



University of Southern Denmark

Phyto-oestrogens in herbage and milk from cows grazing white clover, red clover, lucerne or chicory-rich pastures

Andersen, C; Nielsen, Tina S.; Purup, Stig; Kristensen, Troels; Eriksen, Jørgen; Søegaard, Karen; Sørensen, J.; Fretté, Xavier

Published in:
Animal

DOI:
10.1017/S1751731109004613

Publication date:
2009

Document version:
Submitted manuscript

Citation for pulished version (APA):

Andersen, C., Nielsen, T. S., Purup, S., Kristensen, T., Eriksen, J., Søegaard, K., Sørensen, J., & Fretté, X. (2009). Phyto-oestrogens in herbage and milk from cows grazing white clover, red clover, lucerne or chicory-rich pastures. *Animal*, 3(8), 1189-1195. <https://doi.org/10.1017/S1751731109004613>

Go to publication entry in University of Southern Denmark's Research Portal

Terms of use

This work is brought to you by the University of Southern Denmark.

Unless otherwise specified it has been shared according to the terms for self-archiving.

If no other license is stated, these terms apply:

- You may download this work for personal use only.
- You may not further distribute the material or use it for any profit-making activity or commercial gain
- You may freely distribute the URL identifying this open access version

If you believe that this document breaches copyright please contact us providing details and we will investigate your claim. Please direct all enquiries to puresupport@bib.sdu.dk

Phyto-oestrogens in herbage and milk from cows grazing white clover, red clover, lucerne or chicory-rich pastures

C. Andersen^{1a}, T. S. Nielsen¹, S. Purup^{1†}, T. Kristensen², J. Eriksen², K. Søgaard², J. Sørensen³ and X. C. Fretté³

¹Department of Animal Health, Welfare and Nutrition, Faculty of Agricultural Sciences, University of Aarhus, Blichers Allé 20, Tjele, Denmark; ²Department of Agroecology and Environment, Faculty of Agricultural Sciences, University of Aarhus, Blichers Allé 20, Tjele, Denmark; ³Department of Food Science, Faculty of Agricultural Sciences, University of Aarhus, Blichers Allé 20, Tjele, Denmark

(Received 28 October 2008; Accepted 26 March 2009; First published online 24 April 2009)

A grazing experiment was carried out to study the concentration of phyto-oestrogens in herbage for cattle and in milk during two periods (May and June). Forty-eight Danish Holstein cows were divided into four groups with four treatment diets; white clover, red clover, lucerne and chicory-rich pastures. Each experimental period lasted 15 days. Herbage samples from the first day and individual milk samples from the last day of the experimental period were analysed for phyto-oestrogens using LC-MS technique. The total concentration of phyto-oestrogens was 21 399 mg/kg dry matter (DM) for red clover and 238 to 466 mg/kg DM for the other three herbages mainly due to a much higher concentration of biochanin A, formononetin and glycitein in red clover. In the milk, the total concentration of phyto-oestrogens was 253 to 397 µg/l for red clover milk and 56 to 91 µg/l in the milk from the other three treatments. This was especially due to a higher concentration of equol, daidzein and formononetin in the red clover milk. The concentration of biochanin A was significantly higher in milk from the red clover treatment in May while no differences were observed in June. Enterodiol was similar across treatments while the concentration of enterolactone was significantly lower for red clover milk compared with the other treatments. Of the tested pastures, red clover appears to have the highest concentration and to be the best source of phyto-oestrogens, especially equol, in bovine milk.

Keywords: phyto-oestrogens, bovine milk, pasture

Implications

We have documented that the milk content of specific potential health-affecting compounds may be manipulated in a predicted direction by diet composition. This implicates that milk can be designed to contain defined levels of specific compounds transferred from the diet. Examples of this are that phyto-oestrogens are transferred to milk when cows are fed high amounts of leguminous plants like clover.

Introduction

In order to increase the sale of organic milk, new specially produced organic milk types with improved taste and potential health benefits are developed. This can be done through feeding of cows with legumes or herbs containing flavour and components with possible positive impacts

on health that are transferred to the milk. As for health components phyto-oestrogens are of interest.

