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# Virus-induced brain pathology and the neuroinflammation-inflammation continuum: the neurochemists view

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## Abstract

Fascinatingly, an abundance of recent studies has subscribed to the importance of cytotoxic immune mechanisms that appear to increase the risk/trigger for many progressive neurodegenerative disorders, including Parkinson's disease (PD), Alzheimer's disease (AD), amyotrophic lateral sclerosis, and multiple sclerosis. Events associated with the neuroinflammatory cascades, such as ageing, immunologic dysfunction, and eventually disruption of the blood–brain barrier and the “cytokine storm”, appear to be orchestrated mainly through the activation of microglial cells and communication with the neurons. The inflammatory processes prompt cellular protein dyshomeostasis. Parkinson's and Alzheimer's disease share a common feature marked by characteristic pathological hallmarks of abnormal neuronal protein accumulation. These Lewy bodies contain misfolded  $\alpha$ -synuclein aggregates in PD or in the case of AD, they are A $\beta$  deposits and tau-containing neurofibrillary tangles. Subsequently, these abnormal protein aggregates further elicit neurotoxic processes and events which contribute to the onset of neurodegeneration and to its progression including aggravation of neuroinflammation. However, there is a caveat for exclusively linking neuroinflammation with neurodegeneration, since it's highly unlikely that immune dysregulation is the only factor that contributes to the manifestation of many of these neurodegenerative disorders. It is unquestionably a complex interaction with other factors such as genetics, age, and environment. This endorses the “multiple hit hypothesis”. Consequently, if the host has a genetic susceptibility coupled to an age-related weakened immune system, this makes them more susceptible to the virus/bacteria-related infection. This may trigger the onset of chronic cytotoxic neuroinflammatory processes leading to protein dyshomeostasis and accumulation, and finally, these events lead to neuronal destruction. Here, we differentiate “neuroinflammation” and “inflammation” with regard to the involvement of the blood–brain barrier, which seems to be intact in the case of neuroinflammation but defect in the case of inflammation. There is a neuroinflammation-inflammation continuum with regard to virus-induced brain affection. Therefore, we propose a staging of this process, which might be further developed by adding blood- and CSF parameters, their stage-dependent composition and stage-dependent severeness grade. If so, this might be suitable to optimise therapeutic strategies to fight brain neuroinflammation in its beginning and avoid inflammation at all.

**Keywords** Neurodegeneration · Parkinson's disease · Alzheimer's disease · Viruses · Microglia and neuroinflammation · Inflammation · Cytotoxicity · Immunology

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## Introduction

Parkinson's (PD) and Alzheimer's (AD) are progressive neurodegenerative diseases with unknown aetiology. Nevertheless, a plethora of evidence generated from studies suggests the close involvement of risk factors such as ageing, genetics, and pathogen-induced neuroinflammation. Viral infections in the central nervous system (CNS) are significantly more frequent compared to other pathogens, including bacteria, fungi, and protozoa (Romero and Newland 2003).

Strikingly, a large study of 450,000 people (above the age of 60 years and of European ancestry) from Finland and the UK found that viral infections (mainly neurotropic viruses) increased the risk of development of neurodegenerative disorders, including PD and AD (Levine et al. 2023). The viruses reported by this study included influenza virus, herpes zoster virus, enteroviruses, Epstein–Barr virus (EBV), and Herpes simplex virus.

Incidentally, the acute consequences related to virus infections are associated with a heightened host's inflammatory response rather than the viral load itself (Coates et al. 2017). Thus, neuroinflammation resulting from virus (chiefly neurotropic by nature) exposure appears to be strongly associated with neurological sequelae despite viral clearance and may play an instrumental role in the early stages of disease development. As such, a plethora of evidence indicates neurodegeneration may be a casualty of virus-invoked inflammation and disruption of cellular protein metabolism (Leblanc and Vorberg 2022; Smeyne et al. 2021).

## Putative pathways and mechanisms for viral-associated neurodegeneration

### Neuroinflammation

Viruses access the CNS via two major routes: the blood supply (mumps and measles viruses (Griffin 2014)), and some migrate through the peripheral nerves. Rabies virus and poliovirus employ peripheral motor neurons to enter the CNS (Jackson 2014). It has been reported that viral-infected mononuclear cells can gain access to the CNS within 12 h after infection with coxsackie virus B3 (Tabor-Godwin et al. 2010).

The blood–brain barrier (BBB) is a close-fitting vascular barrier composed of brain microvascular endothelial cells, astrocytes, and pericytes (Ben-Zvi et al. 2014). It tightly regulates the entry of components from the blood respectively peripheral nervous system into the CNS (Rausch et al. 1983). The BBB restricts the entry of toxins, macromolecules, and trespassing pathogens. However, some regions of the CNS, like the circumventricular organs, may allow the entry of pathogens since they do not have complete protection from the BBB (Daneman and Prat 2015). Alternatively, BBB itself may be “leaky” or dysfunctional, allowing the free passage of toxins, pathogens, immune cells, and their products. This would also ireregulate ions and signal homeostasis (Profaci et al. 2020), which may have debilitating effects on the physiological functions of neurons and represent a prelude to neurodegeneration to ensue.

Virus or viral-related consequences may be instrumental in impairing the integrity of the BBB (Fig. 1). Inflammation generated by systemic viral infection may be a potential

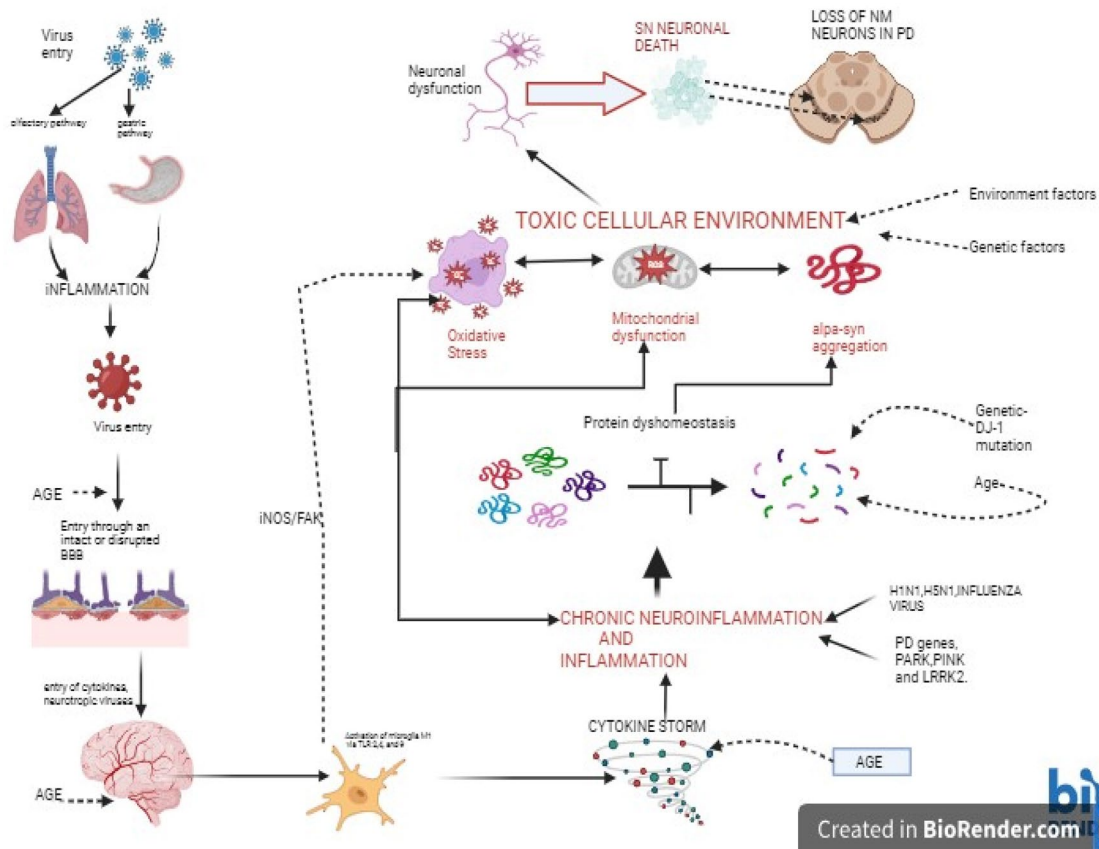
candidate in the damage of the BBB, thereby permitting viral neuroinvasion (Eugenin 2006; Chen et al. 2018).

Increasing evidence supports the involvement of neuro-inflammatory mechanisms as key players in commanding viral-related neurotoxicity (Pathak and Sriram 2023). Following virus-associated infections, neuroinflammation may manifest in various areas in the CNS, such as encephalitis in the brain, meningitis (meninges), and/or myelitis (spinal cord) (Tansey and Goldberg 2010; Klein et al. 2017). Alternatively, the inflammation can occur concurrently in different areas (meningoencephalitis). Interestingly, although viral pathology exerts a significant role in inducing malfunction of the CNS physiological processes and mechanisms, the immune response against the virus can also contribute to dysfunction and disease. Thus, neuroinflammatory cells and mediators may serve as a “Trojan Horse” (Figs. 1, 3) once they gain their illegal entry into the brain, biding their time before invoking chaos and cellular destruction.

Depending on the virus, they can replicate within neurons, glia, and microglia. Of the three glial cells, only microglia and astrocytes execute a cytotoxic effect. The oligodendrocytes like the neuronal cells are the casualties of the destructive inflammatory processes (McGeer and McGeer 2007).

Microglia can serve as a double-edged sword, and in that in response to neuronal damage or an invading pathogen, it can execute both a protective and destructive action (Li and Barres 2018). This dual role probably relates to the two distinct microglial phenotypes, M1 and M2 (Prinz and Jung 2019). M1 exerts pro-inflammatory and neuro-destructive activities (Figs. 2, 5), whereas M2 microglia express markers such as (cluster of differentiation, CD) CD163, and CD206 that reduce the rate of inflammation and tissue repair (Orihuela et al. 2015).

Microglial cells are key regulators of inflammation related to neurodegenerative disorders. Neuronal cell destruction can activate glia/microglia and trigger neuroinflammation via the release of inflammatory mediators and reactive oxygen species (Wolf et al. 2017). Free radicals can initiate cytotoxic processes such as oxidative stress thereby further adding to the neuronal cell death toll. Also, activated microglial can produce pro-inflammatory cytokines and chemokines to suppress and eradicate the viral infection (Amarante-Mendes et al. 2018). Usually, the second wave of cytokine releases (post-infection) promotes viral clearance (Guo and Thomas 2017). However, if this release goes unchecked coupled with dysregulation of anti-inflammatory reactions, a “cytokine storm” follows an aggressive hyperinflammatory response. This can have direct and catastrophic consequences by precipitating serious immune pathology including acute respiratory distress syndrome, coagulopathy, extensive tissue destruction, and multi-organ failure resulting in death (Verbist and Nichols



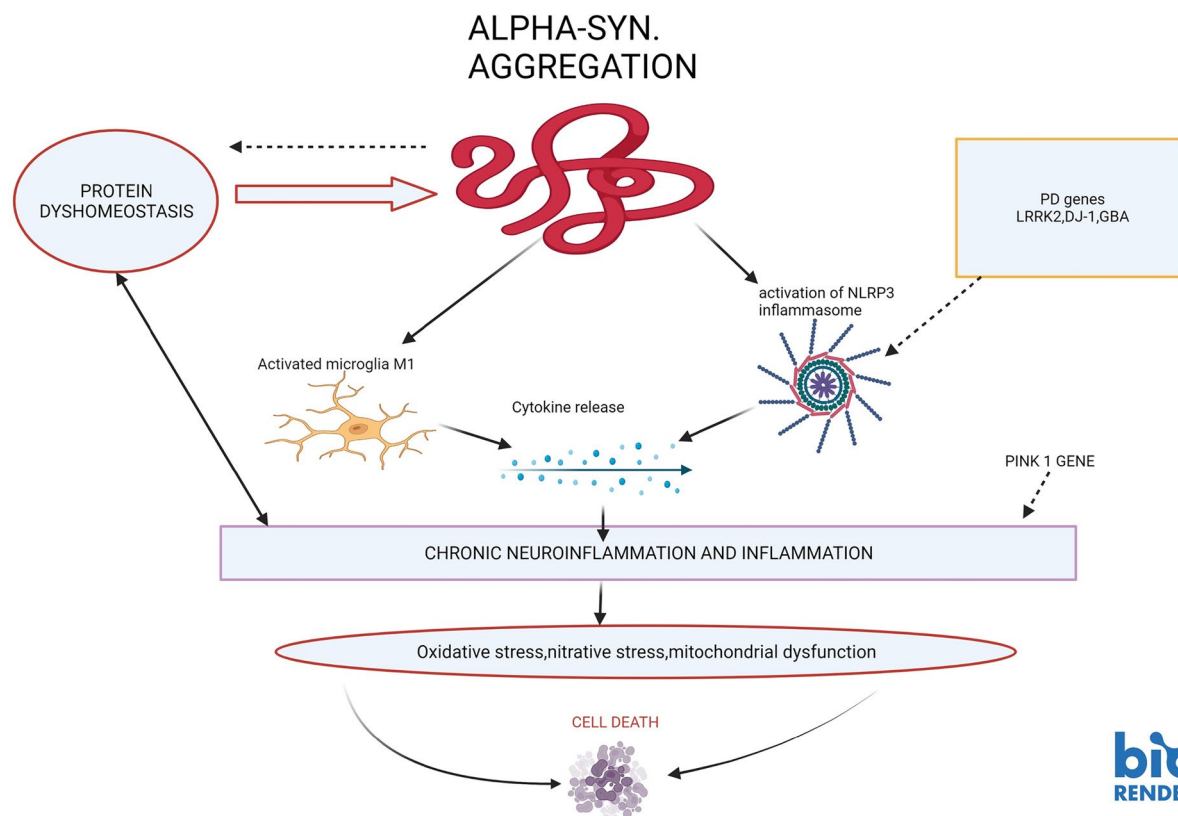
**Fig. 1** Sequence of events that may be related to viral-related infection coupled with other risk factors in Parkinson's disease. Viruses (H1N1, H5N1, FLAVI, HEP.C, HIV, etc) gain entry through the olfactory or/and gastric avenues, inciting neuroinflammation in the respiratory and gastrointestinal tract respectively. This leads to the release of proinflammatory cytokines and chemokines, which can disrupt the integrity of the blood-brain barrier (BBB) by producing fenestrations on its endothelial cells. Thus, making it "leaky" and allowing the passage of pathogens/toxins to the brain. On gaining entry to the brain, these cytokines can activate microglia (M1 cells) via toll-like receptors (2,4, and 9), which generates further production of pro-inflammatory cytokines (IL-1, IL-6, and TNF- $\alpha$ , etc.) and onset neuroinflammation in the brain. Also, chronic neuroinflammation has been implicated in familial PD. Neuroinflammation may con-

tribute to the dysregulation of protein clearance pathways, resulting in the build-up of misfolded  $\alpha$ -synuclein, mitochondrial malfunction, and cytotoxic reactions resp. oxidative/nitrative stress by the generation of reactive free radicals. Eventually, these neurotoxic processes coupled with other risk factors (age, genetic predisposition, and environmental factors) ultimately destroy the dopaminergic nigral neurons in PD. Key: BBB-blood brain barrier; PD-Parkinson's disease; SN substantia nigra; NM neuromelanin; iNOS-inducible nitric oxide/FAK pathway; TLR-Toll-like receptors; IL-Interleukin; TNF- $\alpha$  tumor necrosis factor;  $\alpha$ -syn-alpha synuclein; HSV-1 herpes simplex virus -1; Hep.C hepatitis C virus; Flavi-flaviviruses; H1N1 Influenza A virus; H5N1 avian influenza; and HIV human immunodeficiency virus

2019; Zanza et al. 2022). Microgliosis also generates pro-inflammatory cytokines including, (interleukin, IL) IL-1 $\beta$ , IL-6, and Tumor necrosis factor- $\alpha$  (TNF- $\alpha$ ), which could curtail further damage to the CNS, however, they also have the potential to be destructive to the neurons (Smith et al. 2012). Interestingly, biological features (Fig. 1) of the host such as age, gender, and obesity may affect the pathological magnitude of the cytokine storms (Fajgenbaum 2020). In addition, it was observed in a study in Jakarta that all the patients infected with the SARS-CoV-2 virus (COVID-19) that experienced a cytokine storm in the pulmonary tissue suffered from comorbidities such as hypertension and diabetes (Ramatillah et al. 2022; Gu et al. 2021). Thus,

the health status of the host plays a key role in the vulnerability to the immunopathology of cytokine storms.

Astrocytes can also release both pro and anti-inflammatory molecules and studies have shown that inflammatory cytokines such as IL-1 $\beta$  and TNF- $\alpha$  can induce astrocytes to release chemokines (CXCL10, CCL1, and CXCL13, Szpakowski et al. 2022). Thus, cytokines released from the microglia can activate astrocytes. In addition, astrocytes can modulate microglial activity. Raised levels of cytokines have been reported in serum, cerebrospinal fluid, and postmortem brain areas in PD patients (Blum-Degen et al. 1995; Mogi et al. 1994a, b; 1995a, b; 1996a, b; Wijeyekoon et al. 2020). This in conjunction with evidence for reactive microgliosis



**Fig. 2** Consequences of  $\alpha$ -synuclein aggregation in PD. Accumulates of misfolded  $\alpha$ -synuclein can provoke an array of consequences that exacerbate the burden of neuronal destruction. Including; activation of microglia, further disruption of protein metabolism, and activation

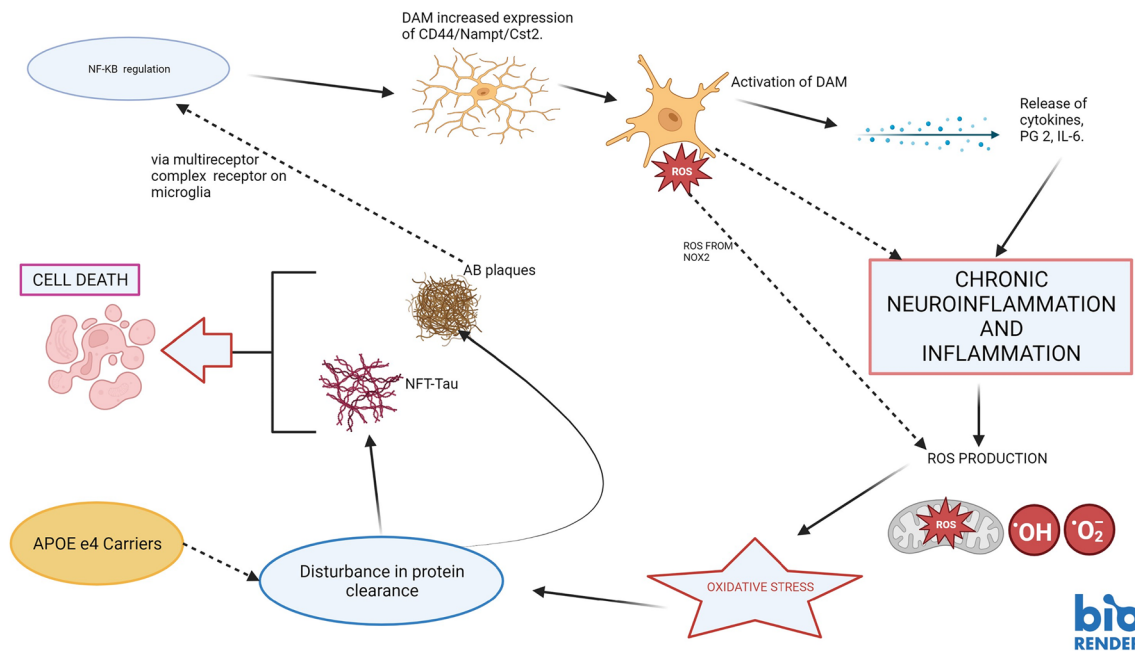
of NLRP3 inflammasome (there also may be involvement of genetic components). Subsequently, these processes marshal in a “cytokine storm” or excess release of cytokines, thereby ushering in a state of chronic neuroinflammation and neuronal death

and astrocytes in the substantia nigra (SN) of PD patients (McGeer et al. 1988; McGeer and McGeer 2004, 2007, 2008; Hunot and Hirsch 2003) and AD (McGeer et al. 1988; Eikelenboom et al. 1994; McGeer et al., 2006), reflects an activated inflammatory situation and supports the involvement of neuroinflammation in the pathogenesis of the disease. (Hirsch and Standaert 2021).

