab A/S (Brøndby, Denmark). The powder was dissolved in 5 mL BSA60 buffer (stock solution). Dextran sulfate sodium salt, with a molecular weight of 500 kDa, was obtained from Pharmacia AB (Uppsala, Sweden). Stock solution of dextran sulfate was prepared by dissolving 2 g of dextran sulfate in 1 L of distilled water. Colloidal silica LUDOX AS-40, a 40% (w/v) silica suspension in H₂O, was obtained from Sigma (stock solution). Polyethylene glycol with an average molecular weight of 6000 Da (PEG-6000) was obtained from Sigma.

Hirep 16/60 Sephacryl S-200 HR column (Sephacryl column) was obtained from GE Healthcare Europe GmbH (GE Healthcare, Brøndby, Denmark). Vivaspin 20™, 100 kDa MWCO, sample concentrator (Vivaspin sample concentrator) was obtained from GE Healthcare. Immulon 2HB round-bottom 96-well plate (2HB plate) was obtained from Thermo Scientific (Rochester, NY, U.S.). Fluoroskan Ascent™ Microplate Fluorometer (fluorometer) was obtained from Thermo Scientific. Thrombinoscope™ software was obtained from Thrombinoscope BV (Maastricht, Netherlands).

2.4. Fluorogenic substrates

The kinetic constants were determined essentially as described elsewhere [23]. The fluorogenic substrate Boc-Leu-Lys-Arg-AMC was diluted from stock solution in BSA60 buffer to obtain a final concentration of 1000, 800, 600, 400, 200, 100, 50, or 0 μM in the well. For Cbz-Pro-Phe-Arg-AMC and Cbz-Phe-Arg-AMC, the final substrate concentration was 2000, 1800, 1400, 1000, 800, 600, 400, or 0 μM in the well. The enzyme to be investigated was diluted from stock solution in BSA5 buffer. The final concentrations were 5.8 nM (kallikrein), 125 nM (FXIIa), 60.25 nM (plasmin), 272.5 nM (thrombin), or 3.125 nM (FXIa) in the well. α₂M-kallikrein was diluted 1:4 in BSA5 and added to the plate (final dilution of 1:24 in the well). The kinetic parameters were determined using the following setup; 100 μL of fluorogenic substrate was added to a 2HB plate and the plate was warmed for 5 min, at 37 °C, in the fluorometer. Twenty μL of enzyme solution (preheated to 37 °C) was then added using the fluorometer dispenser just before measurements were started. Each substrate concentration was run in quadruple and hydrolysis was followed for 40 min. Michaelis-Menten curves were fitted to the obtained data. Kcat was calculated from the determined Vmax, by first converting the fluorescence units per time to concentration per time, using a calibration curve, and then dividing this rate by the enzyme concentration.

2.5. Calibrator

2.5.1. Preparation of α₂M-kallikrein complexes

Human α₂M was obtained using a slight modification of a procedure described elsewhere [18,24]: To 1 vol of human plasma, 0.28 vol PEG-6000 (25% in H₂O) was added and the mixture was incubated for 30 min. After centrifugation (3000g), the precipitate was discarded and the supernatant was mixed with an additional 0.72 vol PEG-6000 (25% in H₂O). After 30 min incubation, the mixture was centrifuged (3000g) and the precipitate was dissolved in 0.1 vol BSA-blank buffer (“crude α₂M”). To prepare α₂M-kallikrein complexes, crude α₂M was diluted 1:8 in BSA-blank buffer and 100 μg/mL of kallikrein (final concentration) was added. The mixture was incubated overnight (ON).
of BSA5 together with 80 μL of citrate pool were added to a 2HB plate and the plate was warmed in the fluorometer to 37 °C. A start reagent for each activator was prepared by diluting the stock solutions of the activators in BSA60 buffer; Dextran sulfate was diluted to 600 μg/mL and silica to 6 mg/mL. The APTT reagent was not diluted. Lower concentrations of the activators were prepared by further dilutions in BSA60. Finally, the fluorogenic substrate was added to each reagent at a concentration of 2.5 mM. Twenty microliter of the solutions were then added to the plate and fluorescence was measured every 12 s for 20 min. Each activator concentration was run in duplicate. The final concentrations of the activators in the well were; dilutions 1:6, 1:12, 1:18, and 1:24 for the APTT reagent, 100, 50, 25, and 12.5 μg/mL for dextran sulfate, and 1, 0.5, 0.25, 0.125, or 0 mg/mL for silica.

2.7. Calculations

The raw data obtained from a generation experiment in a citrate pool (obtained as described in Section 2.8) were transferred to a Microsoft Excel spreadsheet. The corrected calibrator curve was found from a 6th-degree polynomial fit of the recorded calibrator data. The relation between the corrected and measured calibrator curves was calculated [18] and used to convert the obtained fluorescence units of the generation sample into corrected fluorescence units. The contribution from α2M-kallikrein was subtracted as described previously [25]. The fluorescence units of the corrected generation curve were converted to enzyme concentration by comparing them to the calibrator initial velocity. The calculations were also conducted using the Thrombinscope software for comparison.