Phyto-oestrogens are a large group of naturally occurring non-steroidal plant-derived compounds with a diverse structure. Phyto-oestrogens have been shown to behave as weak oestrogen agonists/antagonists in both animals and humans (Benassayag *et al.*, 2002) and studies in humans, animals and cell cultures suggest that dietary phyto-oestrogens may play an important role in the prevention of menopausal symptoms, osteoporosis, hormone-dependent cancers and heart disease (Kurzer and Xu, 1997). Hence, milk with an increased content of phyto-oestrogens could be of significant interest.

Cows mainly ingest phyto-oestrogens from legumes. Among the legumes, red clover has the highest total concentration of phyto-oestrogens varying from 1% to 2.5% of dry matter (DM), while the concentration of phyto-oestrogens in white clover is 0.02% to 0.06% of DM (Saloniemi *et al.*, 1995). Both red and white clovers primarily contain isoflavones such as formononetin, genistein, daidzein and

^a Present address: Faculty of Life Sciences, University of Copenhagen, Bülowsvej 17, Frederiksberg, Denmark.

[†] E-mail: Stig.Purup@agrsci.dk

biochanin A (Steinshamn *et al.*, 2008). Lucerne only contains small quantities of isoflavones (Saloniemi *et al.*, 1995). Lignans are primarily found in cereals, legumes and oilseed used in concentrates (Thompson *et al.*, 1991), and occur in somewhat higher concentrations in white clover compared to red clover (Steinshamn *et al.*, 2008).

In bovine milk, the concentration of the isoflavones, formononetin, biochanin A, daidzein and genistein, have been found to range from 0.1 to 7.7 $\mu\text{g/l}$, the concentration of the isoflavone metabolite equol between 45 and 364 $\mu\text{g/l}$ and the lignan enterolactone between 19 and 96 $\mu\text{g/l}$ (Antignac *et al.*, 2004; Steinshamn *et al.*, 2008; Andersen *et al.*, 2009). In general, organic milk has a higher concentration of isoflavones than conventionally produced milk due to the more widespread use of leguminous plants in organic feeding (Antignac *et al.*, 2004; Purup *et al.*, 2005; Hoikkala *et al.*, 2007). Steinshamn *et al.* (2008) found that milk from cows fed red clover silage had a significantly and several times higher concentration of isoflavones compared to milk from cows fed white clover silage. For example, the concentration of biochanin A, equol and formononetin in the milk was on average 5.0, 4.3 and 2.4 times higher, respectively, in cows fed red clover *v.* white clover silage. In contrast, milk from cows fed white clover silage had the highest concentration of the lignans enterodiol and enterolactone (on average 1.75 and 1.37 times higher, respectively). The effects of silage type on the concentration of the individual phyto-oestrogens in the milk were related to the intake of the compound or its precursor, which implies that it is possible to influence the concentration of the phyto-oestrogens in bovine milk. However, there is limited knowledge on the concentration of phyto-oestrogens in milk from cows on different diets and at different time periods throughout the year.

In this study, the concentrations of phyto-oestrogens in milk from cows grazing red clover, white clover, lucerne or chicory-rich swards were measured during two summer months. Our hypothesis was that the milk content of phytoestrogens can be manipulated by diet composition. No literature exists on phyto-oestrogens in chicory, but chicory was included in the study due to its flavour components. However, only the concentration of phyto-oestrogens is included in this article as the taste of the milk is reported elsewhere.

Material and methods

Pasture

A grazing study was performed in the summer 2006 at the organic research station Rugballegaard (55°52'N, 9°47'E), Denmark, with four treatment diets: white clover, red clover, lucerne and chicory-rich swards.

Swards with lucerne (*Medicago sativa* L., Pondus), red clover (*Trifolium pratense* L., Rajah), white clover (*Trifolium repens* L., Milo) and white clover together with chicory (*Cichorium intybus* L., Puna), respectively, were established in 2005 together with perennial ryegrass (*Lolium perenne* L.), 35% Calibre (medium tetraploid), 30% Sameba (late diploid)

and 35% Tivoli (late tetraploid) in two replicate paddocks. The swards were unfertilized and irrigated at high drought stress. The paddocks were approximately 1.8 ha each (see Eriksen *et al.* (2007) for further details).

Before grazing, the area in each paddock was adjusted with a fence to ensure the same amount of herbage per cow in all paddocks. As little social contact as possible between the groups was ensured during allocation to the paddocks.

The cows were on pasture 20 h daily, supplemented with 6.2 kg DM/cow per day (oats 82%, hay 16%, mineral mix 2%), fed twice daily after milking (0500 h and 1600 h).