Under physiological conditions, microglia are neuroprotective and vital in maintaining neural connections. However, microgliosis may contribute to destructive inflammatory processes in the pathological state (such as PD or AD). More importantly, the SN may be particularly susceptible to the activity of the microglia since there is a fivefold greater number of these cells in this area compared to the cortex or hippocampus as found in the rat model (Kim et al. 2000). Thus, it has been suggested that microglial activation in the SN exerts a more neurotoxic state as opposed to a protective function. The damage executed by activated microglia includes the release of pro- and anti-inflammatory agents and more importantly, cellular toxins such as superoxide radicals. Thus, in the diseased state, the microglia exert a menacing role by triggering free radical-mediated oxidative

stress resulting in dopaminergic cell loss in SN and overall neurodegeneration (Simpson and Oliver 2020). Many in vitro studies and animal models of PD have demonstrated the destructive effects of activated microglia (Joers et al. 2017). In addition, a recent study reported that modulation of reactive microglia cells with cerebral dopamine neurotrophic factor supported fetal mesencephalic graft survival by attenuating the immune responses in a hemi-parkinsonian rat model (Tseng et al. 2022).

Toll-like receptors (TLRs) are expressed chiefly on microglia and play a key role in stimulating inflammatory responses. Indeed, microglial cells are involved in neuro-inflammatory processes in response to invading pathogens (viruses/bacteria) via TLRs. Interestingly, TLRs increase macrophage motility mediated via the inducible nitric oxide synthase (iNOS) /FAK pathway (Maa et al. 2011). The iNOS has been shown to exacerbate oxidative and nitritive stress (Fig. 1) which leads to neuroinflammation and loss of neuronal function in the mouse prion model (Bourgognon et al. 2021). In addition, it has been suggested that interleukin IFN- $\gamma$  pairs with TLRs in the microglia release nitric oxide which in turn produce neuronal malfunctioning (Schilling



**Fig. 3** Neuroinflammatory-mediated degenerative mechanisms that may be operating in the pathogenesis of Alzheimer's disease. NF- $\kappa$ B regulates the enhanced expression of pro-inflammatory DAM genes (Cd44, Nampt, and Cst2). An increase in the expression of these genes can prompt the activation of DAM causing the generation of inflammatory mediators (for example, PG E2, IL-6) and inflammation. Neuroinflammation can elicit cytotoxic processes such as oxidative stress, and disturbance of protein clearance pathways, thereby

resulting in a build-up of A $\beta$  deposits and tau neurofibrillary tangles. The presence of these pathological markers heralds neurodegeneration characteristics of the disease. Key: NF- $\kappa$ B nuclear factor kappa B; ROS reactive oxygen species; DAM disease-associated microglia; PG E2 prostaglandin E2; IL-6-Interleukin 6; NOX2 NADPH OXI-DASE 2; NFT-tau neurofibrillary tangles containing tau and A $\beta$  amyloid beta

et al. 2021). These findings support the involvement and key role that neuroinflammation may play in neurodegenerative disorders' pathogenesis. Interestingly, overexpression of TLRs has been documented in brain tissue and blood cells in PD (Drouin-Ouellet et al. 2014). The upregulation of the TLRs is associated with the aggravation of neuroinflammation. Thus, this led to the notion that TLRs (particularly, TLR2, TLR4, and TLR9) mediated neuroinflammation may exert a dominant neuro-destructive force that can contribute to the casualty of nigral dopaminergic cells loss in PD (Heidari et al. 2022). An overall chronic state of neuroinflammation, coupled with activation of microglial cells and hypercytokinemia can have grave consequences and in the case of PD (Fig. 2), it can prompt the characteristic destruction of nigral neurons (Tansey and Goldberg 2010).

The gathering of the activated microglia in brain areas exhibiting marked neuronal loss confirms the occurrence of destructive neuroinflammation in AD (Hansen et al. 2017). Thus, similar to PD, neuroinflammation generated by microglia has also been implicated in the pathophysiology of AD (McGeer et al. 1988; 2006; Mandreka-Colucci and Landreth 2010; Eikelenboom et al. 1994), although the two disorders differ greatly in symptoms, pathological markers, and areas demonstrating neurodegeneration.

Genetic studies suggest that microglial cells play a major role in the development of the disease and also express risk factors for AD in the brain. The presence of the AD pathological hallmarks amyloid  $\beta$  (A $\beta$ ) deposits/ plaques in close proximity to the microglia (Hansen et al. 2017) offers an opportunity for some possible interaction (s). Indeed, activation and replication of microglial cells close to A $\beta$  plaques is a characteristic siting in AD. Acute microglial inflammation may be initiated by exposure to A $\beta$  aggregates (Baik et al. 2019). Interestingly, pathomechanisms associated with AD (Fig. 3) may be linked to disease-associated microglia (DAM) (Keren-Shaul et al. 2017). The pro-inflammatory DAM genes, *CD44*, *Nampt*, and *Cst2* are implicated in AD (Rangaraju et al. 2018). NF- $\kappa$ B (nuclear factor kappa B) regulates the high expression of *CD44*, and *CD45*, which in turn leads to the activation of the proinflammatory DAM. In vitro studies suggest that the association of neurofibrillary A $\beta$  with microglia leads to the activation of these cells. This accounts for the powerful inflammatory reaction that is initiated in response to the deposition of A $\beta$  plaques in the brain (Fig. 3). It has been proposed that the fibrillar forms of A $\beta$  engage with a multicomponent receptor complex on the microglia (Bamberger et al. 2003). This interaction stimulates the

NF- $\kappa$ B mediated phenotypic microglial activation coupled with amplification of free radical production and cytokine expression. This receptor complex has been reported to play a chief role in the development of AD (Combs et al. 2001). Incidentally, activated microglia cells may also aggravate another pathological feature in AD, tau pathology (Ayyubova 2022).

Furthermore, as discussed earlier the activation of microglia can orchestrate a host of processes such as neuroinflammation, production of cytokines, and cytotoxic events elicited by reactive oxygen and nitrogen species, which can lead to the ultimate destruction of neurons (Hickman et al. 2018). In addition, the activated microglia may facilitate a chronic neuroinflammatory milieu that causes neuronal death directly or by stimulation of astrocytic-mediated neurotoxic processes (astrogliosis; Pekny and Pekna 2016) that ultimately lead to degeneration.

However, there seems to be a continuous development of neuroinflammation to profound inflammation. There is no evidence so far that neuroinflammation is detectable in the brain of young healthy individuals. However, it can be detected in brain tissue of aged people (Chung et al. 2009; DiSabato et al. 2016). Indeed, endogenous toxic metabolites like cytokines, chemokines, oxidative stress parameters etc., generated by the physiological processes of neuronal aging with loss of neurons seem to facilitate low grade activation of microglia, the resident innate immune cells. At this stage, the BBB blocks the transport of peripheral immune cells into brain tissue and there is not much antigen presentation to activate T-cells (Streit et al. 2004; Tohidpour et al. 2017). Of notion, acute and/or chronic infection, traumatic brain injury or exogenous toxic compounds may disturb the BBB and enter the brain so that peripheral immune cells, macrophages, T-cells, B-cells, and other circulating blood components pass compromised BBB and further facilitate neuronal and glial disturbances leading to massive expression of cytokines resp. chemokine levels and expressing enhanced major histocompatibility complex molecules (MHC). In addition, activated astrocytes may release endogenous compounds, inclusive growth factors (DiSabato et al. 2016; Streit et al. 2004). Depending on the type of infectious agents, viruses, bacteria, environmental toxins and their potency to compromise the integrity of the BBB and to enter certain brain areas of preference, they may or may not trigger an inflammatory response (Tohidpour et al. 2017;

Porcher et al. 2021). As such, and as the terminus technicus “neuroinflammation” has not been clearly defined and often is used to be synonymous with “inflammation, Aguzzi et al. (2013) have contributed to the definition of “neuroinflammation”. Here, we enlarge this concept and propose a staging as given in Table 1:

Impairment of the immune system affects both the innate and adaptive responses (Iwasaki 2012; Hill et al. 2022) thus contributing to the susceptibility for a variety of disorders of the aged individual with activated microglia expressing high levels of (Major histocompatibility, MHC) MHC-II and co-stimulatory antigens (Andronie-Cioara et al. 2023) like in PD and AD (McGeer and McGeer 1988).

### Modulation of microglia M1 and M2

As discussed earlier, the microglial cells appear to orchestrate a key role in the neurodegenerative processes. It is vital therefore to highlight the receptors /cytokines/factors/ signal pathways that may alter the microenvironment involved in the modulation of microglia from M2 to M1 to curtail cellular destruction (Yao and Zu 2020). The secretion of interleukin IL-4 or IL-13 from T-helper cells 2, favors the polarization of microglia to the M2 phenotype. In turn, the M2 microglia release anti-inflammatory cytokines to diminish inflammatory reactions thereby supporting the repair of injured cells. Whereas, cellular injury and the related release of pro-inflammatory cytokines TNF- $\alpha$  and interferon-gamma (IFN- $\gamma$ ), results in the microglia polarization to the M1 phenotype. Thus, it is imperative to regulate the activation of the microglia not only to maintain homeostasis in the brain but also to avert or dilute the neuro-destructive pathways (Song et al. 2016).

Indeed, an imbalance of microglial polarization, with a dominance of the M1 phenotype and malfunction of the M2 neuroprotective type, fosters the occurrence of AD (Yao and Zu 2020). Furthermore, A $\beta$  has been reported to activate microglia inflammation via the toll receptor 2 (TLR2) (Jana et al. 2008). Similarly, activation of TLR2 has been suggested to contribute to  $\alpha$ -synuclein aggregation (via the autophagy/lysosomal pathways) in PD (Dzamko et al. 2016). Collectively, this endorses the role of TLR2 in the modulation of microglial polarization. In addition, Toll receptor 4 is activated by lipopolysaccharide (present in Gram-negative

**Table 1** Is there a neuroinflammation-inflammation continuum?

- 1: The young (healthy) brain: no neuroinflammation
- 2: The aged brain: neuroinflammation with intact BBB but endogenous inflammatory processes in the absence of infectious diseases (Chung et al. 2009; DiSabato et al. 2016)
- 3: The neuroinflamed brain: infectious agents; chronic neuroinflammation but intact BBB (Yacoubian et al. 2023)
- 4: The inflamed brain: infectious agents; opening of the BBB, entry of macrophages, T-cells, B-cells, proteins etc. into brain areas causing inflammation (Mestre et al. 2021)

bacterial membrane) resulting in the polarization of M1 microglia (Ciesielska et al. 2020).

Other receptors/molecules/ factors/signal pathways involved in the polarization of microglial include (Fig. 5):

- Receptor-mediated routes for microglial modulation. For instance, the peroxisome proliferator receptor is a chief regulator of M2 phenotype and can therefore attenuate inflammatory responses (McTigue 2008).
- Neuropeptide Y has been shown to significantly hinder microglial activation by blocking the lipopolysaccharide-induced Fc-receptor-mediated phagocytosis (Ferreira et al. 2011). Therefore, neuropeptide Y may play a role in the management of microglial phagocytic activity. In vivo experiments showed that glial cell line-derived neurotrophic factor inhibited the polarization of M1 (Rocha et al. 2012).
- A transcription factor, micro RNA-124 has been suggested to manage the activity of microglia via down-regulating the expression of cytosine-cytosine-adenosine-adenosine-thymidine-enhancer-binding protein- $\alpha$ . This notion was demonstrated by the maintenance of the microglia in a quiescent state after the administration of micro RNA-124 in hyperalgesia induced in rats (Willeman et al. 2012).
- The interleukin family of cytokines is an important signaling molecule in the immune processes (Fields et al 2019). A shift of M1 to M2 phenotype polarization is supported by anti-inflammatory cytokines including: insulin-like growth factor (Labandeira-Garcia et al 2017), IL-4 (Tang et al. 2019), IL-13 (Colonna and Butovsky 2017) and IL-33 (Fu et al. 2016).
- The JAK/STAT transducer signaling pathway exerts a vital role in inflammatory reactions (Ruganzu et al 2021). STAT1 and STAT3 support the polarization of M1 microglia and consequently the release of pro-inflammatory cytokines (Lan et al 2017). In contrast, IL-4 activates STAT6, which in turn induces the appearance of M2 phenotype (Lan et al. 2017).
- Free radicals generated from cellular oxidative stress have the propensity to shift the expression of microglia from M2 to M1 (Ren et al 2020). Unregulated release of these cytotoxic species by NADPH oxidase may precipitate inflammation-related pathology. This concept

is demonstrated by the raised expression of NADPH oxidase in the microglia in AD (Choi et al. 2011).

- NF- $\kappa$ B is a principal transcription factor involved in the activation of the M1 microglia cells (Zhang et al. 2013). Kawai and co-worker (2007) reported that inhibition of NF- $\kappa$ B by modulators produced microglial polarization from M1 to M2 type.
- Transforming growth factor- $\beta$ -activated kinase 1 is a key regulator for inflammation and cell viability via activation of NF- $\kappa$ B and MAPK (Mihaly et al 2014). In addition, MAPK advocates M2 microglial polarization, which augments its anti-inflammatory actions (Zhang et al. 2019a).

Nevertheless, suppression of the M1 microglial phenotype may not be sufficient to halt inflammatory-mediated neurodegenerative pathways (Zhang et al. 2017), M2 polarization may also be required to attain full therapeutic potency (Du et al. 2018).

## Disruption of neuronal protein accumulation

### Parkinson's disease

The presence of misfolded  $\alpha$ -synuclein containing Lewy bodies in the remaining surviving nigral dopaminergic neurons represents the pathological hallmark of PD (Gibb et al. 1991). Interestingly, a mouse lentiviral model was used to mediate the selective accumulation of  $\alpha$ -synuclein in the microglia, which produced the progressive loss of dopaminergic cells (Bido et al. 2021). The inability of the protein homeostasis pathways to cope with excessive  $\alpha$ -synuclein aggregation in the microglia resulted in a “phagocytic exhaustion state” and a cellular toxic environment (perhaps due to the production of free radical species), which caused dopaminergic neuronal destruction. These findings offer a number of plausible pathomechanisms in PD, including malfunction of  $\alpha$ -synuclein homeostasis, a build-up of misfolded  $\alpha$ -synuclein, and microglial-mediated or associated cytotoxic neuroinflammation (Fig. 2). The role of environmental triggers resp. pathogens to modify the structure of  $\alpha$ -synuclein has recently been reviewed in detail (Riederer et al. 2019; Sian-Hulsmann and Riederer, 2021; Linard et al. 2022; Huynh et al. 2023). In addition,  $\alpha$ -synuclein accumulation can stimulate the microglia M-1-like polarization and impairs the microglia protein regulation mechanisms (phagocytosis and autophagy) and activation of NLRP3 inflammasomes (Lv et al. 2023). NLRP3 can also be activated by DAM to start the processing of caspase-1 and release of pro-inflammatory cytokines, IL-1 $\beta$ , and IL-8 (Blevins et al. 2022; Pike et al. 2022). This serves as



the premise of targeting NLRP3 inflammasome to block chronic inflammation in many diseases. In addition, genes implicated in PD including, LRRK2, DJ1, and GBA, also regulate these processes, thereby highlighting the role of disturbed protein metabolism in the pathogenesis of the disease (Lv et al. 2023).

Microglial cells serve an important role in the homeostasis of the  $\alpha$ -synuclein in the brain via autophagy and phagocytosis. Autophagy via microglia also blocks the NLRP3 inflammasomes invoked neuroinflammation (Lv et al. 2023). Consequently, under pathological conditions, DAM and polarized M-1 microglia may accentuate the accumulation of the aggregated  $\alpha$ -synuclein via damage-associated proteins (DAMP), although some studies suggest a neuroprotective role for DAM (Deczkowska 2018). This supports an association between the appearance of  $\alpha$ -synuclein inclusions and the dysregulation of protein metabolism in PD. DAM may support a chronic inflammation and oxidative stress state by generating reactive free radical species. Indeed, free radicals can be generated from NADPH oxidase 2 (NOX2) in DAM, and activated NOX2 is related to inflammation (Fig. 3), DAMP signaling, and  $\beta$ -amyloid plaque deposition (Simpson and Oliver 2020). Interestingly, the transcription factor Nrf2 stimulates the expression of antioxidant NADPH dehydrogenase 1 in microglia. This vital finding can be employed in the advocacy of cellular protective therapy from inflammation and related oxidative stress associated with neurodegeneration (Hickman et al. 2018).

The second mechanism of protein regulation is autophagy. Synucleinophagy is a selective autophagy that is mediated by microglia for clearance of neuron-released  $\alpha$ -synuclein. Thus, microglial  $\alpha$ -synuclein degradative pathway appears to exert a vital role in neuroprotection. This notion is demonstrated by the enhanced accumulation of incorrectly folded  $\alpha$ -synuclein coupled with the destruction of nigral dopaminergic neurons, due to disturbances inflicted on microglial autophagy in mice expressing human  $\alpha$ -synuclein (Choi et al. 2020). Experiments using knock-down DJ-1 genes in microglia showed an impaired autophagic uptake and clearance of  $\alpha$ -synuclein and dysfunction of autophagy-dependent of p62 and LC3 proteins (Nash et al. 2017). Indeed, the synucleinophagy clearance of misfolded  $\alpha$ -synuclein is mediated via TLR4-NF- $\kappa$ B-p62 (Choi et al. 2020). The cellular protective role of DJ-1 against oxidative stress is demonstrated by enhanced vulnerability to the neurotoxicity of dopamine cells in DJ-1 deficiency in microglia (Ariga et al. 2013). Furthermore, DJ-1 mutations ascribe to the autosomal recessive form of PD *park 7* (Repici and Giorgini 2019). Although out of all the causative genes (PARKIN, PINK-1) related to familial PD, DJ-1 mutations only account for a small number. In addition, the presence of the oxidized form of DJ-1 in the brains of PD patients betrays the operation of

cytotoxic oxidative stress. These findings support the concept of the involvement of some underlying genetic default that ascribes/contributes to the overall build-up of misfolded  $\alpha$ -synuclein in PD.