2.8. Validation of kallikrein generation

Calibrated kallikrein generation was determined using four wells for each plasma sample tested. Two wells were used to determine kallikrein generation (generation wells) and two wells were used for calibration of signal (calibrator wells). To generation wells, 20 μL of BSA5 buffer containing either 90 μg/mL of PK-32, 90 μg/mL of nonsense antibody, 480 μg/mL of C1-inhibitor, or buffer alone was added. To calibrator wells, 20 μL of α2M-kallikrein, with an activity corresponding to 200 nM kallikrein, was added. Next, 80 μL of the plasma samples to be tested were added to the wells and the plate was warmed to 37 °C. The following plasmas were tested: Citrate pool, FXII DEP, PK DEP, HAE plasma, and mixtures of the citrate pool and FXII DEP consisting of 50%, 75%, and 90% FXII DEP. Immediately before initiation of experiment, colloidal silica and Boc-Leu-Lys-Arg-AMC were added to BSA60 buffer preheated to 37 °C. Twenty microliter of the solution was added to the wells. The final concentrations of silica and substrate were 0.24 mg/mL and 0.42 mM, respectively. Fluorescence was measured every 12 s for 25 min and calculations were conducted using the Thrombinscope software.

3. Results

3.1. Fluorogenic substrate

The Michaelis-Menten kinetics of three fluorogenic substrates for kallikrein [26] were determined in BSA60 buffer and the results are listed in Table 1. Boc-Leu-Lys-Arg-AMC displayed lower \( k_{\text{cat}} \) and higher \( k_{\text{m}} \) values compared to other fluorogenic substrates.

### Table 1

<table>
<thead>
<tr>
<th>Peptide sequence</th>
<th>( k_{\text{m}} ) (μM)</th>
<th>( k_{\text{cat}} ) (s(^{-1}))</th>
<th>( k_{\text{cat}}/k_{\text{m}} ) (M(^{-1})s(^{-1}))</th>
</tr>
</thead>
<tbody>
<tr>
<td>Chz-Phe-Arg-AMC</td>
<td>0.83 (0.68–1.1)</td>
<td>33 (31–37)</td>
<td>4.0 · 10(^4)</td>
</tr>
<tr>
<td>Chz-Pro-Phe-Arg-AMC</td>
<td>1.4 (0.99–1.5)</td>
<td>45 (39–47)</td>
<td>3.2 · 10(^4)</td>
</tr>
<tr>
<td>Boc-Leu-Lys-Arg-AMC</td>
<td>0.91 (0.84–0.96)</td>
<td>19 (18–20)</td>
<td>2.1 · 10(^4)</td>
</tr>
</tbody>
</table>

The results are presented as median and range of quadruple determinations performed in buffer with high albumin concentration. ND: not determined.
(19 s\(^{-1}\)) for kallikrein compared to Cbz-Phe-Arg-AMC (33 s\(^{-1}\)) and Cbz-Pro-Phe-Arg-AMC (45 s\(^{-1}\)). Cbz-Pro-Phe-Arg-AMC displayed the lowest binding affinity with an apparent KM value of 1.4 mM whereas Boc-Leu-Lys-Arg-AMC and Cbz-Phe-Arg-AMC showed similar binding affinities with apparent KM values of 0.91 and 0.83 mM, respectively. With the aim of obtaining a substrate with a low k\(_{cat}\) and a high KM [27], Boc-Leu-Lys-Arg-AMC was selected for further studies.

In BSA60 buffer, the kinetic constants of the substrate Boc-Leu-Lys-Arg-AMC for other contact system related serine proteases in plasma were investigated. In Table 2, these constants are compared with those obtained for kallikrein. For FXIa, a KM of 3.4 mM and a k\(_{cat}/KM\) of 8.2 \(\times 10^3\) M\(^{-1}\) s\(^{-1}\) were observed. For FXIIa and \(\alpha\)-thrombin, KM values of 0.92 mM and 0.74 mM, respectively, were found with a k\(_{cat}/KM\) for both enzymes of around 30 M\(^{-1}\) s\(^{-1}\). Plasmin showed KM and k\(_{cat}/KM\) values of 3.8 mM and 974 M\(^{-1}\) s\(^{-1}\), respectively.

3.2. Calibrator

A western blot (Fig. 1) demonstrated assembly of \(\alpha_2M\)-kallikrein complexes. A kallikrein related high molecular weight band was observed in the lane containing a mixture of “crude \(\alpha_2M\)" and kallikrein (lane 3). This band was not present in lanes including only kallikrein (lane 1) or "crude \(\alpha_2M\)" (lane 2). After gel filtration, fractions containing only the kallikrein related high-molecular weight protein were obtained (lane 4).