Before each experimental period the cows grazed a traditional white clover/perennial ryegrass field, and from five days before the start of each period, cows were fed the experimental concentrates in the stable.

Animals and experimental design

Forty-eight lactating Holstein dairy cows, 140 ± 98 days in milk (mean \pm s.d.), with milk yield 30.4 ± 5.7 kg energy corrected milk were, before each of the two periods, blocked according to milk yield and parity (1, 2 and >2), randomly within block allocated to one of the four treatments. The cows grazed the paddocks in two periods (May and June). Each period lasted 14 days with start on day 1 after morning milking and finishing on day 15 after morning milking. The cows grazed the two statistical replicates in relation to crop production alternately with one day in each paddock.

Data collection

Individual milk samples were collected on day 15 during morning milking in the experimental period. One sample from June was missing. The milk samples were frozen immediately after collection and stored at -20°C until analysis for phyto-oestrogens.

Individual milk yield was measured and concentration of fat and protein analysed three times during the period. Sward productivity was estimated indirectly in an area fenced off during the period. In the beginning and after one week of grazing the herbage mass and the botanical composition were determined in 0.5 m^2 samples in the grazed area and in the fenced area (Eriksen *et al.*, 2007).

Prior to each grazing period, samples of herbage were collected by tearing off plant parts above stubble height (5 to 6 cm) by hand. This was to get a sample more representative for animal intake than the samples cut at the soil surface. Approximately 2 kg of fresh plant material was sampled. A sub-sample of 1 kg was frozen at -20°C within 15 min and used for analysis of phyto-oestrogens. Botanical composition was assumed similar in both samples. The stubble height was approximated to the lowest grazing height of the cows in the individual paddocks. Samples were collected separately in the two paddocks. The botanical composition was determined by hand separation in sub-samples, drying and weighing.

Registrations made on herd and sward productivity and botanical composition for May and June, respectively, are

Table 1 Proportion of the test species in the sward and herbage mass at the beginning of each period, and growth rate during the first week of the 2-week experimental period

Sward type	Proportion of test species in the sward (% of DM)		Herbage mass (kg DM/ha)		Herbage growth rate (kg DM/ha per day)	
	May	June	May	June	May	June
White clover	60	46	1799	1670	134	109
Red clover	68	47	2383	2178	82	122
Lucerne	27	12	2476	1249	106	124
Chicory	72	52	1738	1609	123	102

DM = dry matter.

Samples were collected from two paddocks and the botanical composition was determined by hand separation in sub-samples and weighing.

shown in Table 1. Generally, the proportion of legumes and chicory was high in the mixtures. However, the proportion of lucerne in both periods was considerably lower than the other three mixtures and only 12% in June caused by a too short rest period from May. The proportion of clover, lucerne and chicory was considerably higher in May compared with June.

Analysis of phyto-oestrogens in herbage and milk samples

Herbage mixture samples from the first day of the experimental period were analysed in single measurements and milk samples were analysed in duplicate for the concentration of chrysin, naringenin, biochanin A, formononetin and glycitein. The milk samples were also analysed for the concentration of daidzein, equol, enterodiol and enterolactone. Standards for daidzein, naringenin, genistein, apigenin and formononetin were purchased from Extrasynthese (Genay, France). All other standards were purchased from Sigma-Aldrich (Brøndby, Denmark).

The flavonoids in the herbage mixture samples were extracted by dipping 50 g frozen aerial parts of pasture sample in liquid nitrogen and immediately placing it in a kitchen blender, resulting in a fine (particle size < 0.5 mm) homogeneous powder. One gram of this powdered grass sample was homogenized in a centrifuge tube (40 ml) for 1 min together with 20 ml 80% aqueous MeOH followed by extraction for 2 h at room temperature. After extraction, the sample was centrifuged (2700 × g for 10 min) and the precipitate discarded. The supernatant was then evaporated to dryness under nitrogen stream. The dried extract was re-solubilized in distilled water and was added 100 µl of β-glucuronidase. The mixture was placed in an oven at 37°C for 1 h and shaken vigorously every 15 min. Post-processing of grass samples after enzymatic hydrolysis is similar to milk samples.