Therefore, although microgliosis does not appear to directly initiate  $\alpha$ -synuclein aggregation, it can evidently contribute to the overall total load of misfolded protein generally. Indeed, PD animal experimental models demonstrate and endorse a principal role of activated microglia and NLRP3 inflammasome in both  $\alpha$ -synuclein neuropathology and dopaminergic neuronal loss (Pike et al. 2022). Incidentally in experimental studies, dopamine has been shown to halt the activation of human microglia NLRP3 inflammasome (Pike et al. 2022). This may account for the vulnerability of the depleted nigral dopamine neurons to the onslaught of cytotoxic neuroinflammatory processes in PD (Fig. 2).

Analogous to PD, the involvement of aggregation of rogue proteins is also a characteristic pathological feature in AD. The presence of abnormal or misfolded A $\beta$  peptide deposits and neurofibrillary tangles (NFTs) containing tau protein in the brain is a characteristic and well-documented histopathological observation in AD (Alzheimer 1907 resp. Alzheimer et al. 1995).

Generally, infections produced by particular pathogens such as viruses or bacteria (*helicobacter pylori*) or fungi (Malassezia) have been linked with an enhanced risk of the manifestation of PD (Wang et al. 2022; Limphaibool et al. 2019). A recent epidemiology study (Cannon and Gruenheid 2022), postulated the “multiple microbe” hypothesis and found that the risk of PD was greater in subjects that were seropositive to a number of microbes (viruses, bacteria, and fungi) compared to controls that were negative. It was suggested that the manifestation of PD-like symptoms due to these microbial infections is probably related to common pathophysiological processes employed including systemic neuroinflammation, accumulation of misfolded  $\alpha$ -synuclein, and dysfunction of mitochondrial respiratory pathways. Also, in the cases of familial PD, the gene mutations may also influence the host-microbe interactions. Indeed, the familial PD genes (PARK2, LRRK2, and PINK 1) have been shown to cause misregulation of immune mechanisms (Figs. 1, 2) and thus genetically mediated modulation of immune consequences may exercise a more integral role in neurodegeneration (Patrick et al. 2019; Baizabal-Carvalho and Alonso-Juarez 2021).

The advent of **post-encephalitis parkinsonism (PEP)** was exhibited after about 10 years in around 50% or more of those that survived encephalitis lethargica produced by **H1N1 influenza virus** (von Economo’s disease) between 1915 and 1927 (Jankovic and Tan 2020; Foley 2009). Interestingly, it shares many features in common with the idiopathic form of PD including motor abnormalities, dystonia,

and neuropathological changes such as degeneration of SN neurons (Davis et al. 2014; Jankovic 2022; Toledano and Davies 2016). However, the characteristic “pill-rolling” movement observed in PD is absent. In addition, there appears to be a difference in the time frame for pathological development in the case of PEP, the loss of dopamine neurons associated with the viral infection (Cocoros et al. 2021) and it shows slower progression compared to idiopathic PD (Jang et al. 2009a).

It has been postulated that two possible mechanisms are related to viral-related parkinsonism (Sulzer et al. 2020). The first one is para-infectious and produces almost immediate neuronal destruction coupled with parkinsonism, whereas the second mechanism is post-infectious and produces a slower neuronal cell death. The two mechanisms are not exclusive and can occur concomitantly. More importantly, both pathways of viral-induced cellular degeneration are mediated through neuroinflammation. Furthermore, there are some hypotheses offered related to viral-associated neurodegeneration (Sulzer et al. 2022). The findings related to PEP led to the evolution of the “dual-hit” hypothesis to the development of PD (Hawkes et al. 2007). It suggests that a neurotropic pathogen (probably of viral origin) gains entry to the brain via two sites, the olfactory bulb and gastric mucosa via the vagus nerve, thus the name dual-hit hypothesis (Fig. 1). At these sites, the neurotropic pathogen triggers  $\alpha$ -synuclein pathology (Braak et al. 2003). Subsequently, the culprit pathogen is associated with neuroinflammation leading to the destruction of the nigral cells. Alternatively, another virus-related neurodegeneration theory has been postulated, the “hit and run” hypothesis (Scarlsbrick and Rodriguez 2003). This suggests that the virus invasion can initiate a powerful inflammatory response that can elicit cytotoxic neuronal destruction which can still operate even after viral load has been cleared (Hara et al. 2021). The “hit and run” hypothesis appears to suggest a very viable possibility for viral-related pathogenesis in neurodegeneration.

The neuropathology of PEP shows marked neuronal loss and gliosis throughout the brainstem with a focus on the SN. PEP is a tauopathy and NFTs have been demonstrated in nearly all brain areas of interest (Jellinger 2009; Hoffmann and Vilensky 2017). However, and in contrast to progressive supranuclear palsy (PSP) but similar to AD NFTs are composed of both 3R and 4R isoforms (Hoffmann and Vilensky 2017). Of note, LBs and Lewy neurites or any other  $\alpha$ -synuclein deposits are absent in PEP (Jellinger 2009).

As human post mortem studies from PEP to identify viral particles so long after the pandemic are difficult and connected with many problems (see Cadar et al. 2021; McCall et al. 2001; Reid et al. 2001) more recent experimental work using **H5N1 influenza virus** gives way to understand, at least in part, the mechanisms underlying virus-induced pathology leading to neuropsychiatric disorders. Jang et al.

(2009b) demonstrated that H5N1 spreads from the peripheral nervous system to the central nervous system. There the virus interacted with  $\alpha$ -synuclein phosphorylation and aggregation, caused nigral degeneration with loss of dopaminergic neurons in the SNpc as well as activation of microglia and neuroinflammation (Jang et al. 2009b). Depending on the experimental conditions H5N1 infection in the long-term induces transient loss of the dopaminergic phenotype but causes long-lasting inflammatory response (Jang et al. 2012).

PEP following exposure to other viruses has been frequently reported in man and rodents (Jang et al. 2009a; Leta et al. 2022). The viruses include influenza A (Gamboa and Yahr 1973), flaviviruses such as West Nile Virus (Robinson et al. 2003; Sejvar 2014), Japanese B encephalitis, St Louis encephalitis, Hepatitis C (Wang et al. 2022), HIV retrovirus and SARS-CoV-2 RNA positive virus. Strikingly, infections with these viruses may result in the manifestation of some cardinal parkinsonian features such as tremor, cogwheel rigidity, and bradykinesia (Beatman et al. 2016) and pathology in the midbrain.

**Flaviviruses** proteins orchestrate the rearrangement of the endoplasmic reticulum membrane required to form replication complexes for the virus (Barrett and Weaver 2012; Sempere and Armando 2019; Thomas et al. 2014). Subsequently, flavivirus reproduction is accompanied by reactive oxygen species-induced endoplasmic reticulum oxidative stress and apoptotic destruction of virally infected cells (Zhang et al. 2019b).  $\alpha$ -Synuclein expression was found to be increased in primary neurons infected with West Nile Virus (Beatman et al. 2016). Thus, native  $\alpha$ -synuclein executes a neuroprotective role which is lost in the pathogenesis of PD. Indeed, it has been documented that deletion of the  $\alpha$ -synuclein gene in mice leads to an increase in viral propagation in the brain coupled with a decrease in neuronal survival rate (Beatman et al. 2016). In contrast, no reports were found relating Japanese encephalitis virus and  $\alpha$ -synucleinopathy, thereby suggesting a different mode of pathogenesis (Misra 2002). Although West Nile Virus and Japanese encephalitis virus as well as mosquito-borne alphavirus infect and cause vast cellular damage to the brain, particularly in the dopaminergic systems including the SN, basal ganglia, and other regions, cerebellum, and brain stem (Solomon 2004; Bantle et al. 2019), the Parkinsonian symptoms appear to be temporary and resolve over time. Another flavivirus, the St Louis encephalitis virus is closely related to the Japanese encephalitis virus. It also produces brain inflammation or encephalitis and consequent parkinsonism (Curren et al. 2018). Up to 20% of Dengue virus infections affect the brain with encephalitis, encephalopathy and movement disorders such as parkinsonism (Hopkins et al. 2022).

**Filovirus**, like Ebola virus infrequently demonstrate neurological signs, but when present they consist of meningitis,

encephalopathy and seizures (Sagui et al. 2015). Ebola virus is suggested to bypass the BBB. Using quantitative MR-relaxometry, 18F-fluorodeoxyglucose PET and immunohistochemistry Schreiber-Stainthorp et al. (2021) in a monkey model describe BBB disruption with extravasation of fluid and albumin into extracellular space. The regional hypermetabolism corresponded to mild neuroinflammation, sporadic neuronal infection and apoptosis as well as increased glucose transporter-3 (GLUT-3) expression (Schreiber-Stainthorp et al. 2021).

Epidemiological evidence is indicative of an association between **Hepatitis C** infections and the manifestation of PD. Indeed, a Taiwanese study reported that a previous infection with the Hepatitis C virus increased the risk of subsequent development of PD (Odd ratio 1.39; Wu and Kang 2015). These findings were confirmed subsequently by Wijarnpreecha and colleagues (2018). The virus primarily causes neuroinflammation-mediated neuronal destruction. It enhances the release of pro-inflammatory cytokines, which leads to the onset of cytotoxic processes particularly in the dopaminergic areas of the brain. The predilection of the pathology in dopamine neurons probably ascribes the subsequent risk for the development of PD. Also, the virus is associated with the downregulation of the astrocyte-derived factor (this is neuroprotective), which exacerbates neuronal cell destruction (Wijarnpreecha et al. 2018).

Recent evidence has been provided to demonstrate a link between **Severe Acute Respiratory Syndrome Corona Virus-2 (SARS-CoV-2)** and PD. Coronavirus disease 2019 (COVID-19) affects the lower and upper respiratory tract with involvement of the peripheral and central nervous system (Harrison et al. 2020; Wu et al. 2023). As such the virus displays difficulties in breathing, cough, fever, headache, loss of taste and/or smell, fatigue and several neurological manifestations including acute cerebrovascular diseases, conscious disturbances, loss of cognitive abilities, attention disorder, anxiety, insomnia, depression and eventually problems of motor coordination as well as typical non-motor symptoms accompanying PD (Conte et al. 2021; Juan et al. 2021; Saniasiaya et al. 2019; Lopez-Leon et al. 2021; Premraj et al. 2022; Efstathiou et al. 2022; Wang and Perez 2023). Therefore, the notion is of interest that human post mortem studies demonstrated neuropathological findings in several brain regions including the basal ganglia, thalamus, hippocampus cingulum and cortical areas (DaSilva-Torres et al. 2022).

TLRs, mainly TLR4 activation seem to cause neuroinflammation (Heidari et al. 2022). SARS-CoV-2 spike protein might bind to TLR4 and the interaction of SARS-CoV-2 and  $\alpha$ -synuclein has been discussed by Conte (2021). As ACE2-virus binding is low in the CNS Szabo et al. (2023) discuss the interaction of SARS-CoV-2 and TLR2. TLR2 is an innate immune receptor, which interacts with multiple viral

components including the envelope protein of SARS-CoV-2 and plays an important role in PD and Alzheimer disease.

Szabo et al. (2023) hypothesize that TLR2 may play a critical role in the response to SARS-CoV-2 infiltration in the CNS.

In vitro studies demonstrate that SARS-CoV-2 proteins directly interact with  $\alpha$ -synuclein and cause Lewy-like pathology in the presence of  $\alpha$ -synuclein overexpression (Wu et al. 2022).

SARS-CoV-2 nucleocapsid (N) protein speeds  $\alpha$ -synuclein aggregation, while SARS-CoV-2 spike (S) protein has no effect on  $\alpha$ -synuclein aggregation (Mesias et al. 2022), thus providing molecular insights into the impact of SARS-CoV-2 post-infection and the oligomerization of  $\alpha$ -synuclein (Semerdzhiev et al. 2022).

SARS-CoV-2 infection in Macaques with mild-to-moderate disease in post mortem brain analyses showed activated microglia in the parenchyma as well as intracellular  $\alpha$ -synuclein aggregates (Philippens et al. 2022). In addition, SARS-CoV-2 infection of transgenic mice expressing human angiotensin converting enzyme 2 (ACE 2) drives NLRP3 inflammasome activation in human microglia (Albornoz et al. 2022). Combined experimental and computational methods show moderate binding between two SARS-CoV-2 proteins and  $\alpha$ -synuclein (Mesias et al. 2022). Altogether, such data suggest SARS-CoV-2 related neurological disorders, which should be confirmed by human post mortem studies (Riederer and Ter Meulen 2020). Indeed, Müller and Benedetto (2023) and Hill et al. (2022) discuss that ineffective immune response may be the reason that the virus is present for longer times, involves the vascular system, spreads within the brain and facilitates processes as that given above leading to chronic neuroinflammation with an increasing risk for the long-covid syndrome (NICE 2020; Müller and Riederer 2023) with psychiatric symptomatology, depression (Benedetti 2023), fatigue and cognitive impairment (Penninx et al. 2021; Riederer and Ter Meulen 2020).

SARS-CoV-2 and its link to  $\alpha$ -synuclein pathology opens the door to speculate about the potency of this virus to trigger parkinsonism. Wang et al. (2023a) conclude a two-year retrospective cohort study following COVID-19 infection is associated with a higher risk of developing neurodegenerative disease, that the risk of developing PD is increased for the first year after COVID-19 but that there is no further risk of PD after that time (Wang et al. 2023b). Indeed, the appearance of parkinsonism during or immediately after COVID-19 infection represents a VERY rare event (Cavallieri et al. 2022; Akilli and Yosunkaya 2021). However, movement disorders started 13 days on average after the initial onset of COVID-19 and in a fraction of patients (22%) ataxia, myoclonus, tremor, parkinsonism was still present after a follow-up period of  $75 \pm 3$  weeks (Schneider et al. 2023).

To conclude this, it is known that SARS-CoV-2 affects neuronal populations relevant for the pathogenesis of PD, but there is no evidence that SARS-CoV-2 triggers neurodegeneration in the acute phase of infection and the early post-infection phase (Lingor et al. 2022; Szabo et al. 2022). This concurs with experimental data in which mice were administered a non-lesion dose of the parkinsonian toxin 1-methyl-4-phenyl-tetrahydropyridine (MPTP) infected with SARS-CoV-2 (Smeyne et al. 2022). There was no loss of dopaminergic neurons in the SNpc but a significant increase in microglial activation in the striatum and the SNpc (Smeyne et al. 2022). Of note, human post mortem studies from brain autopsies in COVID-19 do not show lesions of the nigrostriatal system (Mukerji and Solomon 2021; Goertler et al. 2022; da Silva-Torres et al. 2022; Martin et al. 2022; Crunfli et al. 2022; Solomon et al. 2023), although viral RNA is still present several days after death (Skok et al. 2021). This indicates that there is no evidence of neurotropism for SARS-CoV-2 (Lebrun et al. 2023) although the outcome of the long-covid syndrome cannot be judged at this time (Davis et al. 2023). However, late-stage PD may be highly susceptible to critical COVID-19 infection and risky outcome (Zhai et al. 2021).

**MERS-CoV** infection induces a strong inflammatory response already in the acute phase (Mahallawi et al. 2018). In human DPP4 transgenic mice infected with MERS-CoV viral antigens were detected in brain areas four days post-infection with affection of neurons and astrocytes followed by a damage of the BBB, microglia activation and inflammatory cell infiltration, which may be caused by complement activation (Jiang et al. 2021).

**Human immunodeficiency virus (HIV)** type 1 transactivator of transcription (HIV-1Tat) protein can promote the activity of matrix metalloproteinases (MMPs) such as MMP-9 in astrocytes, which has the potential to damage the BBB and promotes the monocyte infiltration into the CNS (Sung et al. 2009; McArthur et al. 2003). This influx of monocytes symbolizes an early inflammatory event in HIV-associated encephalitis, dementia, and neurocognitive disorders (HAND) (Marino et al. 2020). In addition, mice injected with HIV-1 Tat lead to the activation of cyclooxygenase 2, which in turn subdued the expression of the tight junctions thereby encouraging the opening of the BBB (Pu et al. 2007).

HIV infection affected several million adults and children worldwide and it has been estimated that at least one third of the adults will develop a dementing illness (Janssen et al. 1992; McArthur et al. 1993; see also Koutsilieri et al. 2001) with a poor prognosis (Harrison et al. 2008). HIV-induced dementia with cognitive dysfunction and behavioral abnormalities is associated with motor disorders inclusive parkinsonism. Absence of a direct viral infection of neurons has been described (Takahashi and Yamada 1999)

in contrast to glial cells (astrocytes, microglia; Wahl and Al-Harhi 2023) and it has been assumed that neurotoxins released by infected cells are causal for neurodegeneration leading to a dementia-parkinsonism syndrome (Glass and Johnson 1996). Excitotoxicity has been a focus of research to enrol the mode of action of the HIV dementia-parkinsonism syndrome (Koutsilieri et al. 2001). In fact, enhanced glutamatergic activity has been detected in patient suffering from AIDS as well as in HIV-positive patients without AIDS symptomology as summarised by Koutsilieri et al. (2001). Moreover, this group suggested dysregulation of the dopaminergic system as a pathogenic factor for NeuroAIDS (Koutsilieri et al. 2001) with a loss of dopamine in SIV-infected monkeys (Koutsilieri et al. 2002a, 2002b). Studies with the simian model of HIV as well as the chronically HIV-infected T-lymphoblasts ACH-2 demonstrated that dopamine induced a concentration dependent HIV activation (Scheller et al. 2000), a finding agreeing with those showing that dopaminergic substances such as L-DOPA and the monoamine oxidase inhibitor selegiline accelerate generation of vacuoles in the simian model of HIV infection (Czub et al. 1999, 2001, 2002b; Koutsilieri et al. 2002a) as is with metamphetamine in a HIV-cat model (Phillips et al. 2000). Indeed, dopamine is involved in the regulation of HIV gene expression in neuronal cells and cells of the immune system (Sawaya et al. 1996; Rohr et al. 1999). In contrast, memantine, a glutamatergic N-methyl-D-aspartate receptor antagonist, is beneficial in treating HIV infection in the simian model of HIV (Meisner et al. 2008). HIV-positive individuals show evidence for an activated brain innate immune response and increased brain inflammation associated with poorer cognitive performance (Vera et al. 2016; Wojna 2022) and depression (Mudra-Rakshasa-Loots et al. 2022). Of interest, experimental work in which supernatants of primary monocyte-derived macrophages (MDM) infected with HIV (HIV/MDMs) were used to treat primary cultures of rat oligodendrocyte precursor cells (OPC) during their differentiation to mature oligodendrocytes, HIV/MDMs inhibited OPC maturation (Roth et al. 2021). Additional studies showed that glutamate or AMPA- respective kainic acid receptor agonists phenocopied this effect, probably by inducing an excitotoxic stress response (Roth et al. 2021).

The notion is of interest that depending on the dose of the virus MRI studies of the brain demonstrate variation in the intensity of brain area lesions. For example, HIV shows predominantly cortical atrophy and basal ganglia lesions, EBV demonstrates striatal hyperintensity and basal ganglia necrosis, West Nile Virus (WNV) has a focus on bilateral basal ganglia, thalamus or pontine lesion, while Japanese Encephalitis Virus (JEV) shows bilateral thalamic involvement or substantia nigra lesion (Leta et al. 2022). PEP demonstrates widespread neurodegeneration in subcortical and brainstem areas (Itoh et al. 2000; Jellinger 2009). Such

variations in the intensity of brain area lesion seem to have a clinical relevance insofar as the phenomenology shows differences with the consequence of levodopa treatment response to parkinsonian symptoms (Leta et al. 2022). In fact, HIV, EBV, WNV show poor levodopa response, while JEV PD symptoms can be treated successfully with levodopa and influenza virus demonstrates mild improvement (Leta et al. 2022).