In plasma, the initial amidolytic activity of \(\alpha_2M\)-kallikrein towards the substrate, Boc-Leu-Lys-Arg-AMC, was constant for at least 1 h (data not shown). The data confirm that \(\alpha_2M\)-kallikrein complexes...
possess enzymatic activity towards small substrates that are protected from other inhibitors in plasma.

In BSA60 buffer, the apparent $K_M$ of $\alpha_2M$-kallikrein towards Boc-Leu-Lys-Arg-AMC was determined to 0.58 mM. This is somewhat lower than the apparent $K_M$ of free kallikrein (0.91 mM).

3.3. Activating reagent

The potential of different activators to generate a measureable signal in the current setup is shown in Fig. 2. The results demonstrate a low activation potential of the APTT reagent (Fig. 2A). Both dextran sulfate (Fig. 2B) and colloidal silica (Fig. 2C) were potent activators. However, substantial turbidity of the sample followed the use of dextran sulfate indicating precipitation of proteins in the mixture (data not shown). This was not the case when silica was used as activator. No signal generation was observed in the absence of activator (Fig. 2C, dashed black line).

3.4. Calculations

The recorded fluorescence of the calibrator demonstrated a decreasing reaction velocity at fixed enzyme concentration (Fig. 3A, black line). The "correct" calibrator curve was determined (Fig. 3A, dotted line), and the relation between the corrected and the observed calibrator curve was found (Fig. 3B). Application of this relation to the raw generation data (Fig. 3C, dotted line) resulted in a steady end-level signal (Fig. 3C, black line). Measurements of substrate cleavage by $\alpha_2M$-kallikrein, formed during the generation, were then subtracted (Fig. 3D, black line). Finally, the fluorescence units were converted to enzyme concentration (Fig. 3E, grey dotted line). This curve was identical to the curve obtained when the calculations were performed by the Thrombinscope software (Fig. 3E, black dotted line).

3.5. Validation of kallikrein generation

Fluorescence development was absent in plasmas depleted of FXII or PK while signal generation was recorded in a citrate pool (Fig. 4A). Addition of C1-inhibitor to a citrate pool diminished the fluorescence generated considerably compared to a citrate pool without added C1-inhibitor (Fig. 4B). PK-32 also diminished the generation signal whereas a very modest effect was seen when a nonsense antibody was added to the plasma (Fig. 4B). Furthermore, the signal obtained declined with decreasing levels of FXII (Fig. 4C). A substantially higher signal

![Fig. 3. Application of the mathematical algorithms to correct for the effects from substrate depletion, the inner-filter effects, and cleavage of substrate by $\alpha_2M$-kallikrein. The raw data were obtained from a generation experiment in a citrate pool with silica as activator (0.24 mg/mL, final concentration), Boc-Leu-Lys-Arg-AMC (0.42 mM, final concentration) as substrate, and $\alpha_2M$-kallikrein as calibrator (33.3 nM kallikrein-like activity, final concentration). A: The recorded fluorescence from the calibrator sample (black line) and the "corrected" calibrator curve (dotted line), which was calculated from a 6th-degree polynomial fit of the recorded fluorescence. B: Plot of the raw (measured) data versus the corrected calibrator data. C: First derivate of the raw data from the generation sample (dotted line) and the "corrected" generation data (black line), which was calculated using the relation observed in B. D: The corrected generation data before (dotted line) and after (black line) subtraction of $\alpha_2M$-kallikrein contribution. E: The final curve as obtained by manual calculations (grey dotted line) and by the Thrombinscope software (black dotted line).]
generation was observed in plasmas with decreased C1-inhibitor activity (HAE plasma) compared to the EDTA pool (Fig. 4D).

4. Discussion

In this article we present a series of experiments demonstrating a possible setup for measuring FXII-dependent kallikrein generation in undiluted plasma. The results obtained by the current method are calibrated, thus the final results represent generation of kallikrein activity over time.

Michaelis-Menten kinetics of three fluorogenic substrates for kallikrein [26] were investigated to determine whether they were suitable for continuous generation experiments, i.e. display high $K_M$ and low $k_{cat}$ values [27]. As an effect of albumin on the hydrolysis of a fluorogenic substrate has been reported previously [28], the kinetic constants were determined in buffer with an albumin concentration similar to that of plasma. Under these circumstances, Boc-Leu-Lys-Arg-AMC displayed favorable kinetics for generation experiments with a high $K_M$ (0.91 mM) and a low $k_{cat}$ (19 s$^{-1}$) with respect to kallikrein. Cbz-Pro-Phe-Arg-AMC and Cbz-Phe-Arg-AMC showed $K_M$ values of 1.4 and 0.83 mM, respectively, and considerably higher $k_{cat}$ values (45 and 33 s$^{-1}$, respectively). Furthermore, at low substrate concentrations, a sigmoidal curvature of initial rate versus substrate concentration was observed for both Cbz-Pro-Phe-Arg-AMC and Cbz-Phe-Arg-AMC (data not shown). Because albumin is abundant in plasma, the use of Cbz-Pro-Phe-Arg-AMC or Cbz-Phe-Arg-AMC may be accompanied by significant complications in the current setup.

