Generalized and Naevoid Epidermolytic Ichthyosis in Denmark
Clinical and Mutational Findings
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A Danish–Swedish collaboration was established to identify and classify a Danish cohort of patients with epidermolytic ichthyosis, also known as epidermolytic hyperkeratosis. Patients were recruited from 5 dermatology departments in Denmark, and data were obtained using a structured questionnaire and a systematic examination together with photographs, histopathological descriptions and blood samples for mutational analysis. Sixteen patients from 12 families with generalized or naevoid epidermolytic ichthyosis and ichthyosis bullosa of Siemens were identified. Five families had mutations in K1 and 6 families had mutations in K10. Nine patients had been treated with systemic retinoids (etretinate, acitretin, isotretinoin or alitretinoin), but only 3 patients had acceptable treatment responses and chose to continue therapy. In conclusion epidermolytic ichthyosis is a rare disease with a prevalence of approximately 1 in 350,000 in Denmark and a high percentage of de novo mutations (75%). We identified 4 novel disease-causing mutations. Key words: genomic DNA sequencing; epidermolytic ichthyosis; epidermolytic hyperkeratosis; phenotypic variation.

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Epidermolytic ichthyosis (EI; OMIM 113800), also referred to as epidermolytic hyperkeratosis or bullous congenital ichthyosiform erythroderma, is a rare keratinization disorder caused by dominant negative mutations in keratin genes KRT1 and KRT10, leading to an unstable cytoskeleton of epithelial keratinocytes and epidermal hyperkeratosis (1). In contrast to most other ichthyoses, the histopathological picture of EI is distinctive, with hyperkeratosis, acanthosis and characteristic clumping of tonofilaments, intracellular vacuolization and intraepidermal blisters.

Children with generalized EI are usually born with erythroderma, extensive blistering and denuded skin that later changes into a more ichthyotic phenotype with rippled hyperkeratosis mainly on flexural surfaces. The bullous component gradually becomes less prominent, but patients may continuously have problems with blistering, especially after trauma or skin infections. In addition, some patients also have palmoplantar keratoderma (PPK).

The naevoid form of EI, linear epidermolytic ichthyosis (LEI), occurs as epidermal naevi with cytoskeletal abnormalities of epidermolytic hyperkeratosis representing somatic mosaicism. In case of gonadal mosaicism the offspring of the patient may develop full-blown EI. Therefore epidermolytic naevi are included in the most recent classification of keratinopathic ichthyosis (1).

Ichthyosis bullosa of Siemens (IBS) resembles EI, but has a milder phenotype with less hyperkeratosis, more superficial blistering and no PPK. Mutations in K2, a keratin expressed later during differentiation, are responsible for IBS (1, 2).

In this study we describe the clinical subtypes and mutational findings in a cohort of 16 Danish patients with EI/IBS.

PATIENTS AND METHODS

Patients and samples

Patients were identified through a national study of congenital ichthyosis undertaken in 2004 to 2006 by 3 of the authors (AB, AG and FB). In order to trace all patients seen in Denmark from 1984 to 2003 with a diagnosis of congenital ichthyosis, contact was established with all 6 departments of dermatology. Patients with a clinical phenotype consistent with EI or IBS were included in this study.

Written informed consent was obtained from all patients according to approval by the Danish ethics committee (jr. no. VF20040178).

A complete medical history was obtained from all patients using a structured questionnaire. In order to obtain a uniform classification, patients were systematically examined by at least 2 of 3 authors (AB, AG and FB) together with a local dermatologist to give the best clinical classification. Photographs were taken, previous histopathological descriptions were reviewed and photographs were taken and previous histopathological descriptions were reviewed.

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and new skin biopsies for light or electron microscopy were taken in case of diagnostic doubt. Blood samples were obtained for DNA extraction and mutational analysis in KRT1 and KRT10. In patients where no pathogenic mutation was identified, KRT2 was also examined.