Milk samples (4 ml) were equilibrated to room temperature before addition of 25 µl prunetin (120 ng/ml) as internal standard and 100 µl β-glucuronidase in order to extract aglycones of phyto-oestrogens. The mixture was placed in an oven at 37°C for 1 h and shaken vigorously every 15 min. After centrifugation (10 min at 2700 × g), both the creamy layer and the precipitate were discarded, while the liquid phase was recovered and submitted to solvent extraction with hexane (3 × 3 ml) followed by ethyl acetate (3 × 3 ml). The ethyl acetate extract was evaporated

to dryness under a flow of nitrogen. The residue was then re-suspended in 500 µl acetonitrile, filtered in Mini-Uniprep Amber 0.2 µm PTFE (polytetrafluoroethylene) vials from Whatman, VWR International ApS (Herlev, Denmark) and analysed by LC-MS.

Liquid Chromatography-Mass Spectrometry analyses were performed on an Agilent (Waldbronn, Germany) HPLC-DAD-MS station equipped with an HPLC series 1100 comprising a model G1312A binary pump, a model G1379A micro vacuum degasser, a model G1327A thermostated autosampler, a model G1316A thermo stated column compartment, a model G1315B diode-array detector and a model G2707DA LC/MSD SL detector fitted with a model G1948A atmospheric pressure electrospray ionization source (AP-ESI). The station was controlled and the results were monitored with Agilent's ChemStation software (Rev. A.10.02).

Sample separations were carried out on a Purospher STAR RP-18e column (Merck, Darmstadt, Germany), 250 × 4.0 mm i.d., 5 µm particle size, operated at a temperature of 35°C and with a 0.5 ml/min flow. The solvents used were (A) aqueous 1% formic acid and (B) 1% formic acid in acetonitrile, using a linear gradient programme as follows: 55% B isocratic (10 min), 55% B to 70% B (5 min), 70% B isocratic (5 min), 70% B to 98% B (5 min), 98% B isocratic (5 min), 98% B to 55% B (1 min), 55% B isocratic (10 min). The total time of a run thus was 41 min. The injection volume was 20 µl.

UV spectra were recorded in the range 200 to 600 nm at a rate of 1.25 scans/s. Simultaneously selected wavelengths were monitored separately at 200, 260, 300 and 360 nm. MS spectra of samples were recorded in positive (ESI (electrospray ionisation)) SIM (selected ion-monitoring) mode. The acquisition parameters were as follows: fragmentor 70 V, gain 1 electron multiplier voltage; Spray chamber parameters; Nitrogen was used as drying gas at a flow of 9 l/min and as mobilizing gas at a pressure of 290 kPa and a temperature of 315°C. A potential of +3000 V was used on the capillary.

Retention times (RT), regression equations, R^2 , recovery % and values for limit of quantitation (LOQ) are shown in Table 2. LOQ was calculated according to Simonsen (2006).

Unfortunately, the MS peak for ion m/z 271 (ion generated in API-ES pos. (atmospheric pressure interface-electrospray positive) for genistein molecular weight (MW) 270) with the RT of genistein actually received the contribution of another

Table 2 Retention times (RT), regression equations (slope and intercept), R^2 , recovery (%) and limit of quantitation (LOQ) for analyses of milk samples

Phyto-oestrogen	RT (min)	Intercept	Slope	R^2	Recovery (%)	LOQ ($\mu\text{g/l}$)
Chrysin	12.898	-3242.149	619 824.5	0.9991	73.76	0.029
Naringenin	6.751	-14 004.52	94 464.97	0.9979	76.28	0.700
Biochanin A	14	3538.621	86 698.41	0.9916	68.94	0.065
Daidzein	5.517	16 301.51	855 344.6	0.9931	78.40	0.006
Equol	7.073	-451.8333	6471.807	0.9991	69.58	0.272
Formononetin	9.08	10 225.36	126 204.2	0.9979	91.79	0.160
Glycitein	5.726	-30 338.32	173 167.5	0.9991	68.24	0.170
Enterodiol	5.215	1593.116	108 556.1	0.9994	83.68	0.043
Enterolactone	7.077	25 672.9	55 586.65	0.9911	95.01	0.763

Milk samples were analysed in duplicate for the concentration of phyto-oestrogens.