As the pattern of virus-induced degenerative processes varies with regard to brain areas and especially striatum and substantia nigra we conclude that it is predominantly the nigro-striatal dopaminergic system that is involved in virus-induced neurodegeneration, focusing either the striatum (poor levodopa response (MSA-P type related?) or the substantia nigra (good levodopa response; PD type).

Of note, Theiler's virus and other viruses such as neurotropic strains of mouse hepatitis virus readily and relative selectively infect the SN-tyrosine hydroxylase positive neurons following intranasal inoculation of mice (Oliver et al. 1997). Therefore, it is not farfetched to assume that the dopaminergic neurons of the SN play a primary role in the debilitating viral attack. This is, because in humans about 80–90% of SN neurons contain neuromelanin (NM; Hirsch et al. 1988, 2003). NM under physiological conditions binds neurotoxic substances/pathogens and eventually concentrate them but may release them under conditions of changes in the composition of the cytoplasm (Nagatsu et al. 2023; Riederer et al. 2021, 2023).

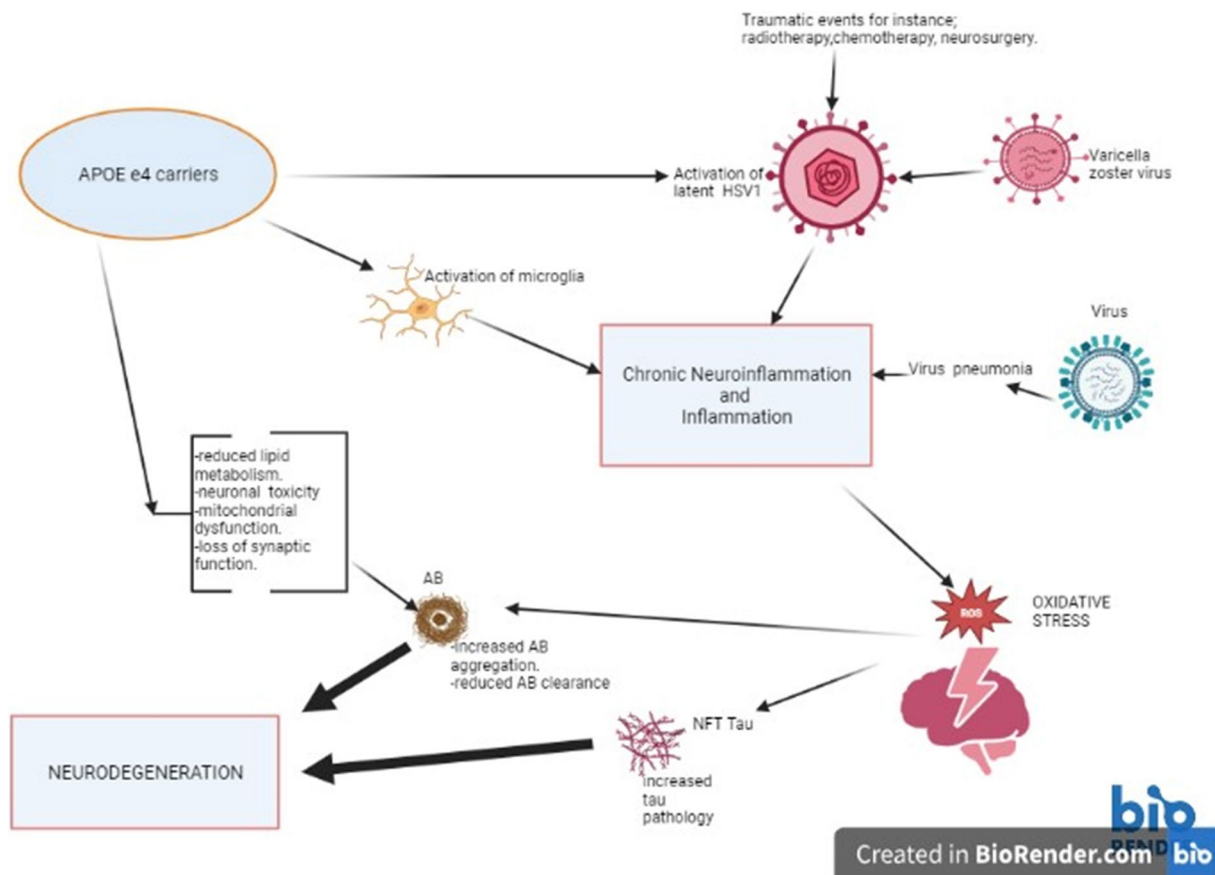
## Alzheimer's disease

There is compelling evidence linking these pathological markers and inflammatory agents/factors. Transgenic mice (TgCRND) treated with an NLRP3 inflammasome inhibitor, JC-124 were shown to decrease microgliosis, A $\beta$  deposits, and oxidative stress (Yin et al. 2018). Another risk factor for inflammation is apolipoprotein Ee4 (APOE e4) (Shi et al. 2017). Interestingly, experiments using animals with a knockdown of the apolipoprotein E gene, show the correct phagocytic/autophagic operation of the microglia/astrocytes to salvage the cells from tau pathology (Wang et al. 2021). The apolipoprotein E gene (Fig. 4) is a risk factor for late-onset AD (Corder et al. 1993). In addition, activation of prostaglandin 2 (which acts as a pro-inflammatory cytokine) is associated with the inhibition of A $\beta$ -related microglial phagocytic activity (Cai et al. 2022), thereby reflecting an inflammatory agent-induced defect in the microglial-autophagy/phagocytic pathway resulting in the aggregation of the misfolded proteins. IL-6 is a cytokine related to inflammation and infection. It has been shown to build up the hyperphosphorylated tau protein (Quintanilla et al. 2004).

It is pertinent to mention that physiologically, microglia exert a key role in the homeostasis of these proteins by phagocytosis of A $\beta$  and tau proteins (Mandreka-Colucci and Landreth 2010). It clears the soluble form of A $\beta$  via micropinocytosis and fibrillar A $\beta$  peptides by phagocytosis. Thus, microglia serves a neuroprotective role since misfolded A $\beta$  aggregates and tau neurofibrilles are toxic to neurons (Yang et al. 2017). However, as previously stated this protective role may be lost in a chronic state of neuroinflammation, resulting in the reduced clearance and consequent accumulation of these unwanted proteins and thus the manifestation of A $\beta$  plaques and NFTs (Cai et al. 2022). Postmortem studies have shown activated microglia to enclose A $\beta$  deposits and tau-containing dying neuronal cells (Serrano-Pozo et al. 2011). Studies using a transgenic AD mouse model show that tau pathology proceeds through microglial activation via the release of cytokines (Maphis et al. 2015). In addition, the difunctional microglia may contribute to the spread of tau pathology (Ayyubova 2022). Strikingly, there have been conflicting findings from postmortem studies showing that tau pathology is more affiliated with neurodegeneration, loss of neuronal synapses, and cognitive deterioration than  $\beta$ -amyloid-accumulated deposits. In contrast, studies of tau pathology in the mouse model have dispelled any involvement of NFTs with memory loss and neuronal cell destruction (Berger et al. 2007). PET studies have demonstrated an association of the tau NFTs with heightened neuroinflammation in the later prodromal stages of AD (Ismail et al. 2020). This concurs with the amyloid cascade hypothesis, which suggests that neuroinflammation mediates the amyloid aggregations during disease progression to prompt tau pathology. Subsequently, the tau aggregates can further exacerbate neuroinflammation and neuronal destruction (Sun et al. 2021). Indeed, experiments using the AD mouse model, show that rutin (a flavonoid glycoside) barred the development of tau aggregation and tau-related cytotoxicity and neuroinflammation (Sun et al. 2021). Some studies have even suggested that the A $\beta$  toxicity is dependent to a certain degree on the presence of NFTs (Roberson et al. 2007).

There has been an association between AD and infections with some pathogens such as influenza virus, **Herpes simplex virus -1 (HSV1)**, SARS-CoV-2 virus, HIV, bacteria (Spirobacteria, Streptococcus pneumonia). Around 5.9% of viral encephalitis patients subsequent to a viral infection developed AD (Levine et al. 2023). Furthermore, viral encephalitis increased the risk by 31-fold to develop AD, and are 40 times more likely to have dementia (Levine et al. 2023).

Indeed, viral infections including influenza, and pneumonia exhibited similar associations with dementia (Levine et al. 2023). However, others have disagreed with such an association (Imfeld et al. 2016). Nevertheless, the infectious agent may induce systemic and chronic neuroinflammation,



**Fig. 4** Apolipoprotein E e4 (ApoE e4) function in Alzheimer's disease, Carriers of APOE e4 gene bestow the operation of many cellular toxic actions. These events are exacerbated in the presence of particular viral infections (HSV-1, influenza virus, and varicella-zoster virus).

These pathways share a common microglial-mediated neuroinflammatory route that eventually contributes to the appearance of pathological markers and neurodegeneration typical of the disease. Key: HSV-1 herpes simplex virus -1

which consequently produces cellular/neuronal damage and destruction (Piekut et al. 2022.) In addition, the viral pathogen may also foster typical pathological changes observed in AD, such as A $\beta$  aggregates, and stimulate hyperphosphorylation of tau. The presence of AD pathological markers due to virus/pathogen-related infections endorses the relevance of translating the findings from such studies to the pathogenic processes occurring in AD. A study that employed frequent intermittent lipopolysaccharide-induced infections in mice were reported to impair cognition, disturbed neuronal communications, and had some permanent effects on brain function (Engler-Chiurazzi et al. 2023). Extrapolation of these findings to man would imply that neuroinflammation induced by pathogenic-related infections may have produced a collective destructive effect (s) on the cells. Subsequently, in advancing age, the cells may not be so robust (diminished brain plasticity with age) and they succumb to the onslaught of the cytotoxic processes related to chronic neuroinflammation triggered by the pathogen/virus, leading to cognitive decline amongst other neurodegenerative effects. Indeed, a

recent cohort study reported that the influenza vaccination decreases the risk of AD in adults 65 years and above (Bukhbinder et al. 2022).

Similarly, activation of latent HSV1 (present in the brain) has been linked to the manifestation of AD (Mangold and Szpara 2019). Epidemiological studies furnish evidence indicating that frequent HSV1 infection poses a potential risk factor for the illness (Piacentini et al. 2014). Under normal conditions HSV1 resides dormant in the neurons, however, when it is activated it can produce signature pathological changes typical to AD (Cairns et al. 2022). Varicella zoster virus-related infections have been suspected to cause activation of the latent HSV1 virus. In addition, traumatic events such as chemotherapy and radiotherapy can also reactivate HSV1 in the brain (Itzhaki 2016, 2017). The recurrent HSV1 infection probably poses a threat due to neuronal events associated with HSV1 infections such as neuroinflammation, oxidative stress, A $\beta$  accumulation, hyperphosphorylation of tau proteins, and disruption of synaptic cellular communications (Jamieson et al. 1991). These events make up the very

foundations for the pathogenesis of AD. Other factors may be involved in viral-specific susceptibility to the illness, such as a genetic component. Indeed, numerous studies have supported the significant risk factor for AD (Fig. 4) in ApoE e4 carriers in association with HSV1 infection (Itzhaki 2017). In contrast, carriers of another isoform, the ApoE e2 allele are protected (Husain et al 2021). Nevertheless, although ApoEe4 is the strongest risk factor for AD, that does not imply that carriers will assuredly develop the disease but they do carry the risk (Yamazaki et al. 2019). Physiologically the isomers of ApoE differentially control the accumulation and clearance of A $\beta$  in the brain and the capacity of binding lipids. However, the presence or expression of ApoE e4 is linked to two mechanisms that can result in neurodegeneration. First, the ApoE e4 may participate in the pathogenesis of AD via A $\beta$ -independent processes, including neuroinflammation, disturbing neuronal-synapsis function, and cholesterol homeostasis. In contrast, the second mechanism is related to the aggregation of A $\beta$  deposits and the cytotoxic events triggered by these deposits (Verghese et al. 2013). Fascinatingly, it has been suggested that the pathogenesis of AD may be linked to mishandling of lipids (Yin 2022). This idea is supported by *in vitro* studies using human astrocytes, that demonstrated ApoE e4 to disrupt cellular lipid homeostasis (Sienski et al. 2021). Consequently, this can result in cellular destruction, since imbalances in lipids can hinder many vital cellular processes. Therefore, in the diseased state (Fig. 4), the disturbed lipid homeostasis may exert catastrophic effects such as disrupting protein metabolism pathways (which may lead to the build-up of A $\beta$ ), amyloidogenesis, disturbing cell signaling, disrupting cellular calcium homeostasis, damaging the blood–brain barrier (allowing easier access to pathogens into the brain) and support the operation of oxidative stress and neuroinflammation (Chew et al. 2020).

### The relevance of Janus-faced molecules in the pathogenesis of PD and AD

There are some molecules/ enzymes/proteins present in the human body that can serve both a physiological and/ or a pathological role, thus they are referred to as Janus-faced molecules, in reference to the two-faced Roman god (Sierra et al. 2013). These include: transcription activators, pro-inflammatory cytokines (such as IL-6), nitric oxide,  $\alpha$ -synuclein,  $\beta$ -amyloid, and microglial cells.

Inflammatory stimuli including pathogens (such as viruses and bacteria) or non-pathogenic products, or signal molecules (including cytokines, interleukin Il-1 $\beta$ , Il-6) for cellular destruction can interact with toll receptors and receptors (Kaminska et al. 2005). This leads to the activation of these receptors that can trigger a host of important

inflammatory pathways including Janus kinase (JAK)-signal transducers, nuclear factor kappa B (NF- $\kappa$ B), mitogen-activated protein kinase (MAPK), and STAT pathways (activator of transcription), finally resulting in the activation of inflammatory cells (Hendrayani et al. 2016; Chen et al. 2017).

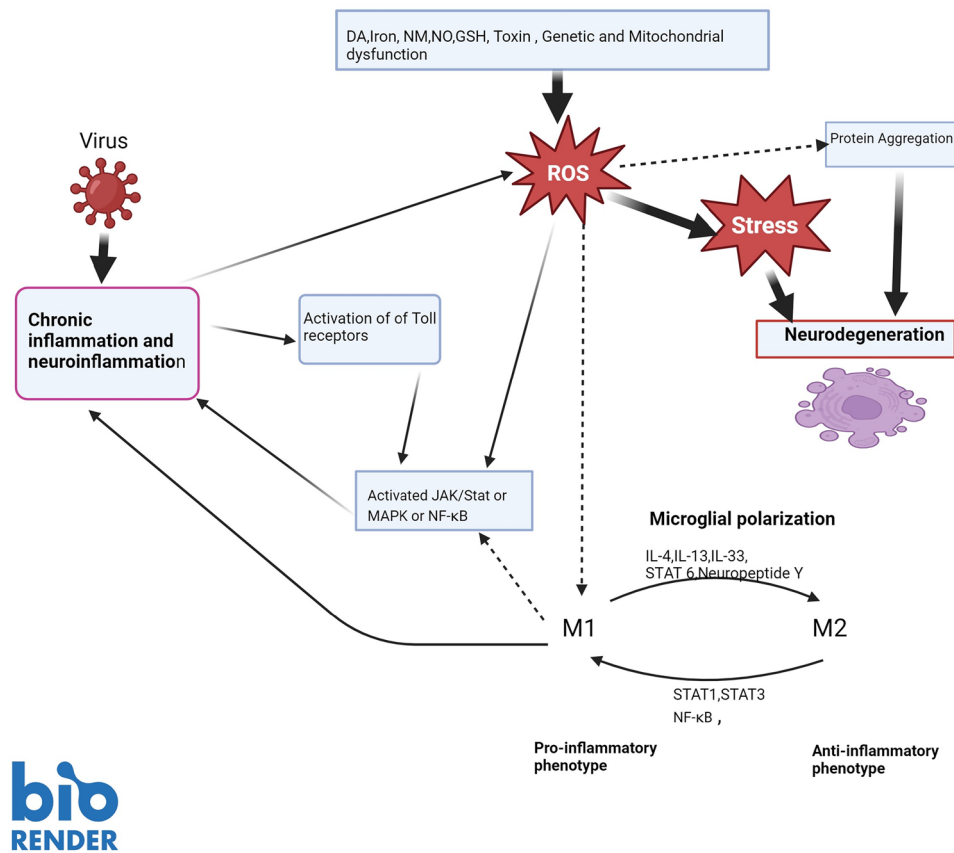
The Janus kinase signal transducers (Fig. 5) activate the transcription of the JAK/STAT pathway, which mediates the cellular response to cytokines such as IL-6. JAK-related receptors are activated by ligands via phosphorylation thereby forming docking areas for latent cytoplasmic STAT transcription factors (Harrison 2012). In addition, the binding of the IL-6 cytokines membrane receptors in turn activates the JAK-STAT pathway prompting the transcription of inflammatory genes and activation of cytokines (Boengler et al. 2008). Physiologically the signaling of JAK/STAT is vital for many homeostatic and development processes including immune cell development, and hematopoiesis (Ghoreschi et al. 2009).

However, an imbalance of the JAK-STAT pathway, MAPK, or NF- $\kappa$ B pathways is related to inflammatory, metabolic, and autoimmune diseases (Oekinghaus et al. 2011). This may be ascribed to transcription factor-related cytokine secretion and in addition disturbances on a genetic level since many of the transcription agents regulate inflammatory genes including TNF- $\alpha$ , IL-1, and IL-6, interferons (Rahman and Adcock 2006).

In addition, JAK mutations lead to many diseases such as severe combined immune deficiency, hyper IgE syndromes, and cancer (Jatiani et al. 2010). This highlights the pivotal role of this signaling transducer in the inflammatory response of the immune system, thereby offering a lucrative site for targeting anti-inflammatory therapy.

Interestingly, elevated free radicals (and 8-hydroxy-2-deoxyguanosine) generated from oxidative stress can activate JAK/STAT and other transcription factors, resulting in the enhanced expression of pro-inflammatory cytokines and chemokines. Consequently, this prompts the immune system to respond to inflammatory reactions (Al-Samahari et al. 2015). Extrapolation of these findings to the brain in the diseased state may suggest that perhaps the products of oxidative (and nitrogen) stress such as the free radicals may prompt cytotoxic neuroinflammatory pathways, thereby augmenting the neuronal destruction.

Nitric oxide (NO) is produced by nitric oxide synthase during the conversion of L-arginine to L-citrulline. It plays multiple physiological roles including vasodilation, neurotransmission, regulation of neuronal survival, and activation of the JAK signal transducer. Subsequently, superoxide free radicals (produced by oxidative stress) may react with NO to produce highly oxidant peroxynitrite, which can cause further oxidative cellular damage to DNA, lipids, and proteins (Calabrese et al. 2009; Piacenza et al. 2022). In addition, NO is a potent and highly reactive free radical in its own right, it



**Fig. 5** Signal molecules and proteins associated with chronic inflammation and neuroinflammation resulting in neurodegeneration. Molecules released from inflammatory processes can activate Toll-receptors, which in turn stimulate signal molecules (such as JAK/STAT or MAPK or NF- $\kappa$ B) and their pathways. In addition, pro-inflammatory M1 microglia phenotype can also stimulate these signal pathways. The polarization of the M1 microglial is supported by, NF- $\kappa$ B, STAT1, and STAT3, whereas the appearance of anti-inflammatory M2 microglial is favored by IL-4, IL-13, IL-33, STAT 6 and Neuropeptide Y. Consequently, chronic inflammation and neuroinflammation generate reactive oxygen species that induce oxidative stress.