### Genetic analyses

Genomic DNA (gDNA) was extracted from 2 ml ethylenediaminetetraacetic acid (EDTA) blood using E:Z:N:A:Midikit’s (Omega Bio-Tek Norcross, GA, USA). 100 ng of gDNA was used for PCR amplification of all KRT1 exons and the hot-spot regions in KRT2 and KRT10. The DNA was added to a mix of 1×Taq buffer, 0.2 mM of each dNTP, forward and reverse primers (10 µM), 2.5 mM MgCl2, and 0.05 U Taq polymerase (Applied Biosystems, Stockholm, Sweden) in a total volume of 50 µl.

All patients were initially screened for mutations in hotspot regions of KRT1 and KRT10 by denaturing high-performance liquid chromatography (HPLC) (Transgenomic, Omaha, USA). Before the analysis the PCR fragments were partially denatured by decreasing the temperature from 94ºC to 40ºC during 30 min. An aliquot of PCR fragments was purified by using GTX™ PCR DNA and GelBand purification Kit (GE Healthcare, Uppsala, Sweden) followed by DNA sequencing. Automated sequencing was done using Big Dye Terminator kit and analysed on an ABI Prism 377 DNA sequencer (Perkin-Elmer Applied Biosystems, Foster City, CA, USA).

### RESULTS

A total of 16 patients from 12 families fulfilled the inclusion criteria for EI/IBS. The symptoms varied in intensity from mild to severe and in 8 of the patients an associated PPK was observed (Table I). For 2 families, 3 affected members were found in each family. In one of the families (number 5) an affected mother and 2 children with EI were found (Fig. 1), and in the other family (number 6) EI was observed in 3 generations, starting with a naevoid lesion in the grandfather (Fig. S1; available from: http://www.medicaljournals.se/acta/content/?doi=10.2340/00015555-1447), had no identifiable KRT1, KRT2 or KRT10 mutation, except for a previously reported polymorphism, an 18-bp deletion in exon 1 of KRT2 (13).

### DISCUSSION

We identified 16 Danish patients with generalized or naevoid EI and IBS, corresponding to a prevalence of approximately 1 in 350,000 in Denmark. This is 3 times higher than the estimated prevalence in other Scandinavian countries (14), but similar to prevalence estimates of 1 in 100,000–300,000 reported in the literature (15, 16).

The patients with generalized EI typically had disease onset from birth, with erythroderma and blisters with denuded skin areas (Fig. S2; available from: http://www.medicaljournals.se/acta/content/?doi=10.2340/00015555-1447). Some patients subsequently developed mild flexural involvement, while others had generalized erythroderma with blistering/erosion tendency continuing in adulthood, but overall symptoms improved with age. A few patients had very severe PPK with contractures (Fig. S4; available from http://www.medicaljournals.se/acta/content/?doi=10.2340/00015555-1447).

In this study, the percentage of de novo mutations was very high; 75% compared with a previously reported spontaneous mutation rate of about 50% (15–18). The majority of reported mutations were heterozygous missense mutations. When mutations are located at the conserved helix boundary motifs, the helix initiation or termination peptides and the non-helical H1 domain of K1 and K10, they will result in severe EI (17, 19). In family number 5 with a moderate EI phenotype, a partial deletion of L1 and L2 region in K1 was identified. Insertion/deletion mutations and splice site-defects or dinucleotide alterations in KRT10 have also been described and more than 100 mutations in KRT1 and KRT10 have already been reported (17, www.interfil.org).
All 6 patients with K1 mutations had PPK, while only 1 out of 6 patients with K10 mutation had PPK. This is in accordance with earlier published studies (15, 17, 20). The absence of PPK in patients with mutation in K10 is explainable by palmoplantar expression of K9, which is a functional substitute for K10. In our study 2 unrelated females (patient XIII and XIV) had the same mutation in K10, but only one of them had a mild PPK, suggesting that other genetic or environmental factors also influence the phenotype.