Table 3 Effect of dietary treatment on milk yield, fat and protein from 48 lactating Holstein dairy cows in two 15-day periods in May and June, respectively

	May				June				P-values		
	White clover	Red clover	Lucerne	Chicory	White clover	Red clover	Lucerne	Chicory	Diet	Period	Diet \times Period
Milk (kg ECM per day)	31.3	30.1	31.4	33.9	30.9	30.3	31.2	30.1	0.76	0.37	0.60
Fat (g/kg milk)	3.62	3.96	3.79	4.12	3.82	3.68	3.66	3.77	0.48	0.26	0.40
Protein (g/kg milk)	3.39	3.46	3.33	3.34	3.36	3.27	3.33	3.33	0.94	0.35	0.63

ECM = energy corrected milk.

P-values for the effect of diet, period and their interaction are shown.

compound with the same MW, probably apigenin. Therefore, it was not possible to measure the concentration of genistein. Furthermore, the concentration of daidzein could not be measured in herbage samples.

Statistical analysis

All data were analysed using the GLM procedure of SAS (SAS Institute, 1999). The following statistical model was used:

$$Y_{ijkl} = \mu + \alpha_i + \beta_j + (\alpha \times \beta)_{ij} + \varepsilon_{ijkl},$$

where: Y = dependent variable, μ = mean, α = fixed effect of period i (May, June), β = fixed effect of diet j (white clover, red clover, lucerne, chicory); random residual variation $\varepsilon_{ijkl} \sim N(0, \sigma^2)$. For the analysis of phyto-oestrogens in herbage mixtures the fixed effect of period was excluded. The results are presented as least squares means.

Results

Milk yield

The milk yield, fat and protein concentration, reported in Table 3, show no overall effect of dietary treatment or period.

Phyto-oestrogens in herbage samples

The concentrations of phyto-oestrogens in the herbage are reported in Table 4. The total concentration of phyto-oestrogens was 45 to 90 times higher in red clover mixture

Table 4 Concentration of phyto-oestrogens (mg/kg DM) in pasture mixtures in relation to dietary treatments

Phyto-oestrogen	White clover	Red clover	Lucerne	Chicory	P-value
Chrysin	3.5 ^a	4.5 ^a	2.5 ^a	64.5 ^a	0.29
Naringenin	31.5 ^a	175.5 ^b	87.0 ^a	72.5 ^a	0.0097
Biochanin A	17.0 ^a	888 ^b	11.5 ^a	3.5 ^a	0.0009
Formononetin	405 ^a	11 420 ^b	156 ^a	60 ^a	0.0001
Glycitein	8.5 ^a	913 ^b	6.0 ^a	37.5 ^a	0.0047
Total concentration	466 ^a	21 399 ^b	263 ^a	237.5 ^a	0.0002

DM = dry matter.

Samples were obtained from May and June.

P-values for the effect of diet are shown. The superscript symbols a, b designate significant difference ($P < 0.05$) between dietary treatments.

compared to the other three herbage mixtures. In particular, the concentration of naringenin, biochanin A, formononetin and glycitein was significantly higher in red clover mixture than in the other herbage mixtures. However, although not significant, the concentration of chrysin was many times higher in the chicory mixture. The total level of phyto-oestrogens in the other three herbage mixtures was similar. In the red clover mixture both biochanin A and formononetin occurred in high concentrations compared to the other phyto-oestrogens. In the white clover mixture the by-far most quantitative phyto-oestrogen was formononetin. In the lucerne and chicory mixtures none of the phyto-oestrogens occurred in relatively high concentrations compared with the mixtures.

Phyto-oestrogens in milk samples

The concentration of phyto-oestrogens in the milk is shown in Table 5 with equol, naringenin and enterolactone being the quantitatively most important phyto-oestrogens. The total concentration of phyto-oestrogens was 4 to 5.6 times higher in milk from the red clover diet compared with the other treatments and more or less comparable for the other three treatments. The concentration of equol, daidzein and formononetin was 6.1 to 11.8, 2.7 to 6.6 and 2.7 to 4 times higher ($P < 0.001$), respectively, in milk from cows fed the red clover mixture compared with the other treatments, whereas there were no significant differences between white clover, chicory and lucerne. The concentration of biochanin A was significantly higher for red clover compared to the other treatments in May, while no differences were observed in June. For naringenin and glycitein the concentrations were similar across treatments in May but significantly lower for white clover compared to the other treatments in June. The concentration of chrysin was significantly lower for white clover in June compared with the other treatments. Enterolactone was 1.3 to 2 times lower for red clover compared with the other treatments in both periods. Treatment had no effect on the concentration of enterodiol but overall the concentration was lower in June compared with May.