Other factors may also initiate the production of reactive oxygen species including dopamine, elevated iron, neuromelanin, nitric oxide, depleted glutathione, endo/exotoxin, genetic predisposition, and mitochondrial dysfunction. Eventually, oxidative stress, accumulation of protein aggregates, and other cytotoxic processes lead to the destruction of cells. Key: ROS reactive oxygen species; JAK/STAT Janus kinase and signal transducer of activation; MAPK mitogen-activated protein kinase; NF- $\kappa$ B nuclear factor kappa B; IL interleukin; DA dopamine; NM neuromelanin; NO nitric oxide; GSH reduced glutathione

can thus contribute to the cell carnage associated with neurodegeneration. Neurons appear to be more susceptible to the cytotoxic effects of NO in contrast to astrocytes (Tewari et al. 2020). This is probably due to the presence of a higher content of cellular antioxidant, glutathione in the astrocytes compared to that present in the neurons (Chen et al 2001). This concept is endorsed by the glutathione-dependent protection offered by the astrocytes against oxidative stress.

Neuroinflammation in the brain can lead to excessive production of NO by glia, which can in turn exacerbate cellular deleterious effects associated with chronic neuro-inflammation. Thus, it therefore unequivocally qualifies as a Janus-faced molecule. Indeed, NO can effectively also exert a sinister role in both neurodegeneration and neuroinflammation

and is therefore implicated in PD, AD, and other degenerative disorders (Tewari et al. 2020).

Stykel and associates (2022), suggested that about 30% of the proteins present in LB are nitrosylated by reactive nitrogen species. The nitration of  $\alpha$ -synuclein by NO may also contribute to its misfolding and aggregation in PD. More importantly, this reflects the involvement of NO and reactive nitrogen species in the degenerative processes occurring in LB-containing diseases or other  $\alpha$ -synucleinopathies.

As discussed in depth,  $\alpha$ -synuclein also exhibits Janus-faced qualities as demonstrated by its physiological function and pathological role involved in LB formation (Ray et al. 2021).



Interestingly, aggregates of amyloid  $\beta$  proteins are considered to have very harmful properties and exert neurotoxic effects in a myriad of ways in AD as analyzed earlier. However, it has two Janus faces too, since the core hexapeptide structure which is present in many amyloids including A $\beta$  and tau has particularly notable immune suppressant qualities (Steinman et al 2014). Steinman and co-workers (2014) also reported that administration of this hexapeptide (derived from  $\beta$ -amyloid) rapidly decreased levels of pro-inflammatory cytokines (IL-2, IL-6). It would therefore be beneficial in the management of AD to determine the conditions warranted to activate this component of  $\beta$ amyloid, perhaps to halt or reduce the onslaught of degeneration in AD.

The divergent cellular effects of microglia M1 and M2 clearly exhibit its Janus-faced characteristics. M1 induces inflammatory neurotoxicity, in contrast, M2 exerts anti-inflammatory and neuroprotective actions (Colonna and Butovsky 2017).

It would appear that by virtue of evoking an inflammatory response, viruses may be indirectly involved in the activation of the sinister and cytotoxic component of some of these Janus-faced molecules.

## Conclusion

Neurodegenerative disorders including PD and AD are associated with neuroinflammatory cascades, which may be triggered by eg viral infections. Reviewing these cascades, like ROS/oxidative stress related to dopamine, iron, nitric oxide, loss of glutathione, mitochondrial dysfunction, excitotoxicity, cytotoxic cytokines, microglial M1/M2 activities, JAK-STAT pathway, the role of Toll-receptors, NF $\kappa$ B we aim to elucidate the viral impact leading to accumulation/aggregation of proteins characteristic for PD and AD,  $\alpha$ -synuclein (PD), and  $\beta$ -amyloid respective Tau-protein (AD).

Multiple cytotoxic neuroinflammatory processes leading to protein dyshomeostasis and finally neuronal destruction may depend on “dose” and duration of such pathologic processes induced/triggered by a variety of endotoxins including viruses. As such, involvement of the blood–brain barrier is of crucial importance. The BBB may be intact in the case of neuroinflammation but defect in the case of inflammation. This should be elucidated in future experimental studies and clinical as well as human post mortem brain studies.

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## References

- Aguzzi A, Barres BA, Bennett ML (2013) Microglia: scapegoat, saboteur, or something else? *Science* 339(6116):156–161. <https://doi.org/10.1126/science.1227901>. PMID:23307732;PMCID:PMC4431634
- Akilli NB, Yosunkaya A (2021) Part of the Covid19 puzzle: Acute parkinsonism. *Am J Emerg Med*. <https://doi.org/10.1016/j.ajem.2021.02.050>
- Albornoz EA, Amarilla AA, Modhiran N, Parker S, Li XX, Wijesundara DK, Aguado J, Zamora AP, McMillan CLD, Liang B, Peng NYG, Sng JDJ, Saima FT, Fung JN, Lee JD, Paramitha D, Parry R, Avumegah MS, Isaacs A, Lo MW, Miranda-Chacon Z, Bradshaw D, Salinas-Rebolledo C, Rajapakse NW, Wolvetang EJ, Munro TP, Rojas-Fernandez A, Young PR, Stacey KJ, Khromykh AA, Chappell KJ, Watterson D, Woodruff TM (2022) SARS-CoV-2 drives NLRP3 inflammasome activation in human microglia through spike protein. *Mol Psychiatry*. <https://doi.org/10.1038/s41380-022-01831-0>
- Al-Samhari MM, Al-Rasheed NM, Al-Rejaie S, Al-Rasheed NM, Hasan IH, Mahmoud AM, Dzimir N (2015) Possible involvement of the JAK/STAT signaling pathway in N-acetylcysteine-mediated antidepressant-like effects. *Exp Biol Med* 241(5):509–518. <https://doi.org/10.1177/1535370215619707>
- Alzheimer A, Stelzmann RA, Schnitzlein HN, Murtagh FR (1995) An english translation of Alzheimer’s 1907 Paper, ‘Über Eine Eigenartige Erkrankung Der Hirnrinde.’ *Clinical Anatomy* (new York, NY) 8(6):429–431. <https://doi.org/10.1002/ca.980080612>
- Alzheimer A, (1907) Über eine eigenartige Erkrankung der Hirnrinde. *Allgemeine Zeitschrift für Psychiatrie und Psychisch-gerichtliche Medizin*. 64:146-8
- Amarante-Mendes GP, Adjemian S, Migliari Branco L, Zanetti LC, Weinlich R, Bortoluci KR (2018) Pattern recognition receptors and the host cell death molecular machinery. *Frontiers Immunology*. <https://doi.org/10.3389/fimmu.2018.02379>
- Andronie-Cioara FL, Ardelean AI, Nistor-Cseppento CD, Jurcau A, Jurcau MC, Pascalau N, Marcu F (2023) Molecular Mechanisms of Neuroinflammation in Aging and Alzheimer’s Disease Progression. *Int J Molecular Sci*. 24(3):1869. <https://doi.org/10.3390/ijms24031869>
- Ariga H, Takahashi-Niki K, Kato I, Maita H, Niki T, Iguchi-Ariga SMM (2013) Neuroprotective function of DJ-1 in parkinson’s disease. *Oxid Med Cell Longev* 2013:1–9. <https://doi.org/10.1155/2013/683920>
- Ayyubova G (2022) dysfunctional microglia and tau pathology in Alzheimer’s disease. *Rev Neurosci* 34(4):443–458. <https://doi.org/10.1515/revneuro-2022-0087>. (PMID: 36302379)
- Baik SH, Kang S, Lee W, Choi H, Chung S, Kim JI, Mook-Jung I (2019) A breakdown in metabolic reprogramming causes microglia dysfunction in Alzheimer’s disease. *Cell Metab*

- 30(3):493–507. <https://doi.org/10.1016/j.cmet.2019.06.005>. (Epub 2019 Jun 27 PMID: 31257151)
- Baizabal-Carvalho JF, Alonso-Juarez M (2021) The role of viruses in the pathogenesis of parkinson's disease. *Neural Regen Res* 16(6):1200. <https://doi.org/10.4103/1673-5374.300437>
- Bamberger ME, Harris ME, McDonald DR, Husemann J, Landreth GE (2003) A cell surface receptor complex for fibrillar  $\beta$ -amyloid mediates microglial activation. *J Neurosci* 23(7):2665–2674. <https://doi.org/10.1523/jneurosci.23-07-02665.2003>
- Bantle CM, Phillips AT, Smeyne RJ, Rocha SM, Olson KE, Tjalkens RB (2019) Infection with mosquito-borne alphavirus induces selective loss of dopaminergic neurons, neuroinflammation and widespread protein aggregation. *Npj Parkinson's Disease*. <https://doi.org/10.1038/s41531-019-0090-8>
- Barrett ADT, Weaver SC, (2012). Arboviruses: Alphaviruses, flaviviruses and bunyaviruses: Encephalitis; yellow fever; dengue; haemorrhagic fever; miscellaneous tropical fevers; undifferentiated fever. In *Medical Microbiology: Eighteenth Edition* (pp. 520–536). Elsevier Inc. <https://doi.org/10.1016/B978-0-7020-4089-4.00066-4>
- Beatman E, Massey A, Shives KD, Burrack KS, Chamanian M, Morrison TE, Beckham JD (2016) "Alpha-synuclein expression restricts RNA viral infections in the brain" edited by S. Perlman. *J Virology* 90(6):2767–2782. <https://doi.org/10.1128/jvi.02949-15>
- Benedetti F (2023) Immunopsychiatry after COVID-19. *J Affect Disord Rep*. <https://doi.org/10.1016/j.jadr.2023.100521>. (Epub 2023 Mar 31. PMID: PMC10063369)
- Ben-Zvi A, Lacoste B, Kur E, Andreone BJ, Mayshar Y, Yan H, Gu C (2014) Mfsd2a is critical for the formation and function of the blood-brain barrier. *Nature* 509(7501):507–511. <https://doi.org/10.1038/nature13324>. (PMID: 24828040; PMID: PMC4134871)
- Berger Z, Roder H, Hanna A, Rangachari V, Yue M, Wszolek Z, Ashe K, Knight J, Dickson D, Andorfer C, Rosenberry TL, Lewis J, Hutton M, Janus C, (2007). Accumulation of Pathological Tau Species and Memory Loss in a Conditional Model of Tauopathy. *Journal of Neuroscience*, 27 (14) 3650–3662. Available at: [https://aquila.usm.edu/fac\\_pubs/7541](https://aquila.usm.edu/fac_pubs/7541)
- Bido S, Muggeo S, Massimino L, Marzi MJ, Giannelli SG, Melacini E, Nannoni M et al (2021) Author correction: microglia-specific overexpression of  $\alpha$ -synuclein leads to severe dopaminergic neurodegeneration by phagocytic exhaustion and oxidative toxicity. *Nat Commun*. <https://doi.org/10.1038/s41467-021-27737-z>
- Blevins HM, Xu Y, Biby S, Zhang S (2022) The NLRP3 inflammasome pathway: a review of mechanisms and inhibitors for the treatment of inflammatory diseases. *Frontiers Aging Neurosci*. <https://doi.org/10.3389/fnagi.2022.879021>
- Blum-Degen D, Muller T, Kuhn W, Gerlach M, Przuntek H, Riederer P (1995) Interleukin-1 $\beta$  and interleukin-6 are elevated in the cerebrospinal fluid of Alzheimer's and de novo Parkinson's disease patients. *Neurosci Lett* 202:17–20
- Boengler K, Hilfikerkleiner D, Drexler H, Heusch G, Schulz R (2008) The myocardial JAK/STAT pathway: From protection to failure. *Pharmacol Ther* 120(2):172–185. <https://doi.org/10.1016/j.pharmthera.2008.08.002>
- Bourgognon JM, Spiers JG, Robinson SW, Scheiblich H, Glynn P, Ortori C, Bradley SJ, Tobin AB, Steinert JR (2021) Inhibition of neuroinflammatory nitric oxide signaling suppresses glycation and prevents neuronal dysfunction in mouse prion disease. *Proc Natl Acad Sci USA* 118(10):e2009579118. <https://doi.org/10.1073/pnas.2009579118>
- Braak H, Del Tredici K, Rüb U, de Vos RA, Jansen Steur EN, Braak E (2003) Staging of brain pathology related to sporadic Parkinson's disease. *Neurobiol Aging* 24(2):197–211. [https://doi.org/10.1016/s0197-4580\(02\)00065-9](https://doi.org/10.1016/s0197-4580(02)00065-9). (PMID: 12498954)
- Bukhbinder AS, Ling Y, Hasan O, Jiang X, Kim Y, Phelps KN, Schmandt RE et al (2022) Risk of alzheimer's disease following influenza vaccination: a claims-based cohort study using propensity score matching. *J Alzheimer's Disease: JAD* 88(3):1061–1074. <https://doi.org/10.3233/JAD-220361>
- Cadar D, Jellinger KA, Riederer P, Strobel S, Monoranu CM, Tappe D (2021) No Metagenomic evidence of causative viral pathogens in Postencephalitic Parkinsonism Following Encephalitis Lethargica. *Microorganisms* 9(8):1716. <https://doi.org/10.3390/microorganisms9081716>. PMID:34442795;PMCID:PMC8398509
- Cai Y, Liu J, Wang B, Sun M, Yang H (2022) Microglia in the neuro-inflammatory pathogenesis of Alzheimer's disease and related therapeutic targets. *Front Immunol*. <https://doi.org/10.3389/fimmu.2022.856376>. PMID:35558075;PMCID:PMC9086828
- Cairns DM, Itzhaki RF, Kaplan DL (2022) Potential involvement of varicella zoster virus in alzheimer's disease via reactivation of quiescent herpes simplex virus type 1. *J Alzheimer's Disease* 88(3):1189–1200. <https://doi.org/10.3233/jad-220287>
- Calabrese V, Cornelius C, Rizzarelli E, Owen JB, Dinkova-Kostova AT, Butterfield DA (2009) Nitric oxide in cell survival: a janus molecule. *Antioxid Redox Signal* 11(11):2717–2739. <https://doi.org/10.1089/ars.2009.2721>
- Cannon T, Gruenheid S (2022) Microbes and Parkinson's disease: from associations to mechanisms. *Trends Microbiol* 30(8):749–760. <https://doi.org/10.1016/j.tim.2022.01.004>
- Cavallieri F, Fioravanti V, Toschi G, Grisanti S, Napoli M, Moratti C, Pascarella R, Versari A, Fraternali A, Casali M, Paul JJ, Moro E, Bauer P, Zedde M, Valzania F (2022) COVID-19 and parkinson's disease: a casual association or a possible second hit in neurodegeneration? *J Neurol* 269(1):59–61. <https://doi.org/10.1007/s00415-021-10694-4>
- Chen Y, Vartiainen NE, Ying W, Chan PH, Koistinaho J, Swanson RA (2001) Astrocytes protect neurons from nitric oxide toxicity by a glutathione-dependent mechanism. *J Neurochem* 77(6):1601–1610. <https://doi.org/10.1046/j.1471-4159.2001.00374.x>
- Chen G, Xu T, Yan Y, Zhou Y, Jiang Y, Melcher K, Xu HE (2017) Amyloid beta: structure, biology and structure-based therapeutic development. *Acta Pharmacol Sin* 38(9):1205–1235. <https://doi.org/10.1038/aps.2017.28>
- Chen L, Deng H, Cui H, Fang J, Zuo Z, Deng J, Li Y, Wang X, Zhao L (2018) Inflammatory responses and inflammation-associated diseases in organs. *Oncotarget* 9(6):7204–7218. <https://doi.org/10.18632/oncotarget.23208>
- Chew H, Solomon VA, Fonteh AN (2020) Involvement of lipids in Alzheimer's disease pathology and potential therapies. *Frontiers Physiology*. <https://doi.org/10.3389/fphys.2020.00598>
- Choi S-H, Aid S, Kim H-W, Jackson SH, Bosetti F (2011) Inhibition of NADPH oxidase promotes alternative and anti-inflammatory microglial activation during neuroinflammation. *J Neurochem* 120(2):292–301. <https://doi.org/10.1111/j.1471-4159.2011.07572.x>
- Choi I, Zhang Y, Seegobin SP, Pruvost M, Wang Q, Purtell K, Zhang B, Yue Z (2020) Microglia clear neuron-released  $\alpha$ -synuclein via selective autophagy and prevent neurodegeneration. *Nat Commun* 11(1):1386. <https://doi.org/10.1038/s41467-020-15119-w>. PMID:32170061;PMCID:PMC7069981
- Chung HY, Cesari M, Anton S, Marzetti E, Giovannini S, Seo AY, Carter C, Yu BP, Leeuwenburgh C (2009) Molecular inflammation: underpinnings of aging and age-related diseases. *Ageing Res Rev* 8(1):18–30. <https://doi.org/10.1016/j.arr.2008.07.002>
- Ciesielska A, Matyjek M, Kwiatkowska K (2020) TLR4 and CD14 trafficking and its influence on LPS-induced pro-inflammatory signaling. *Cellular Molecular Life Sci* 78(4):1233–1261. <https://doi.org/10.1007/s00018-020-03656-y>
- Coates BM, Staricha KL, Ravindran N, Koch CM, Cheng Y, Davis JM, Shumaker DK (2017) Inhibition of the NOD-Like receptor