LEI is manifested as streaks of hyperkeratosis following the lines of Blaschko. LEI is caused by somatic mutations in K10, or rarely K1, arising post-zygotically during embryogenesis (21–23). Mutations have been demonstrated in keratinocytes from affected skin, whereas the mutations are absent in non-lesional skin and blood

**Table I. Summary of clinical and mutational data in 16 Danish patients with various types of epidermolytic ichthyosis (EI), showing those with KRT1 mutations at the top (new mutations are shown in bold)**

<table>
<thead>
<tr>
<th>Pat. No.</th>
<th>Diagnosis</th>
<th>Sex/age (years)</th>
<th>Familial (family No.)</th>
<th>Mutation</th>
<th>Protein</th>
<th>Clinical presentation</th>
<th>At birth</th>
<th>At inclusion in study</th>
<th>Histology showing EHK</th>
</tr>
</thead>
<tbody>
<tr>
<td>I</td>
<td>EI</td>
<td>M/34</td>
<td>No (1)</td>
<td><strong>KRT1 c.1457T&gt;A</strong></td>
<td>p.Leu486Gln</td>
<td>Blistering</td>
<td>Erosions</td>
<td>Few erosions</td>
<td>Hyperkeratosis</td>
</tr>
<tr>
<td>II</td>
<td>EI</td>
<td>M/20</td>
<td>No (2)</td>
<td><strong>KRT1 c.1424T&gt;C</strong></td>
<td>p.Leu475Pro</td>
<td>Blistering</td>
<td>Erosions</td>
<td>Focal blisters</td>
<td>Hyperkeratosis</td>
</tr>
<tr>
<td>III</td>
<td>EI</td>
<td>M/27</td>
<td>No (3)</td>
<td>KRT1 c.145A&gt;G</td>
<td>p.Tyr482Cys</td>
<td>Erythroderma</td>
<td>Blistering</td>
<td>Erythroderma</td>
<td>Hyperkeratosis</td>
</tr>
<tr>
<td>IV</td>
<td>EI</td>
<td>M/40</td>
<td>No (4)</td>
<td><strong>KRT1 c.1465G&gt;A</strong></td>
<td>p.Glu489Lys</td>
<td>Blistering</td>
<td>Erosions</td>
<td>Focal blisters</td>
<td>Erythroderma</td>
</tr>
<tr>
<td>V</td>
<td>EI</td>
<td>F/33</td>
<td>Yes (5)</td>
<td><strong>KRT1 c.673_702 del 30</strong></td>
<td>p.His225_Phe 234del</td>
<td>Erythroderma</td>
<td>Blistering</td>
<td>Focal blisters</td>
<td>Hyperkeratosis</td>
</tr>
<tr>
<td>VI</td>
<td>EI</td>
<td>M/3</td>
<td>Yes (5)</td>
<td><strong>KRT1 c.673_702 del 30</strong></td>
<td>p.His225_Phe 234del</td>
<td>Erythroderma</td>
<td>Blistering</td>
<td>Focal blisters</td>
<td>Hyperkeratosis</td>
</tr>
<tr>
<td>VII</td>
<td>EI</td>
<td>F/1</td>
<td>Yes (5)</td>
<td>NT</td>
<td>Erythroderma</td>
<td>Blistering</td>
<td>Erythroderma</td>
<td>Hyperkeratosis</td>
<td>PPK</td>
</tr>
<tr>
<td>VIII</td>
<td>LEI</td>
<td>M/65</td>
<td>Yes (6)</td>
<td>No mutation found</td>
<td>Naevoid/striate thickening of skin on right-sided extremities</td>
<td>Striate Hyperkeratosis</td>
<td></td>
<td>Yes</td>
<td></td>
</tr>
<tr>
<td>IX</td>
<td>EI</td>
<td>F/38</td>
<td>Yes (6)</td>
<td><strong>KRT10 c.1333G&gt;A</strong></td>
<td>p.Glu445Lys</td>
<td>Erythroderma</td>
<td>Blistering</td>
<td>Erythroderma</td>
<td>Hyperkeratosis</td>
</tr>
<tr>
<td>X</td>
<td>EI</td>
<td>F/6</td>
<td>Yes (6)</td>
<td>NT</td>
<td>Erythroderma</td>
<td>Blistering</td>
<td>Hyperkeratosis</td>
<td>EK</td>
<td>NT</td>
</tr>
<tr>
<td>XI</td>
<td>EI</td>
<td>M/18</td>
<td>Yes (7)</td>
<td>KRT10 c.466C&gt;T</td>
<td>p.Arg156Cys</td>
<td>Erythroderma</td>
<td>Hyperkeratosis</td>
<td>Focal blisters</td>
<td>NT</td>
</tr>
<tr>
<td>XII</td>
<td>EI</td>
<td>M/3</td>
<td>Yes (8)</td>
<td>KRT10 c.466C&gt;A</td>
<td>p.Arg156Ser</td>
<td>Erythroderma</td>
<td>Hyperkeratosis</td>
<td>Focal blisters</td>
<td>NT</td>
</tr>
<tr>
<td>XIII</td>
<td>EI</td>
<td>F/44</td>
<td>No (9)</td>
<td>KRT10 c.482T&gt;C</td>
<td>p.Leu161Ser</td>
<td>Erythroderma</td>
<td>Hyperkeratosis</td>
<td>Collodion baby</td>
<td>NT</td>
</tr>
<tr>
<td>XIV</td>
<td>EI</td>
<td>F/38</td>
<td>No (10)</td>
<td>KRT10 c.482T&gt;C</td>
<td>p.Leu161Ser</td>
<td>Hyperkeratosis</td>
<td>Erythroderma</td>
<td>Hyperkeratosis</td>
<td>PPK</td>
</tr>
<tr>
<td>XV</td>
<td>EI</td>
<td>M/42</td>
<td>No (11)</td>
<td><strong>KRT10 c.452A&gt;C</strong></td>
<td>p.Gln151Pro</td>
<td>Blistering</td>
<td>Erythroderma</td>
<td>Hyperkeratosis</td>
<td>NT</td>
</tr>
<tr>
<td>XVI</td>
<td>IBS</td>
<td>F/29</td>
<td>Adopted (12)</td>
<td>No mutation found</td>
<td>Unknown</td>
<td>Hyperkeratosis</td>
<td></td>
<td></td>
<td>NT</td>
</tr>
</tbody>
</table>