When looking at the effect of period, the total concentration of phyto-oestrogens was significantly decreased from May to June, mainly because of a significant decrease of phyto-oestrogens in milk from cows fed the red clover mixture.

Discussion*Phyto-oestrogens in herbage*

To our knowledge only two previous studies have related the concentration of phyto-oestrogens in bovine milk to the concentration of phyto-oestrogen in the diet. Steinshamn *et al.* (2008) used two diets based on either white clover or red clover silages while Andersen *et al.* (2009) among other had one diet based on lucerne silage. When comparing the concentration of phyto-oestrogens in red and white clover mixtures in this study to the red and white clover silages used by Steinshamn *et al.* (2008), the concentration of formononetin in this study was considerably higher (3.6 to 4 and 2.5 to 3 times, respectively). For biochanin A, the concentration was more or less the same for white clover but considerably higher for red clover. The concentration of formononetin was considerably higher and the concentration of daidzein considerably lower in the lucerne mixture in the present study compared to the lucerne silage in Andersen *et al.* (2009). Thus, there was an overall higher concentration of isoflavones in the pastures used in this study than the silages fed in the other two studies. This is likely to reflect a higher concentration of phyto-oestrogens in fresh grass than in silage. Supporting this, Sivesind and Seguin (2005) found that the concentration of phyto-oestrogen was more or less similar among cultivars of red

Table 5 Concentration ($\mu\text{g/l}$) of phyto-oestrogens in milk in relation to dietary treatments

Phyto-oestrogen	May						June						P-values						
	White clover milk		Red clover milk		Lucerne milk		Chicory milk		White clover milk		Red clover milk		Lucerne milk		Chicory milk		Diet	Period	Diet \times Period
Chrysin	0.27 ^{ab}	0.30 ^b	0.26 ^{ab}	0.25 ^a	0.12 ^c	0.18 ^d	0.19 ^d	0.19 ^d	0.0290	0.0001	0.0001	0.0290	0.0001	0.0001	0.0001	0.0001	0.0099		
Naringenin	13.43 ^{ab}	14.30 ^{ac}	14.00 ^{ac}	13.97 ^{ac}	12.29 ^b	13.90 ^{ac}	14.43 ^{ac}	14.79 ^c	0.0063	0.8361	0.1730	14.79 ^c	14.79 ^c	0.0063	0.8361	0.1730	0.0099		
Biochanin A	0.34 ^{ad}	0.55 ^b	0.25 ^{ade}	0.23 ^{adef}	0.05 ^c	0.20 ^{cd}	0.09 ^{ce}	0.08 ^{cf}	0.0010	0.0001	0.2758	0.08 ^{cf}	0.08 ^{cf}	0.0010	0.0001	0.2758	0.0001		
Daidzein	0.32 ^a	1.14 ^b	0.42 ^a	0.26 ^a	0.37 ^a	1.92 ^c	0.44 ^a	0.29 ^a	0.0001	0.0020	0.0001	0.44 ^a	0.29 ^a	0.0001	0.0020	0.0001	0.0001		
Equol	30.04 ^a	355.42 ^b	46.20 ^a	58.21 ^a	21.76 ^a	214.97 ^c	19.85 ^a	18.82 ^a	0.0001	0.0001	0.0003	19.85 ^a	18.82 ^a	0.0001	0.0001	0.0003	0.0001		
Formononetin	1.98 ^{ae}	6.28 ^b	2.28 ^a	1.73 ^{ace}	0.99 ^{ce}	4.03 ^d	1.16 ^c	1.13 ^{ce}	0.0001	0.0001	0.1063	1.16 ^c	1.13 ^{ce}	0.0001	0.0001	0.1063	0.0001		
Glycitein	3.55 ^a	3.66 ^a	3.68 ^a	3.94 ^{ac}	3.62 ^a	4.45 ^b	4.36 ^{bc}	4.28 ^b	0.0020	0.0001	0.069	4.36 ^{bc}	4.28 ^b	0.0020	0.0001	0.069	0.0001		
Enterodiol	7.76 ^a	7.53 ^{ab}	7.63 ^{ab}	7.13 ^{abc}	6.47 ^{bc}	5.92 ^c	6.48 ^{bc}	6.57 ^{bc}	0.8084	0.0004	0.7060	6.48 ^{bc}	6.57 ^{bc}	0.8084	0.0004	0.7060	0.0001		
Enterolactone	12.47 ^a	7.96 ^b	16.25 ^c	11.15 ^a	10.50 ^{ab}	8.39 ^b	12.47 ^a	12.68 ^a	0.0001	0.1605	0.0266	12.47 ^a	12.68 ^a	0.0001	0.1605	0.0266	0.0001		
Total concentration	70.16 ^a	397.14 ^b	91.00 ^a	96.87 ^a	56.19 ^a	253.97 ^c	59.47 ^a	58.83 ^a	0.0001	0.0001	0.0006	59.47 ^a	58.83 ^a	0.0001	0.0001	0.0006	0.0001		