The selectivity of Boc-Leu-Lys-Arg-AMC for plasma kallikrein compared to other serine proteases is of major importance. The selectivity of the substrate was investigated by comparing the kinetic constants for kallikrein, FXIIa, FXIa, thrombin, and plasmin (Table 2). The substrate demonstrated higher $K_M$ for FXIIa and plasmin than for kallikrein but comparable or lower $K_M$ for FXIa and thrombin. However, very low $k_{cat}/K_M$ values were observed for both FXIIa and thrombin compared to kallikrein, and a significant contribution from these enzymes upon signal generation is therefore unlikely. In addition, many of the proteases are present in plasma in much lower concentrations than that of prekallikrein. For example, the plasma concentration of FXI is around twenty times lower than that of prekallikrein [16]. Even though a two-fold difference between the $k_{cat}/K_M$ for kallikrein and FXIa was recorded, the signal development from cleavage by FXIa will be minimal [21]. Furthermore, the initial phase of contact activation does not require presence of calcium. Consequently, re-calciﬁcation is omitted securing no or very little activation of calcium dependent proteases.

The enzymatic capacity of the $\alpha_2M$-thrombin complex, used as calibrator in the CAT-method [18], is not affected by inhibitors in plasma but retains the ability to cleave small substrates. $\alpha_2M$-kallikrein was likewise unaffected by inhibitors in plasma as no decrease in activity was observed after incubation in plasma for 1 h. It is preferable that the affinity of the substrate for kallikrein and $\alpha_2M$-kallikrein is similar for calibration. However, complex formation between kallikrein and $\alpha_2M$ affected the kinetic constant $K_M$, which appeared lower towards the substrate when kallikrein was bound to $\alpha_2M$. It is peculiar that interaction between the enzyme and an inhibitor result in an apparently improved binding affinity (i.e. lower $K_M$) of the substrate. However, this finding is in accordance with previous reports, in which a similar trend was observed for human $\alpha_2M$-kallikrein [29], $\alpha_2M$-trypsin [30], and $\alpha_2M$-thrombin [31] complexes. The effect of difference in $K_M$ upon calibration was not investigated here.

Polyphosphates [32] and misfolded proteins [33] have been proposed as physiological activators of the contact system, but the subject is still under study. Other assays relying on contact activation, such as the APTT and contact dependent thrombin generation, employ non-physiological activators. In light of this, we investigated several of the known non-physiological activators in the current setup. When the APTT reagent was used as activator, a moderate signal was observed. The signal diminished dose-dependently with lower concentration of APTT reagent. At high concentrations, both dextran sulfate and colloidal silica generated a stronger signal than the APTT reagent. The
use of dextran sulfate, however, resulted in significant turbidity, which presumably is caused by dextran sulfate induced precipitation of proteins. This was not observed when silica was used as the activator.

Variation in pre-analytical handling of samples can affect test consistency [34,35]. While the effects of pre-analytical factors were not investigated in the present study, we did not record any signal development in the absence of contact activator. These findings indicate that the proposed method is not affected by the surface of the microtiter plate. Furthermore, the findings also indicate that the reported activity of PK-HMWK complexes [36] is not detected by the current method.

The current analytical setup does, however, suffer from other obstacles seen in the CAT-method including effects of substrate depletion, the inner-filter effect, and cleavage of substrate by the formation of α-M-enzyme complexes. In the CAT-method, mathematical algorithms are used to circumvent these obstacles and such algorithms should be applicable in the current setup as well. The manual calculations presented here showed that the algorithms could be used to overcome these complications.

The validation experiments strongly indicated that the observed signal was related to FXII-driven kallikrein generation. We observed no signal in plasma depleted of FXII or PK confirming that the presence of these proteins is required. Furthermore, adding citrate pool to the FXII DEP dose-dependently restored signal generation. This indicates a considerable effect from plasma levels of FXII. The amount of C1-inhibitor considerably affected by levels of FXII, PK, and C1-inhibitor. Consequently, the experiments show that such a method could be useful to study of a functional, sensitive, and accurate global assay for measurements of high-molecular-weight kininogen cleavage correlates with diseases in the bradykinin-mediated angioedema due to hereditary C1-inhibitor deficiency, Clin. Exp. Allergy 44 (2014) 1503–1514.