- protein 3 Inflammasome Is protective in juvenile influenza a virus infection. *Front Immunol* 10(8):782. <https://doi.org/10.3389/fimmu.2017.00782>. PMID:28740490;PMCID:PMC5502347
- Cocoros NM, Svensson E, Szépligeti SK et al (2021) Long-term risk of Parkinson disease following influenza and other infections. *Jama Neurol* 78(12):1461–1470. <https://doi.org/10.1001/jamanneurol.2021.3895>
- Colonna M, Butovsky O (2017) Microglia function in the central nervous system during health and neurodegeneration. *Annu Rev Immunol* 35(1):441–468. <https://doi.org/10.1146/annurev-immunol-051116-052358>
- Combs CK, Karlo JC, Kao S-C, Landreth GE (2001)  $\beta$ -amyloid stimulation of microglia and monocytes results in TNF $\alpha$ -dependent expression of inducible nitric oxide synthase and neuronal apoptosis. *J Neurosci* 21(4):1179–1188. <https://doi.org/10.1523/jneurosci.21-04-01179.2001>
- Conte M, Feltracco M, Chirizzi D, Trabucco S, Dinoi A, Gregoris E, Barbaro E, La Bella G, Ciccarese G, Belosi F, La Salandra G, Gambaro A, Contini D (2021) Airborne concentrations of SARS-CoV-2 in indoor community environments in Italy. *Environ Sci Pollut Res*. <https://doi.org/10.1007/s11356-021-16737-7>
- Corder EH, Saunders AM, Strittmatter WJ, Schmechel DE, Gaskell PC, Small GW, Roses AD, Haines JL, Pericak-Vance MA (1993) Gene dose of apolipoprotein E type 4 allele and the risk of Alzheimer's disease in late onset families. *Science* 261(5123):921–923. <https://doi.org/10.1126/science.8346443>. (PMID: 8346443)
- Crunfli F, Carregari VC, Veras FP, Silva LS, Nogueira MH (2022) Morphological, cellular, and molecular basis of brain infection in COVID-19 patients. *Proc Natl Acad Sci U S A* 119(35):e2200960119. <https://doi.org/10.1073/pnas.2200960119>. PMID:35951647;PMCID:PMC9436354
- Curren EJ, Hills SL, Fischer M, Lindsey NP (2018) St. louis encephalitis virus disease in the united states, 2003–2017. *Am J Trop Med Hyg* 99(4):1074–1079. <https://doi.org/10.4269/ajtmh.18-0420>
- Czub M, Czub S, Gosztonyi G, Koutsilieri E, Sopper S, Müller JG, Gerlach M, Riederer P (1999) Effects of Selegiline in a retroviral rat model for neurodegenerative disease. *J Neurovirol* 5(5):458–464. <https://doi.org/10.3109/13550289909045374>. (PMID: 10568882)
- Czub S, Koutsilieri E, Sopper S, Czub M, Stahl-Hennig C, Müller JG, Pedersen V, Gsell W, Heeney JL, Gerlach M, Gosztonyi G, Riederer P (2001) Enhancement of central nervous system pathology in early simian immunodeficiency virus infection by dopaminergic drugs. *Acta Neuropathol* 101(2):85–91. <https://doi.org/10.1007/s004010000313>. (PMID: 11271377)
- da Silva Torres MK, Bichara CDA, de Almeida MNDS, Vallinoto MC, Queiroz MAF, Vallinoto IMVC, Dos Santos EJM, de Carvalho CAM (2022) The complexity of SARS-CoV-2 infection and the COVID-19 pandemic. *Front Microbiol* 10(13):789882. <https://doi.org/10.3389/fmicb.2022.789882>. PMID:35222327;PMCID:PMC8870622
- Daneman R, Prat A (2015) The Blood-Brain Barrier. *Cold Spring Harb Perspect Biol* 7(1):a020412. <https://doi.org/10.1101/cshperspect.a020412>
- Davis LE, Koster F (2014) Neurologic aspects of influenza viruses. *Handb Clin Neurol* 123:619–645. <https://doi.org/10.1016/B978-0-444-53488-0.00030-4>
- Davis HE, McCorkell L, Vogel JM et al (2023) (2023) Long COVID: major findings, mechanisms and recommendations. *Nat Rev Microbiol* 21:133–146. <https://doi.org/10.1038/s41579-022-00846-2>
- Deczkowska A (2018) Disease-associated microglia: a universal immune sensor of Neurodegeneration. *Cell* 173(5):1073–1081. <https://doi.org/10.1016/j.cell.2018.05.003>
- DiSabato DJ, Quan N, Godbout JP (2016) Neuroinflammation: the devil is in the details. *J Neurochem*. <https://doi.org/10.1111/jnc.13607>
- Drouin-Ouellet J, St-Amour I, Saint-Pierre M, Lamontagne-Proulx J, Kriz J, Barker RA, Cicchetti F (2014) Toll-like receptor expression in the blood and brain of patients and a mouse model of Parkinson's disease. *Int J Neuropsychopharmacol*. <https://doi.org/10.1093/ijnp/pyu103>. PMID:25522431;PMCID:PMC4438545
- Du RH, Sun HB, Hu ZL, Lu M, Ding JH, Hu G (2018) Kir6.1/K-ATP channel modulates microglia phenotypes: implication in Parkinson's disease. *Cell Death Dis* 9(3):404. <https://doi.org/10.1038/s41419-018-0437-9>
- Dzambo N, Gysbers A, Perera G, Bahar A, Shankar A, Gao J, Fu Y, Halliday GM (2016) Toll-like receptor 2 is increased in neurons in Parkinson's disease brain and may contribute to alpha-synuclein pathology. *Acta Neuropathol* 133(2):303–319. <https://doi.org/10.1007/s00401-016-1648-8>
- Efstathiou V, Stefanou MI, Demetriou M, Siafakas N, Makris M, Tsigvoulis G, Zoumpourlis V, Kypouropoulos SP, Tsoporis JN, Spandidos DA, Smyrnis N, Rizos E (2022) Long COVID and neuropsychiatric manifestations (Review). *Exp Ther Med* 23(5):363. <https://doi.org/10.3892/etm.2022.11290>
- Eikelenboom P, Zhan SS, van Gool WA (1994) Inflammatory mechanisms in Alzheimer's disease. *Trends Pharmacol Sci* 15(12):447–450. [https://doi.org/10.1016/0165-6147\(94\)90057-4](https://doi.org/10.1016/0165-6147(94)90057-4). (PMID: 7886816)
- Engler-Chiurazzi EB, Russell AE, Povroznik JM, McDonald KO, Porter KN, Wang DS, Hammock J, Billig BK, Felton CC, Yilmaz A, Schreurs BG, O'Callaghan JD, Zvezdaryk KJ, Simpkins JW (2023) Intermittent systemic exposure to lipopolysaccharide-induced inflammation disrupts hippocampal long-term potentiation and impairs cognition in aging male mice. *Brain Behav Immun* 108:279–291. <https://doi.org/10.1016/j.bbi.2022.12.013>. (Epub 2022 Dec 19. PMID: 36549577; PMCID: PMC10019559)
- Eugenin EA (2006) CCL2/Monocyte chemoattractant protein-1 mediates enhanced transmigration of Human immunodeficiency virus (HIV)-infected Leukocytes across the Blood-brain barrier: a potential mechanism of HIV-CNS invasion and NeuroAIDS. *J Neurosci* 26(4):1098–1106. <https://doi.org/10.1523/jneurosci.3863-05.2006>
- Fajgenbaum DC (2020) Cytokine storm. *N Engl J Med* 383(23):2255–2273. <https://doi.org/10.1056/NEJMr2026131>. PMID:33264547; PMCID:PMC7727315
- Ferreira R, Santos T, Cortes L, Cochaud S, Agasse F, Silva AP, Xapelli S, Malva JO (2011) Neuropeptide Y inhibits interleukin-1 beta-induced microglia motility. *J Neurochem* 120(1):93–105. <https://doi.org/10.1111/j.1471-4159.2011.07541.x>
- Fields JK, Günther S, Sundberg EJ (2019) Structural Basis of IL-1 family cytokine signaling. *Frontiers Immunology*. <https://doi.org/10.3389/fimmu.2019.01412>
- Foley PB, (2009) Encephalitis lethargica and influenza. I. The role of the influenza virus in the influenza pandemic of 1918/1919. *J Neural Transm (Vienna)*. 116, (2): 143–50. <https://doi.org/10.1007/s00702-008-0161-1>
- Fu AKY, Hung K-W, Yuen MYF, Zhou X, Mak DSY, Chan ICW, Cheung TH, Zhang B, Fu W-Y, Liew FY, Ip NY (2016) IL-33 ameliorates Alzheimer's disease-like pathology and cognitive decline. *Proceed Nat Academy Sci*. <https://doi.org/10.1073/pnas.1604032113>
- Gamboa ET, Yahr MD (1973) Influenza a virus as a possible cause of postencephalitic parkinsonism. *Trans Am Neurol Assoc* 98:177–180
- Ghoreschi K, Laurence A, O'Shea JJ (2009) Janus kinases in immune cell signaling. *Immunol Rev* 228(1):273–287. <https://doi.org/10.1111/j.1600-065x.2008.00754.x>

- Gibb WRG, Scott T, Lees AJ (1991) Neuronal inclusions of parkinson's disease. *Mov Disord* 6(1):2–11. <https://doi.org/10.1002/mds.870060103>
- Glass JD, Johnson RT (1996) Human immunodeficiency virus and the brain. *Annu Rev Neurosci* 19(1):1–26
- Goerttler T, Kwon E-H, Fleischer M, Stettner M, Tönges L, Klebe S (2022) SARS-CoV-2, COVID-19 and parkinson's disease—many issues need to be clarified—a critical review. *Brain Sci* 12(4):456. <https://doi.org/10.3390/brainsci12040456>
- Griffin DE (2014) Measles virus and the nervous system. *Handb Clin Neurol* 123:577–590. <https://doi.org/10.1016/B978-0-444-53488-0.00027-4>. (PMID: 25015505)
- Gu Y, Zuo X, Zhang S, Ouyang Z, Jiang S, Wang F (2021) The mechanism behind influenza virus cytokine storm. *Viruses* 13(7):1362. <https://doi.org/10.3390/v13071362>. PMID: 34372568; PMCID: PMC8310017
- Guo XJ, Thomas PG (2017) New Fronts Emerge in the influenza cytokine storm. *Seminars Immunopathology* 39(5):541–550. <https://doi.org/10.1007/s00281-017-0636-y>
- Hansen DV, Hanson JE, Sheng M (2017) Microglia in alzheimer's disease. *J Cell Biol* 217(2):459–472. <https://doi.org/10.1083/jcb.201709069>
- Hara H, Chida J, Uchiyama K, Pasiana AD, Takahashi E, Kido H (2021) Neurotropic influenza a virus infection causes prion protein misfolding into infectious prions in neuroblastoma cells. *Sci Rep* 11(1):10109. <https://doi.org/10.1038/s41598-021-89586-6>. PMID: 33980968; PMCID: PMC8115602
- Harrison DA (2012) The JAK/STAT pathway. *Cold Spring Harb Perspect Biol*. <https://doi.org/10.1101/cshperspect.a011205>
- Harrison KM, Kajese T, Hall HI, Song R (2008) Risk factor redistribution of the national HIV/AIDS surveillance data: an alternative approach. *Public Health Rep* 123(5):618–627. <https://doi.org/10.1177/003335490812300512>
- Harrison AG, Lin T, Wang P (2020) Mechanisms of SARS-CoV-2 transmission and pathogenesis. *Trends Immunol* 41(12):1100–1115. <https://doi.org/10.1016/j.it.2020.10.004>
- Hawkes CH, Del Tredici K, Braak H (2007) Parkinson's disease: a dual-hit hypothesis. *Neuropathol Appl Neurobiol* 33(6):599–614. <https://doi.org/10.1111/j.1365-2990.2007.00874.x>
- Heidari A, Yazdanpanah N, Rezaei N (2022) The role of toll-like receptors and neuroinflammation in parkinson's Disease. *J Neuroinflammation*. <https://doi.org/10.1186/s12974-022-02496-w>
- Hendrayani S-F, Al-Harbi B, Al-Ansari MM, Silva G, Abousekhra A (2016) The inflammatory/cancer-related IL-6/STAT3/NF- $\kappa$ B positive feedback loop includes AUF1 and maintains the active state of breast myofibroblasts. *Oncotarget* 7(27):41974–41985. <https://doi.org/10.18632/oncotarget.9633>
- Hickman S, Izzy S, Sen P, Morsett L, Khoury JL (2018) Microglia in Neurodegeneration. *Nat Neurosci* 21(10):1359–1369. <https://doi.org/10.1038/s41593-018-0242-x>
- Hill JM, Lukiw WJ (2022) microRNA, the innate-immune system and SARS-CoV-2. *Front Cell Infect Microbiol* 12:887800. <https://doi.org/10.3389/fcimb.2022.887800>
- Hirsch E, Graybiel AM (1988) Melanized dopaminergic neurons are differentially susceptible to degeneration in Parkinson's disease. *Nature* 334(6180):345–348. <https://doi.org/10.1038/334345a0>. (PMID: 2899295)
- Hirsch EC, Standaert DG (2021) Ten unsolved questions about neuroinflammation in parkinson's disease. *Mov Disord* 36(1):16–24. <https://doi.org/10.1002/mds.28075>. (Epub 2020 May 1 PMID: 32357266)
- Hirsch EC, Breidert T, Rousset E, Hunot S, Hartmann A (2003) The role of glial reaction and inflammation in Parkinson's disease. *Ann N Y Acad Sci* 991:214–228. <https://doi.org/10.1111/j.1749-6632.2003.tb07478.x>. (PMID: 12846989)
- Hoffman LA (2017) Encephalitis lethargica: 100 years after the epidemic. *Brain* 140(8):2246–2251. <https://doi.org/10.1093/brain/awx177>
- Hopkins HK, Traverse EM, Barr KL (2022) Viral Parkinsonism: an underdiagnosed neurological complication of Dengue virus infection. *PLoS Negl Trop Dis* 16(2):e0010118. <https://doi.org/10.1371/journal.pntd.0010118>
- Hunot S, Hirsch EC (2003) Neuroinflammatory processes in Parkinson's disease. *Ann Neurol*. <https://doi.org/10.1002/ana.10481>
- Husain MA, Laurent B, Plourde M (2021) APOE and Alzheimer's disease: from lipid transport to physiopathology and therapeutics. *Front Neurosci*. <https://doi.org/10.3389/fnins.2021.630502>
- Huynh VA, Takala TM, Murros KE, Diwedi B (2023) Saris PEJ (2023) *Desulfovibrio* bacteria enhance alpha-synuclein aggregation in a *Caenorhabditis elegans* model of Parkinson's disease. *Front Cell Infect Microbiol* 1(13):1181315. <https://doi.org/10.3389/fcimb.2023.1181315>. PMID: 37197200; PMCID: PMC10183572
- Imfeld P, Toovey S, Jick SS (2016) Meier CR (2016) Influenza infections and risk of Alzheimer's disease. *Brain Behav Immun* 57:187–192. <https://doi.org/10.1016/j.bbi.2016.03.014>. (Epub 2016 Mar 19 PMID: 27006278)
- Ismail R, Parbo P, Madsen LS, Hansen AK, Hansen KV, Schaldemose JL, Kjeldsen PL, Stokholm MG, Gottrup H, Eskildsen SF (2020) The relationships between neuroinflammation, beta-amyloid and tau deposition in Alzheimer's disease: a longitudinal PET study. *J Neuroinflammation* 17(1):151. <https://doi.org/10.1186/s12974-020-01820-6>. PMID: 32375809; PMCID: PMC7203856
- Itoh K, Mehraein P, Weis S (2000) Neuronal damage of the substantia nigra in HIV-1 infected brains. *Acta Neuropathol* 99:376–384
- Itzhaki RF (2016) Herpes and Alzheimer's disease: subversion in the central nervous system and how it might be halted. *J Alzheimer's Disease* : JAD 54(4):1273–1281. <https://doi.org/10.3233/JAD-160607>
- Itzhaki RF (2017) Herpes simplex virus type 1 and Alzheimer's disease: possible mechanisms and signposts. *FASEB J* 31(8):3216–3226. <https://doi.org/10.1096/fj.201700360>
- Iwasaki AA (2012) (2012) virological view of innate immune recognition. *Annu Rev Microbiol* 66:177–196. <https://doi.org/10.1146/annurev-micro-092611-150203>. PMID: 22994491; PMCID: PMC3549330
- Jackson AC (2014) Rabies. *Neurovirology*. <https://doi.org/10.1016/b978-0-444-53488-0.00029-8>
- Jamieson GA, Maitland NJ, Wilcock GK, Craske J, Itzhaki RF (1991) Latent Herpes simplex virus type 1 in normal and alzheimer's disease brains. *J Med Virol* 33(4):224–227. <https://doi.org/10.1002/jmv.1890330403>
- Jana M, Palencia CA, Pahan K (2008) Fibrillar amyloid-beta peptides activate microglia via TLR2: implications for Alzheimer's disease. *J Immunol* 181(10):7254–7262. <https://doi.org/10.4049/jimmunol.181.10.7254>
- Jang H, Boltz DA, Webster RG, Smeyne RJ (2009a) Viral parkinsonism. *Biochem Biophys Acta* 1792(7):714–721. <https://doi.org/10.1016/j.bbdis.2008.08.001>
- Jang H, Boltz D, Sturm-Ramirez K, Shepherd KR, Jiao Y, Webster R, Smeyne RJ (2009b) Highly pathogenic H5N1 influenza virus can enter the central nervous system and induce neuroinflammation and neurodegeneration. *Proc Natl Acad Sci U S A* 106(33):14063–14068. <https://doi.org/10.1073/pnas.0900096106>
- Jang H, Boltz D, McClaren J, Pani AK, Smeyne M, Korff A, Webster R (2012) Inflammatory effects of highly pathogenic H5N1 influenza virus infection in the CNS of mice. *J Neurosci* 32(5):1545–1559. <https://doi.org/10.1523/JNEUROSCI.5123-11.2012>. PMID: 22302798; PMCID: PMC3307392