P-values for the effect of diet, period and their interaction are shown. The superscript symbols a, b, c, d, e, f designate significant difference ($P < 0.05$) between dietary treatments.

clover, but that the concentration of total isoflavones was 22% higher in fresh herbage compared to silage and hay. However, it is important to note that ensiling is a dynamic process dependent on many factors (pH, microbial population, temperature, initial herbage composition) that fluctuate, and therefore these factors potentially influence the concentration of isoflavones in silage.

Phyto-oestrogens in milk

Comparing the phyto-oestrogen level in this study to commercial milk samples analysed by Antignac *et al.* (2004), the concentration of enterodiol was higher for all dietary treatments, but for white clover, lucerne and chicory the concentration of biochanin, daidzein, equol and enterolactone was lower. Thus, white clover, lucerne and chicory pastures do not seem effective in increasing the concentration of phyto-oestrogens in milk. However, for red clover there was an increased concentration of formononetin and equol compared to the commercial milk samples analysed by Antignac *et al.* (2004). A high concentration of equol has also been observed in other studies with milk from cows fed red clover pastures or in organic milk production where the high concentration of equol seems to be linked to the frequent use of leguminous plants, probably specifically the use of red clover (Antignac *et al.*, 2004; Purup *et al.*, 2005; Hoikkala *et al.*, 2007).

Equol appeared to be the most abundant phyto-oestrogen in milk regardless of the dietary treatment and period. This is in accordance with previous findings (Antignac *et al.*, 2004; Hoikkala *et al.*, 2007; Steinshamn *et al.*, 2008). The very high concentration of equol in milk from especially cows grazing red clover was expected, since red clover has a high concentration of formononetin (0.8 to 11 mg/g DM) but also biochanin A (0.8 to 5 mg/g DM) depending on the part (flower, stem or leaves) and maturity of the plant, cultivar and environment (Sivesind and Seguin, 2005; Booth *et al.*, 2006). Formononetin is metabolized by microbes in the rumen via daidzein to equol which is the major isoflavone absorbed to the blood circulation after feeding red clover/grass silage, whereas biochanin A is demethylated to genistein to form mainly p-ethyl phenol (Lundh, 1995). Unfortunately, the concentration of daidzein in herbage, and genistein in herbage and milk, could not be analysed in this experiment as previously described.

The concentration of enterodiol was high while the concentration of equol, enterolactone and formononetin was low in the white clover milk compared to results obtained by Steinshamn *et al.* (2008). The red clover diet resulted in a lower concentration of enterolactone and biochanin A compared to Steinshamn *et al.* (2008). For the lucerne diet, the concentration of enterodiol and equol was much higher in the present study than observed in milk from cows fed lucerne silage (Andersen *et al.*, 2009). In contrast, this study found a lower concentration of enterolactone and a slightly lower concentration of biochanin A and daidzein. The variation between experiments in the concentration of phyto-oestrogen in the milk is likely to reflect initial differences in

the concentration of phyto-oestrogens in the diet but could also be due to differences in the metabolism between pasture and silage.

The concentration of naringenin and chrysin in feedstuffs and milk has not been reported before. Similarly, there are no previous studies on phyto-oestrogens in chicory. The concentration of glycitein in red clover has been reported by Tsao *et al.* (2006) who found similar levels as those found in this study. The concentration of glycitein in the other herbage and milk has not been reported previously.

Compared to sources rich in phyto-oestrogens like soy milk (5 to 10 mg isoflavones/kg) and tofu (13.5 to 67 mg isoflavones/kg) (Committee on Toxicity of Chemicals in Food, 2003), the concentration of phyto-oestrogens in bovine milk is low. Nevertheless, the mixture of many phyto-oestrogens could have an additive or synergistic effect and thus a biological effect. For instance, a recent experiment showed that soy extract is more potent than genistein, the most potent phyto-oestrogen in soy, in inhibiting tumour growth. This is presumably due to the synergistic effect of the various bioactive components in the soy extract (Kim *et al.*, 2008). Future studies will have to investigate if there is a biological effect of phyto-oestrogens in bovine milk. This study verifies that it is possible to affect the concentration of phyto-oestrogens in the milk through the feeding and that red clover diets seem to be the most effective way of increasing the concentration of phyto-oestrogens in bovine milk.