*a* No such mutation found in 50 controls. 
*b* Analysed at the Department of Clinical Genetics, Aarhus University Hospital, Denmark. 
*c* Analysed by Dr Paul Bowden, UWCM, Cardiff, UK.

EHK: epidermolytic hyperkeratosis; LM: light microscopy; EM: electron microscopy; LEI: linear epidermolytic ichthyosis; IBS: ichthyosis bullosa of Siemens; PPK: palmoplantar keratoderma; ND: not diagnostic; NT: not tested.
leukocytes (17). There seems to be a correlation between widespread LEI and the risk of germ-line transmission (22). The patient with LEI (VIII) had rippled and hyperkeratotic streaks on his right-side extremities, as well as thickened macerated skin in his right axilla (Fig. S1). Blood leukocytes, but no keratinocytes from affected skin, were available for molecular genetic analysis; no mutations in K1 or K10 could be detected.

We could not identify any mutation in patient XVI. The patient was adopted from Korea, and had clinical features of IBS (Fig. S2). Mutations outside the hotspot region of KRT2 are possible.

The treatment options for EI are less than satisfactory. Retinoids are used in more severe cases of EI, but are usually moderately effective and carry a risk of side-effects when given systemically (24). Nine of our patients had been treated with systemic retinoids (etretinate, acitretin, isotretinoin or alitretinoin). Of these, 3 patients (patient I with K1 mutation and patients XIV and XV with K10 mutations) had acceptable treatment responses and are continuously on retinoid therapy. However, in another 2 patients (II and III) with K1 mutations skin symptoms such as blistering and erosions worsened during treatment; presumably they are more vulnerable to a retinoid down-regulation of K2, which may otherwise, to some extent, compensate for a mutated or missing K1 protein (14).

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The authors declare no conflicts of interest.

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