- Jankovic J (2020) Parkinson's disease: etiopathogenesis and treatment journal of neurology. *Neurosurgery Psychiatry* 91:795–808
- Janssen RS, Nwyanwu OC, Selik RM, Stehr-Green JK (1992) Epidemiology of human immunodeficiency virus encephalopathy in the united states. *Neurology* 42:1472–1476
- Jatiani SS, Baker SJ, Silverman LR, Reddy EP (2010) JAK/STAT pathways in cytokine signaling and myeloproliferative disorders: approaches for targeted therapies. *Genes Cancer* 1(10):979–993. <https://doi.org/10.1177/1947601910397187>
- Jellinger KA (2009) Absence of alpha-synuclein pathology in postencephalitic parkinsonism. *Acta Neuropathol* 118(3):371–379. <https://doi.org/10.1007/s00401-009-0537-9>
- Jiang Y, Chen Y, Sun H, Zhang X, He L, Li J, Zhao G (2021) MERS-CoV infection causes brain damage in human DPP4-transgenic mice through complement-mediated inflammation. *J Gen Virol* 102(10):001667. <https://doi.org/10.1099/jgv.0.001667>. PMID: 34704923; PMCID: PMC8604193
- Joers V, Tansey MG, Mulas G, Carta AR (2017) Microglial phenotypes in parkinson's disease and animal models of the disease. *Prog Neurobiol* 155:57–75. <https://doi.org/10.1016/j.pneurobio.2016.04.006>
- Kaminska, B. (2005). MAPK signalling pathways as molecular targets for anti-inflammatory therapy—from molecular mechanisms to therapeutic benefits. *Biochimica et Biophysica Acta (BBA) - Proteins and Proteomics*, 1754 (1–2) 253–262. <https://doi.org/10.1016/j.bbapap.2005.08.017>
- Kawai T, Akira S (2007) Signaling to NF- $\kappa$ B by Toll-like receptors. *Trends Molecular Med* 13(11):460–469. <https://doi.org/10.1016/j.molmed.2007.09.002>
- Keren-Shaul H, Spinrad A, Weiner A, Matcovitch-Natan O, Dvir-Szternfeld R, Ulland TK, David E, Baruch K, Lara-Astaiso D, Toth B, Itzkovitz S, Colonna M, Schwartz M (2017) A unique microglia type associated with restricting development of Alzheimer's disease. *Cell* 169(7):1276–1290.e17. <https://doi.org/10.1016/j.cell.2017.05.018>. (Epub 2017 Jun 8 PMID: 28602351)
- Kim WG, Mohny RP, Wilson B, Jeohn GH, Liu B (2000) Regional difference in susceptibility to lipopolysaccharide-induced neurotoxicity in the rat brain: role of microglia. *J Neurosci* 20(16):6309–6316. <https://doi.org/10.1523/JNEUROSCI.20-16-06309.2000>. PMID: 10934283; PMCID: PMC6772569
- Klein RS, Garber C, Howard N (2017) Infectious immunity in the central nervous system and brain function. *Nat Immunol* 18(2):132–141. <https://doi.org/10.1038/ni.3656>
- Koutsilieri E, Ter Meulen V, Riederer P (2001) Neurotransmission in HIV associated dementia: a short review. *J Neural Transm* 108:767–775. <https://doi.org/10.1007/s007020170051>
- Koutsilieri E, Scheller C, Czub S, Sopper S, ter Meulen V, Riederer P (2002a) Dopamine Is a Pathogenetic Factor in HIV-Induced Neuro-AIDS. In: Nagatsu T, Nabeshima T, McCarty R, Goldstein DS (eds) Catecholamine Research. *Advances in Behavioral Biology*, Springer, Boston, MA
- Koutsilieri E, Sopper S, Scheller C, ter Meulen V (2002b) Parkinsonism in HIV dementia. *J Neural Transm* 109(5–6):767–775. <https://doi.org/10.1007/s007020200063>
- Labandeira-Garcia JL, Costa-Besada MA, Labandeira CM, Villar-Cheda B, Rodríguez-Perez AI (2017) Insulin-Like growth factor-1 and Neuroinflammation. *Frontiers Aging Neurosci*. <https://doi.org/10.3389/fnagi.2017.00365>
- Lan X, Han X, Li Q, Yang Q-W, Wang J (2017) Modulators of microglial activation and polarization after intracerebral haemorrhage. *Nat Rev Neurol* 13(7):420–433. <https://doi.org/10.1038/nrneurol.2017.69>
- Leblanc P, Vorberg IM (2022) Viruses in neurodegenerative diseases: more than just suspects in crimes. *PLoS Pathog* 18(8):e1010670. <https://doi.org/10.1371/journal.ppat.1010670>
- Lebrun L, Absil L, Rummelink M, De Mendonça R, D'Haene N, Gaspard N, Rusu S, Racu ML, Collin A, Allard J, Zindy E, Schiavo AA, De Clercq S, De Witte O, Decaestecker C, Lopes MB (2023) SARS-Cov-2 infection and neuropathological findings: a report of 18 cases and review of the literature. *Acta Neuropathol Commun* 11(1):78. <https://doi.org/10.1186/s40478-023-01566-1>. PMID: 37165453; PMCID: PMC10170054
- Leta V, Urso D, Batzu L, Lau YH, Mathew D, Boura I, Raeder V, Falup-Pecurariu C, van Wamelen D, Ray Chaudhuri K (2022) Viruses, parkinsonism and parkinson's disease: the past, present and future. *J Neural Transm (vienna)* 129(9):1119–1132
- Levine KS, Leonard HL, Blauwendraat C, Iwaki H, Johnson N, Bandler-Ciga S, Ferrucci L, Faghri F, Singleton AB, Nalls MA (2023) Virus exposure and neurodegenerative disease risk across national biobanks. *Neuron* 111(7):1086–1093. <https://doi.org/10.1016/j.neuron.2022.12.029>
- Li Q, Barres B (2018) Microglia and macrophages in brain homeostasis and disease. *Nat Rev Immunol* 18:225–242. <https://doi.org/10.1038/nri.2017.125>
- Limphaibool N, Iwanowski P, Holstad MJV, Kobylarek D, Kozubski W (2019) Infectious etiologies of parkinsonism: pathomechanisms and clinical implications. *Front Neurol* 19(10):652. <https://doi.org/10.3389/fneur.2019.00652>. PMID: 31275235; PMCID: PMC6593078
- Linard M, Ravier A, Mougé L, Grgurina I, Boutillier AL, Foubert-Samier A, Blanc F, Helmer C (2022) Infectious agents as potential drivers of  $\alpha$ -Synucleinopathies. *Mov Disord* 37(3):464–477. <https://doi.org/10.1002/mds.28925>. (Epub 2022 Jan 18 PMID: 35040520)
- Lingor P, Demleitner AF, Wolff AW, Feneberg E, (2022) SARS-CoV-2 and neurodegenerative diseases: what we know and what we don't. *J Neural Transm (Vienna)*. 129 (9):1155–1167. doi: <https://doi.org/10.1007/s00702-022-02500-w>. Epub 2022 Apr 17. PMID: 35434769; PMCID: PMC9013492.
- Lopez-Leon S, Wegman-Ostrosky T, Perelman C, Sepulveda R, Rebolledo PA, Cuapio A, Villapol S (2021) More than 50 Long-term effects of COVID-19: a systematic review and meta-analysis. *medRxiv [Preprint]*. 2021 Jan 30:2021.01.27.21250617. <https://doi.org/10.1101/2021.01.27.21250617>. Update in: *Sci Rep*. 2021 Aug 9;11(1):16144. PMID: 33532785; PMCID: PMC7852236.
- Lv QK, Tao KX, Wang XB, Yao XY, Pang MZ, Liu JY, Wang F (2023) Role of  $\alpha$ -synuclein in microglia: autophagy and phagocytosis balance neuroinflammation in Parkinson's disease. *Inflamm Res* 72(3):443–462. <https://doi.org/10.1007/s00011-022-01676-x>. (Epub 2023 Jan 4 PMID: 36598534)
- Maa MC, Chang M, Jiarung Li Y, Li Y, Hsieh MY, Yang CF, Chen YJ et al (2011) The INOS/Src/FAK Axis Is critical in toll-like receptor-mediated cell motility in macrophages. *Biochim Biophys Acta* 1813(1):136–147. <https://doi.org/10.1016/j.bbamcr.2010.09.004>
- Mahallawi WH, Khabour OF, Zhang Q, Makhdoum HM, Suliman BA (2018) MERS-CoV infection in humans is associated with a pro-inflammatory Th1 and Th17 cytokine profile. *Cytokine*. <https://doi.org/10.1016/j.cyto.2018.01.025>
- Mandrekar-Colucci S, Landreth GE (2010) Microglia and inflammation in Alzheimers disease. *CNS Neurological Disorders Drug Targets* 9(2):156–167. <https://doi.org/10.2174/187152710791012071>
- Mangold CA, Szpara ML (2019) Persistent Infection with herpes simplex virus 1 and Alzheimer's Disease—a call to study how variability in both virus and host may impact disease. *Viruses* 11(10):966. <https://doi.org/10.3390/v11100966>
- Maphis N, Xu G, Kokiko-Cochran ON, Jiang S, Cardona A, Ransohoff RM, Lamb BT, Bhaskar K (2015) Reactive microglia drive tau

- pathology and contribute to the spreading of pathological tau in the brain. *Brain*. <https://doi.org/10.1093/brain/awv081>
- Marino J, Maubert ME, Mele AR, Spector C, Wigdahl B, Nonnemacher MR (2020) Functional Impact of HIV-1 Tat on cells of the CNS and Its role in HAND. *Cellular Molecular Life Sci : CMLS* 77(24):5079–5099. <https://doi.org/10.1007/s00018-020-03561-4>
- Martin MDGM, Paes VR, Cardoso EF, Neto CEBP, Kanamura CT, Leite CDC, Otaduy MCG, Monteiro RAA, Mauad T, da Silva LFF, Castro LHM, Saldiva PHN, Dolhnikoff M (2022) Duarte-Neto AN Postmortem brain 7T MRI with minimally invasive pathological correlation in deceased COVID-19 subjects. *Insights Imaging* 13(1):7. <https://doi.org/10.1186/s13244-021-01144-w>. PMID: 35032223; PMCID: PMC8760871
- McArthur JC, Hoover DR, Bacellar H, Miller EN, Cohen BA, Becker JT, Graham NM, McArthur JH, Selnes OA, Jacobson LP et al (1993) Dementia in AIDS patients: incidence and risk factors. *Multicenter AIDS Cohort Study Neurology* 43(11):2245–2252. <https://doi.org/10.1212/wnl.43.11.2245>. (PMID: 8232937)
- McArthur JC, Haughey N, Gartner S et al (2003) Human immunodeficiency virus-associated dementia: An evolving disease. *J Neurovirol* 9:205–221
- McCall S, Henry JM, Reid A (2001) Influenza RNA not detected in archival brain tissues from acute encephalitis lethargica cases or in postencephalitic parkinson cases. *J Neuropathol Exp Neurol* 60:696–704. <https://doi.org/10.1093/jnen/60.7.696>
- McGeer PL (2004) Inflammation and neurodegeneration in Parkinson's disease. *Parkinsonism Relat Disord* 10(Suppl 1):S3-7. <https://doi.org/10.1016/j.parkreldis.2004.01.005>. (PMID: 15109580)
- McGeer EG (2007) The role of anti-inflammatory agents in Parkinson's disease. *CNS Drugs* 21(10):789–797. <https://doi.org/10.2165/00023210-200721100-00001>. (PMID: 17850169)
- McGeer PL, McGeer EG (2008) Glial reactions in Parkinson's disease. *Mov Disord* 23(4):474–483. <https://doi.org/10.1002/mds.21751>
- McGeer PL, Rogers J (2006) Inflammation, anti-inflammatory agents and Alzheimer disease: the last 12 years. *J Alzheimers Dis* 9(3 Suppl):271–276. <https://doi.org/10.3233/jad-2006-9s330>. (PMID: 16914866)
- McGeer PL, Itagaki S, Boyes BE (1988) Reactive microglia are positive for HLA-DR in the substantia nigra of Parkinson's and Alzheimer's disease brains. *Neurology* 38(8):1285–1291. <https://doi.org/10.1212/wnl.38.8.1285>. (PMID: 3399080)
- McTigue DM (2008) Potential therapeutic targets for PPAR after spinal cord injury. *PPAR Res* 2008:1–7. <https://doi.org/10.1155/2008/517162>
- Meisner F, Neuen-Jacob E, Sopper S, Schmidt M, Schlamme S, Scheller C, Vosswinkel D, Ter Meulen V, Riederer P (2008) Disruption of excitatory amino acid transporters in brains of SIV-infected rhesus macaques is associated with microglia activation. *J Neurochem* 104(1):202–209. <https://doi.org/10.1111/j.1471-4159.2007.05007.x>. (Epub 2007 Nov 6 PMID: 17986224)
- Mesias VSD, Zhu H, Tang X, Dai X, Liu W, Guo Y, Huang J (2022) Moderate binding between two SARS-CoV-2 protein segments and  $\alpha$ -synuclein alters its toxic oligomerization propensity differently. *J Phys Chem Lett* 13(45):10642–10648. <https://doi.org/10.1021/acs.jpcclett.2c02278>
- Mestre L, Alonso G, Feliú A, Mecha M, Martín C, Villar LM, Guaza C (2021) Aging and neuroinflammation: changes in immune cell responses, axon integrity, and motor function in a viral model of progressive multiple sclerosis. *Aging Cell*. <https://doi.org/10.1111/ace1.13440>
- Mihaly SR, Ninomiya-Tsuji J, Morioka S (2014) TAK1 control of cell death. *Cell Death Differentiation* 21(11):1667–1676. <https://doi.org/10.1038/cdd.2014.123>
- Misra UK (2002) Prognosis of Japanese encephalitis patients with dystonia compared to those with parkinsonian features only. *Postgrad Med J* 78(918):238–241. <https://doi.org/10.1136/pmj.78.918.238>
- Mogi M, Harada M, Riederer P, Narabayashi H, Fujita K, Nagatsu T (1994) Tumor necrosis factor- $\alpha$  (TNF- $\alpha$ ) increases both in the brain and cerebrospinal fluid from parkinsonian patients. *Neurosci Lett* 165:208–210
- Mogi M, Harada M, Kondo T, Riederer P, Inagaki H, Minami M (1994) Interleukin-1 beta, interleukin-6, epidermal growth factor and transforming growth factor-alpha are elevated in the brain from parkinsonian patients. *Neurosci Lett* 180(2):147–150. [https://doi.org/10.1016/0304-3940\(94\)90508-8](https://doi.org/10.1016/0304-3940(94)90508-8). (PMID: 7700568)
- Mogi M, Harada M, Kondo T, Narabayashi H, Riederer P, Nagatsu T (1995a) Transforming growth factor- $\beta$ 1 levels are elevated in the striatum and in ventricular cerebrospinal fluid in Parkinson's disease. *Neurosci Lett* 193:129–132
- Mogi M, Harada M, Kondo T, Riederer P, Nagatsu T (1995b) Brain  $\beta$ <sub>2</sub>-microglobulin levels are elevated in the striatum in Parkinson's disease. *J Neural Transm* 9:87–92
- Mogi M, Harada M, Narabayashi H, Inagaki H, Minami M, Nagatsu T (1996a) Interleukin (IL)-1 $\beta$ , IL-2, IL-4, IL-6, and transforming growth factor- $\beta$  levels are elevated in ventricular cerebrospinal fluid of juvenile parkinsonism and Parkinson's disease. *Neurosci Lett* 193:129–132
- Mogi M, Harada M, Kondo T, Mizuno Y, Riederer P, Nagatsu T (1996b) Bcl-2 protein is increased in the brain from parkinsonian patients. *Neurosci Lett* 215:137–139
- Mudra Rakshasa-Loots A, Whalley HC, Vera JH, Cox SR (2022) Neuroinflammation in HIV-associated depression: evidence and future perspectives. *Mol Psychiatry* 27(9):3619–3632. <https://doi.org/10.1038/s41380-022-01619-2>. (Epub 2022 May 26. PMID: 35618889; PMCID: PMC9708589)
- Mukerji SS, Solomon IH (2021) What can we learn from brain autopsies in COVID-19? *Neurosci Lett*. <https://doi.org/10.1016/j.neulet.2020.135528>
- Müller L, Di Benedetto S (2023) Aged brain and neuroimmune responses to COVID-19: post-acute sequelae and modulatory effects of behavioral and nutritional interventions. *Immun Ageing* 20:17. <https://doi.org/10.1186/s12979-023-00341-z>
- Müller T, Riederer P (2023) Aminoadamantanes: from treatment of Parkinson's and Alzheimer's disease to symptom amelioration of long COVID-19 syndrome? *Expert Rev Clin Pharmacol* 16(2):101–107. <https://doi.org/10.1080/17512433.2023.2176301>. (Epub 2023 Feb 8 PMID: 36726198)
- Nagatsu T, Nakashima A, Watanabe H, Ito S, Wakamatsu K, Zucca FA, Zecca L, Youdim M, Wulf M, Riederer P, Dijkstra JM (2023) The role of tyrosine hydroxylase as a key player in neuromelanin synthesis and the association of neuromelanin with Parkinson's disease. *J Neural Transm (vienna)*. <https://doi.org/10.1007/s00702-023-02617-6>
- Nash Y, Schmukler E, Trudler D, Pinkas-Kramarski R (2017) DJ-1 deficiency impairs autophagy and reduces alpha-synuclein phagocytosis by microglia. *J Neurochem* 143(5):584–594. <https://doi.org/10.1111/jnc.14222>. (Epub 2017 Oct 30 PMID: 28921554)
- NICE: COVID-19 rapid guideline: managing the long-term effects of COVID-19. London: National Institute for Health and Care Excellence (NICE); 2020 Dec 18. PMID: 33555768.
- Oeckinghaus A, Hayden MS, Ghosh S (2011) Crosstalk in NF- $\kappa$ B signaling pathways. *Nat Immunol* 12(8):695–708. <https://doi.org/10.1038/ni.2065>
- Oliver KR, Brennan P (1997) Specific infection and destruction of dopaminergic neurons in the substantia nigra by Theiler's virus. *J Virol* 71(8):6179–6182. <https://doi.org/10.1128/JVI.71.8.6179-6182.1997>. PMID: 9223513; PMCID: PMC191879