Acknowledgements

This work was supported by the Ministry of Food, Agriculture and Fisheries (project No. 3304-FOJO-05-38-01).

References

- Andersen C, Hansen-Møller J, Weisbjerg MR and Sejrsen K 2009. The content of phyto-oestrogens in milk and feedstuffs from cows fed clover grass silage or different concentrations of alfalfa silage. *Animal* 3, 617–622.
- Antignac JP, Cariou R, Le Bizec B and André F 2004. New data regarding phytoestrogens content in bovine milk. *Food Chemistry* 87, 275–281.
- Benassayag C, Perrot-Applanat M and Ferre F 2002. Phytoestrogens as modulators of steroid action in target cells. *Journal of Chromatography B, Analytical Technologies in the Biomedical and Life Sciences* 777, 233–248.
- Booth NL, Cassia RO, Ping Y, Totura S, Deng Y, Hedayat AS, Bolton JL, Pauli GF and Farnsworth NR 2006. Seasonal variation of red clover (*Trifolium pratense* L., Fabaceae) isoflavones and estrogenic activity. *Journal of Agricultural and Food Chemistry* 54, 1277–1282.
- Committee on Toxicity of Chemicals in Food 2003. COT report – phytoestrogens and health. Food Standards Agency, London.
- Eriksen J, Søgaard K, Kristensen T, Fretté XC, Wiking L and Nielsen JH 2007. Herbage and milk productivity and quality when grazing different legumes and herbs. *Grassland Science in Europe* 12, 212–214.
- Hoikkala A, Mustonen E, Saastmoinen I, Jokela T, Teponen J, Saloniemi H and Wähälä K 2007. High levels of equol in organic skimmed Finnish cow milk. *Molecular Nutrition & Food Research* 51, 782–786.
- Kim HA, Jeong KS and Kim YK 2008. Soy extract is more potent than genistein on tumor growth inhibition. *Anticancer Research* 28, 2837–2841.
- Kurzer MS and Xu X 1997. Dietary phytoestrogens. *Annual Review of Nutrition* 17, 353–381.

Lundh T 1995. Metabolism of estrogenic Isoflavones in domestic animals. *Proceedings of the Society of Experimental Biology and Medicine* 208, 33–39.

Purup S, Hansen-Møller J, Sejrsen K, Christensen LP, Lykkesfeldt AE, Leffers H and Skakkebæk NE 2005. Increased phytoestrogen content in organic milk and the biological importance. Newsletter from Danish Research Centre for Organic Farming, No. 2. Danish Research Centre for Organic Farming, Tjele, Denmark.

Saloniemi H, Wähälä K, Nykänen-Kurki P, Kallela K and Saastamoinen I 1995. Phytoestrogen content and estrogenic effect of legume fodder. *Proceedings of the Society for Experimental Biology and Medicine* 208, 13–17.

SAS Institute 1999. SAS/STAT Software. Release 8.02. SAS Institute Inc., Cary, NC, USA.

Simonsen F 2006. *Analyseteknik, instrumentering og metoder*, 2nd edition. Nyt Teknisk Forlag, Copenhagen V, Denmark.

Sivesind E and Seguin P 2005. Effects of the environment, cultivar, maturity, and preservation method on red clover isoflavone concentration. *Journal of Agriculture and Food Chemistry* 53, 6397–6402.

Steinshamn H, Purup S, Thuen E and Hansen-Møller J 2008. Effects of clover silages and concentrate supplementation on the content of phytoestrogens in dairy cow milk. *Journal of Dairy Science* 91, 2715–2725.

Thompson LU, Robb P, Serraino M and Cheung F 1991. Mammalian lignan production from various foods. *Nutrition and Cancer* 16, 43–52.

Tsao R, Papadopoulus Y, Yang R, Young JC and McRae K 2006. Isoflavone profiles of red clovers and their distribution in different parts harvested at different growing stages. *Journal of Agricultural and Food Chemistry* 54, 5797–5805.