- Orihuela R, McPherson CA, Harry GJ (2015) Microglial M1/M2 Polarization and Metabolic States. *Br J Pharmacol* 173(4):649–665. <https://doi.org/10.1111/bph.13139>
- Pathak D, Sriram K (2023) Molecular mechanisms underlying neuroinflammation elicited by occupational injuries and toxicants. *Int J Mol Sci* 24(3):2272. <https://doi.org/10.3390/jms24032272>
- Patrick KL, Bell SL, Weindel CG and Watson RT, (2019). “Exploring the ‘Multiple-Hit Hypothesis’ of Neurodegenerative Disease: Bacterial Infection Comes up to Bat” <https://doi.org/10.3389/fcimb.2019.00138>.
- Pekny M, Pekna M, (2016). “Reactive Gliosis in the Pathogenesis of CNS Diseases.” *Biochimica et Biophysica Acta (BBA) Molecular Basis of Disease* 1862 (3): 483–91. <https://doi.org/10.1016/j.bbadis.2015.11.014>.
- Penninx BWJH (2021) Psychiatric symptoms and cognitive impairment in “Long COVID”: the relevance of immunopsychiatry. *World Psychiatry* 20:357–358. <https://doi.org/10.1002/wps.20913>
- Philippens IHCHM, Böszörményi KP, Wubben JAM, Fagrouch ZC, van Driel N, Mayenburg AQ, Lozovaglia D, Roos E, Schurink B, Bugiani M, Bontrop RE, Middeldorp J, Bogers WM, de Geus-Oei LF, Langermans JAM, Verschoor EJ, Stammes MA, Verstrepen BE (2022) Brain inflammation and intracellular  $\alpha$ -synuclein aggregates in macaques after SARS-CoV-2 infection. *Viruses* 14(4):776. <https://doi.org/10.3390/v14040776>
- Phillips TR, Billaud JN (2000) Methamphetamine and HIV-1: potential interactions and the use of the FIV/cat model. *J Psychopharmacol* 14(3):244–250. <https://doi.org/10.1177/026988110001400309>. (PMID: 11106303)
- Piacentini R, De Chiara G, Li Puma DD, Ripoli C, Marcocci ME, Garaci E, Palamara AT (2014) HSV-1 and Alzheimer’s disease: more than a hypothesis. *Front Pharmacol* 7(5):97. <https://doi.org/10.3389/fphar.2014.00097>. PMID:24847267;PMCID:PMC4019841
- Piacenza L, Zeida A, Trujillo M, Radi R (2022) The superoxide radical switch in the biology of nitric oxide and peroxynitrite. *Physiol Rev*. <https://doi.org/10.1152/physrev.00005.2022>
- Piekut T, Hurla M, Banaszek N, Szejn P, Dorszewska J, Kozubski W (2022) Infectious agents and Alzheimer’s disease. *J Integr Neurosci* 21(2):73. <https://doi.org/10.31083/j.jin2102073>. (PMID: 35364661)
- Pike AF, Longhena F, Faustini G, van Eik JM, Gombert I, Herrebut MAC, Fayed MMHE, Sandre M, Varanita T, Teunissen CE, Hoozemans JJM, Bellucci A, Veerhuis R (2022) Dopamine signaling modulates microglial NLRP3 inflammasome activation: implications for Parkinson’s disease. *J Neuroinflammation* 19(1):50. <https://doi.org/10.1186/s12974-022-02410-4>. PMID: 35172843;PMCID:PMC8848816
- Porcher L, Bruckmeier S, Burbano SD, Finnell JE, Gorny N, Klett J, Wood SK (2021) Aging triggers an upregulation of a multitude of cytokines in the male and especially the female rodent hippocampus but more discrete changes in other brain regions. *J Neuroinflammation* 18(1):219. <https://doi.org/10.1186/s12974-021-02252-6>. PMID:34551810;PMCID:PMC8459490
- Premraj L, Kannapadi NV, Briggs J, Seal SM, Battaglini D, Fanning J, Suen J, Robba C, Fraser J (2022) Mid and long-term neurological and neuropsychiatric manifestations of post-COVID-19 syndrome: a meta-analysis. *J Neurol Sci* 15(434):120162. <https://doi.org/10.1016/j.jns.2022.120162>
- Prinz M, Jung S (2019) Microglia biology: one century of evolving concepts. *Cell* 179(2):292–311. <https://doi.org/10.1016/j.cell.2019.08.053>
- Profaci CP, Munji RN, Pulido RS (2020) The blood-brain barrier in health and disease: important unanswered questions. *J Exp Med* 217(4):e20190062. <https://doi.org/10.1084/jem.20190062>. PMID: 32211826;PMCID:PMC7144528
- Pu H, Hayashi K, Andras IE, Eum SY, Hennig B (2007) Limited role of COX-2 in HIV Tat-induced alterations of tight junction protein expression and disruption of the blood-brain barrier. *Brain Res* 12(1184):333–344. <https://doi.org/10.1016/j.brainres.2007.09.063>. (Epub 2007 Oct 2 PMID: 17976544)
- Quintanilla RA, Orellana DI, González-Billault C (2004) Interleukin-6 induces Alzheimer-type phosphorylation of tau protein by deregulating the cdk5/p35 pathway. *Exp Cell Res* 295(1):245–257. <https://doi.org/10.1016/j.yexcr.2004.01.002>. (PMID: 15051507)
- Rahman I (2006) Oxidative stress and redox regulation of lung inflammation in COPD. *Eur Respir J* 28(1):219–242. <https://doi.org/10.1183/09031936.06.00053805>
- Ramatillah DL, Gan SH, Pratiwi I, Syed Sulaiman SA, Jaber AAS, Jusnita N, Lukas S (2022) Impact of cytokine storm on severity of COVID-19 disease in a private hospital in West Jakarta prior to vaccination. *PLoS ONE* 17(1):e0262438. <https://doi.org/10.1371/journal.pone.0262438>. PMID:35077495;PMCID:PMC8789122
- Rangaraju S, Dammer EB, Raza SA, Rathakrishnan P, Xiao H, Gao T, Duong DM, Pennington MW, Lah JJ, Seyfried NT (2018) Identification and therapeutic modulation of a pro-inflammatory subset of disease-associated-microglia in Alzheimer’s disease. *Mol Neurodegener* 13(1):24. <https://doi.org/10.1186/s13024-018-0254-8>. PMID:29784049;PMCID:PMC5963076
- Rausch WD, Rossmannith W, Gruber J, Riederer P, Jellinger K, Weiser M, (1983). Studies on the Neurotransmitter Binding to Pig Brain Microvessels. In: Goldstein, M., Jellinger, K., Riederer, P. (eds) Basic Aspects of Receptor Biochemistry. Journal of Neural Transmission, Springer, Vienna
- Ray B, Mahalakshmi AM, Tuladhar S, Bhat A, Srinivasan A, Pellegrino C, Kannan A, Bolla SR, Chidambaram SB, Sakharkar MK (2021) ‘Janus-faced’  $\alpha$ -Synuclein: Role in Parkinson’s Disease. *Frontiers Cell Develop Biol*. <https://doi.org/10.3389/fcell.2021.673395>
- Reid AH, McCall S, Henry JM (2001) Experimenting on the past: the enigma of von Economo’s encephalitis lethargica. *J Neuropathol Exp Neurol* 60(7):663–670. <https://doi.org/10.1093/jnen/60.7.663>. (PMID: 11444794)
- Ren C, Li D, Zhou Q, Hu X (2020) Mitochondria-targeted TPP-MoS2 with dual enzyme activity provides efficient neuroprotection through M1/M2 microglial polarization in an Alzheimer’s disease model. *Biomaterials* 232:119752. <https://doi.org/10.1016/j.biomaterials.2019.119752>
- Repici M, Giorgini F (2019) DJ-1 in Parkinson’s disease: clinical insights and therapeutic perspectives. *J Clin Med* 8(9):1377. <https://doi.org/10.3390/jcm8091377>
- Riederer P, Berg D, Casadei N, Cheng F, Classen J, Dresel C, Jost W, Krüger R, Müller T, Reichmann H, Rieß O, Storch A, Strobel S, van Eimeren T, Völker HU, Winkler J, Winklhofer KF, Wülner U, Zunke F, Monoranu CM (2019)  $\alpha$ -Synuclein in Parkinson’s disease: causal or bystander? *J Neural Transm (vienna)* 126(7):815–840. <https://doi.org/10.1007/s00702-019-02025-9>. (Epub 2019 Jun 25 PMID: 31240402)
- Riederer P, Monoranu C, Strobel S, Iordache T, Sian-Hulsmann J (2021) Iron as the concert master in the pathogenic orchestra playing in sporadic Parkinson’s disease. *J Neural Transm (vienna)* 128(10):1577–1598. <https://doi.org/10.1007/s00702-021-02414-z>
- Riederer P, Nagatsu T, Youdim MBH, Wulf M, Dijkstra JM, Sian-Hulsmann J (2023) Lewy bodies, iron, inflammation and neuromelanin: pathological aspects underlying Parkinson’s disease. *J Neural Transm (vienna)* 130(5):627–646
- Riederer P, Ter Meulen V, (2020) Coronaviruses: a challenge of today and a call for extended human postmortem brain analyses. *J Neural Transm (Vienna)*. 127 (9):1217–1228. doi: <https://doi.org/10.1007/s00702-020-02230-x>. Epub 2020 Jul 28. Erratum

- in: *J Neural Transm* (Vienna). 2022 Jan;129(1):127. PMID: 32725545; PMCID: PMC7386201.
- Roberson ED, Scearce-Levie K, Palop JJ, Yan F, Cheng IH, Wu T, Gerstein H, Yu GQ (2007) Reducing endogenous tau ameliorates amyloid beta-induced deficits in an Alzheimer's disease mouse model. *Science* 316(5825):750–754. <https://doi.org/10.1126/science.1141736>. (PMID: 17478722)
- Robinson R, Shahida S, Madan N, Rao S and Khardori N (2003). "Transient Parkinsonism in West Nile Virus Encephalitis" 115 (3): 252–53. [https://doi.org/10.1016/s0002-9343\(03\)00291-2](https://doi.org/10.1016/s0002-9343(03)00291-2).
- Rocha SM, Cristovão AC, Campos FL, Fonseca CP, Baltazar G (2012) Astrocyte-derived GDNF is a potent inhibitor of microglial activation. *Neurobiol Dis* 47(3):407–415. <https://doi.org/10.1016/j.nbd.2012.04.014>
- Rohr O, Sawaya BE, Lecestre D, Aunis D, Schaeffer E (1999) Dopamine stimulates expression of the human immunodeficiency virus type 1 via NF-kappaB in cells of the immune system. *Nucleic Acids Res* 27:3291–3299
- Romero JR, Newland JG (2003) Viral meningitis and encephalitis: traditional and emerging viral agents. *Seminars Pediatric Infectious Diseases* 14(2):72–82. <https://doi.org/10.1053/spid.2003.127223>
- Roth LM, Akay-Espinoza C, Grinspan JB, Jordan-Sciutto KL (2021) HIV-induced neuroinflammation inhibits oligodendrocyte maturation via glutamate-dependent activation of the PERK arm of the integrated stress response. *Glia* 69(9):2252–2271. <https://doi.org/10.1002/glia.24033>
- Ruganzu JB, Zheng Q, Wu X, He Y, Peng X, Jin H, Zhou J, Ma R, Ji S, Ma Y, Qian Y, Wang Y, Yang W (2021) TREM2 overexpression rescues cognitive deficits in APP/PS1 transgenic mice by reducing neuroinflammation via the JAK/STAT/SOCS signaling pathway. *Exp Neurol* 336:113506. <https://doi.org/10.1016/j.expneurol.2020.113506>
- Sagui E, Janvier F, Baize S, Foissaud V, Koulibaly F, Savini H, Maugé N, Aletti M, Granier H (2015) Severe ebola virus infection with encephalopathy: evidence for direct virus involvement. *Clin Infect Dis* 61(10):1627–1628. <https://doi.org/10.1093/cid/civ606>. (Epub 2015 Jul 21 PMID: 26197842)
- Saniasiaya J, Islam MA, Abdullah B (2019) Prevalence of olfactory dysfunction in coronavirus disease 2019 (COVID-19): A Meta-analysis of 27,492 patients. *Laryngoscope* 131(4):865–878. <https://doi.org/10.1002/lary.29286>
- Sawaya BE, Rohr O, Aunis D (1996) Chicken ovalbumin upstream promoter transcription factor, a transcriptional activator of HIV-1 gene expression in human brain cells. *J Biol Chem* 271(38):23572–23576. <https://doi.org/10.1074/jbc.271.38.23572>. (PMID: 8798567)
- Scarlsbrick IA, Rodriguez M (2003) Hit-Hit and hit-Run: viruses in the playing field of multiple sclerosis. *Curr Neurol Neurosci Rep* 3(3):265–271. <https://doi.org/10.1007/s11910-003-0087-9>
- Scheller C, Sopper S, Jassoy C, ter Meulen V, Riederer P (2000) Dopamine activates HIV in chronically infected T lymphoblasts. *J Neural Transm* (vienna) 107(12):1483–1489. <https://doi.org/10.1007/s007020070012>. (PMID: 11459001)
- Schilling S, Chausse B, Dikmen HO, Almouhanna F, Hollnagel JO, Lewen A (2021) TLR2- and TLR3-activated microglia induce different levels of neuronal network dysfunction in a context-dependent manner. *Brain Behav Immun* 96:80–91. <https://doi.org/10.1016/j.bbi.2021.05.013>. (Epub 2021 May 17 PMID: 34015428)
- Schneider SA, Desai S, Phokaewvarangkul O, Rosca EC, Sringean J, Anand P, Bravo GA, Cardoso F, Cervantes-Arslanian AM, Chovatiya H, Crosiers D, Dijkstra F, Fearon C, Grandas F, Guedj E, Méndez-Guerrero A, Hassan M, Jankovic J, Lang AE, Makhouk K, Muccioli L, O'Shea SA, Ostovan VR, Perez-Sanchez JR, Ramdhani R, Ros-Castelló V, Schulte C, Shah P, Wojtecki L, Pal PK (2023) COVID19-associated new-onset movement disorders: a follow-up study. *J Neurol* 270(5):2409–2415. <https://doi.org/10.1007/s00415-023-11661-x>
- Schreiber-Stainthorp W, Solomon J, Lee JH, Castro M, Shah S, Martinez-Orengo N, Reeder R, Maric D, Gross R, Qin J, Hagen KR, Johnson RF (2021) Longitudinal in vivo imaging of acute neuropathology in a monkey model of Ebola virus infection. *Nat Commun* 12(1):2855. <https://doi.org/10.1038/s41467-021-23088-x>. PMID:34001896;PMCID:PMC8129091
- Sejvar J (2014) Clinical Manifestations and Outcomes of West Nile Virus Infection. *Viruses* 6(2):606–623. <https://doi.org/10.3390/v6020606>
- Semerdzhev SA, Fakhree MAA, Segers-Nolten I, Blum C, Claessens MMAE (2022) Interactions between SARS-CoV-2 N-protein and  $\alpha$ -synuclein accelerate amyloid formation. *ACS Chem Neurosci* 13(1):143–150. <https://doi.org/10.1021/acscchemneuro.1c00666>
- Sempere RN, Armando A (2019) Establishment of a cell culture model of persistent flaviviral infection: usutu virus shows sustained replication during passages and resistance to extinction by antiviral Nucleosides. *Viruses*. <https://doi.org/10.3390/v11060560>
- Serrano-Pozo A, Mielke ML, Gómez-Isla T, Betensky RA, Growdon JH, Frosch MP, Hyman BT (2011) Reactive glia not only associates with plaques but also parallels tangles in Alzheimer's disease. *Am J Pathol* 179(3):1373–1384. <https://doi.org/10.1016/j.ajpath.2011.05.047>
- Shi Y, Yamada K, Liddelow SA, Smith ST, Zhao L, Luo W, Tsai RM, Spina S, Grinberg LT, Rojas JC, Gallardo G, Wang K, Roh J, Robinson G, Finn MB, Jiang H, Sullivan PM, Baufeld C, Wood MW, Sutphen C, McCue L, Xiong C, Del-Aguila JL, Morris JC, Cruchaga C; Alzheimer's Disease Neuroimaging Initiative; Fagan AM, Miller BL, Boxer AL, Seeley WW, Butovsky O, Barres BA, Paul SM, Holtzman DM, (2017) ApoE4 markedly exacerbates tau-mediated neurodegeneration in a mouse model of tauopathy. *Nature* <https://doi.org/10.1038/nature24016>. Epub 2017 Sep 20. PMID: 28959956; PMCID: PMC5641217.
- Sian-Hulsmann J (2021) The nigral coup in parkinson's disease by  $\alpha$ -synuclein and its associated rebels. *Cells* 10(3):598. <https://doi.org/10.3390/cells10030598>. PMID:33803185;PMCID:PMC8000327
- Sienski G, Narayan P, Bonner JM, Kory N, Boland S, Arczewska AA, Ralvenius WT, Akay L, Lockshin E, He L, Milo B, Graziosi A, Baru V, Lewis CA, Kellis M, Sabatini DM, Tsai LH, Lindquist S (2021) *APOE4* disrupts intracellular lipid homeostasis in human iPSC-derived glia. *Sci Transl Med*. <https://doi.org/10.1126/scitranslmed.aaz4564>. PMID:33658354;PMCID:PMC8218593
- Sierra A, Abiega O, Shahraz A, Neumann H (2013) Janus-faced microglia: beneficial and detrimental consequences of microglial phagocytosis. *Frontiers Cellular Neurosci*. <https://doi.org/10.3389/fncel.2013.00006>
- Simpson DSA, Oliver PL (2020) ROS Generation in microglia: understanding oxidative stress and inflammation in neurodegenerative disease. *Antioxidants* 9(8):743. <https://doi.org/10.3390/antiox9080743>
- Skok K, Stelzl E, Trauner M, Kessler HH (2021) Post-mortem viral dynamics and tropism in COVID-19 patients in correlation with organ damage. *Virchows Arch* 478(2):343–353. <https://doi.org/10.1007/s00428-020-02903-8>
- Smeyne RJ, Noyce AJ, Byrne M, Savica R (2021) Infection and risk of parkinson's disease. *J Parkinsons Dis* 11(1):31–43. <https://doi.org/10.3233/JPD-202279>. PMID:33361610;PMCID:PMC7990414
- Smeyne RJ, Eells JB, Chatterjee D, Byrne M, Akula SM, Sriramula S, O'Rourke DP, Schmidt P (2022) COVID-19 infection enhances susceptibility to oxidative stress-induced parkinsonism. *Mov Disord* 37(7):1394–1404. <https://doi.org/10.1002/mds.29116>
- Smith JA, Das A, Ray SK, Banik NL (2012) Role of pro-inflammatory cytokines released from microglia in neurodegenerative diseases.



- Brain Res Bull 87(1):10–20. <https://doi.org/10.1016/j.brainresbu.2011.10.004>
- Solomon T (2004) Flavivirus encephalitis. *N Engl J Med* 351(4):370–378. <https://doi.org/10.1056/nejmra030476>
- Solomon IH, Singh A, Folkerth RD (2023) What can we still learn from brain autopsies in COVID-19? *Semin Neurol* 43(2):195–204. <https://doi.org/10.1055/s-0043-1767716>. (Epub 2023 Apr 6 PMID: 37023787)
- Song GJ, Nam Y, Jo M, Jung M, Koo JY, Cho W, Koh M, Park SB, Suk K (2016) A novel small-molecule agonist of PPAR- $\gamma$  potentiates an anti-inflammatory M2 glial phenotype. *Neuropharmacology* 109:159–169. <https://doi.org/10.1016/j.neuropharm.2016.06.009>
- Steinman L, Rothbard JB, Kurnellas MP (2014) Janus faces of amyloid proteins in neuroinflammation. *J Clin Immunol* 34(S1):61–63. <https://doi.org/10.1007/s10875-014-0034-3>
- Streit WJ, Mrak RE, Griffin WST (2004) Microglia and neuroinflammation: a pathological perspective. *J Neuroinflammation* 1:14. <https://doi.org/10.1186/1742-2094-1-14>
- Stykel MG, and Scott RD, (2022). *Nitrosative stress in Parkinson's disease*. [online] Available at: <https://www.nature.com/articles/s41531-022-00370-3>.
- Sulzer D, Antonini A, Leta V, Nordvig A, Smeyne RJ, Goldman JE, Al-Dalahmah O, Zecca L, Sette A, Bubacco L, Meucci O, Moro E, Harms AS, Xu Y, Fahn S (2020) COVID-19 and possible links with Parkinson's disease and parkinsonism: from bench to bedside. *NPJ Parkinsons Dis* 20(6):18. <https://doi.org/10.1038/s41531-020-00123-0>. PMID:32885037;PMCID:PMC7441399
- Sun XY, Li LJ, Dong QX, Zhu J, Huang YR, Hou SJ, Yu XL (2021) Rutin prevents tau pathology and neuroinflammation in a mouse model of Alzheimer's disease. *J Neuroinflammation* 18(1):131. <https://doi.org/10.1186/s12974-021-02182-3>. PMID:34116706; PMCID:PMC8196535
- Sung MJu, Song HY, Ji Lee JH, Lee SH, Choi SY and Park J (2009). "Extracellular HIV-1 Tat Up-Regulates Expression of Matrix Metalloproteinase-9 via a MAPK-NF-KB Dependent Pathway in Human Astrocytes" 41 (2): 86–86. <https://doi.org/10.3858/emmm.2009.41.2.011>.
- Szabo MP, Iba M, Nath A, Maslah E, Kim C (2022) Does SARS-CoV-2 affect neurodegenerative disorders? TLR2, a potential receptor for SARS-CoV-2 in the CNS. *Exp Mol Med* 54(4):447–454. <https://doi.org/10.1038/s12276-022-00755-7>
- Szabo S, Gyires K, Zayachkivska O (2023) Introduction to the 'long COVID' special issue. *Inflammopharmacology* 31(2):549. <https://doi.org/10.1007/s10787-023-01197-x>
- Szpakowski P, Ksiazek-Winiarek D, Turniak-Kusy M, Pacan I, Glabinski A (2022) Human primary astrocytes differently respond to pro- and anti-inflammatory stimuli. *Biomedicines*. 10(8):1769. <https://doi.org/10.3390/biomedicines10081769>. PMID:35892669;PMCID:PMC9331936
- Tabor-Godwin JM, Ruller CM, Bagalzo N, An N, Pagarigan RR, Harkins S, Gilbert PE, Kiesses WB, Gude NA, Cornell CT, Doran KS, Sussman MA, Whitton JL (2010) A novel population of myeloid cells responding to coxsackievirus infection assists in the dissemination of virus within the neonatal CNS. *J Neurosci* 30(25):8676–8691. <https://doi.org/10.1523/JNEUROSCI.1860-10.2010>. PMID:20573913;PMCID:PMC2902258
- Takahashi M (1999) Viral etiology for Parkinson's disease—a possible role of influenza a virus infection. *Jpn J Infect Dis* 52(3):89–98 (PMID: 10507986)
- Tang RH, Qi RQ, Liu HY (2019) Interleukin-4 affects microglial autophagic flux. *Neural Regen Res* 14(9):1594–1602. <https://doi.org/10.4103/1673-5374.255975>
- Tansey MG, Goldberg MS (2010) Neuroinflammation in Parkinson's disease: its role in neuronal death and implications for therapeutic intervention. *Neurobiol Dis* 37(3):510–518. <https://doi.org/10.1016/j.nbd.2009.11.004>
- Tewari D, Sah AN, Bawari S, Nabavi SF, Dehpour AR, Shirooie S, Braidy N, Fiebich BL, Vacca RA, Nabavi SM (2020) Role of nitric oxide in neurodegeneration: function, regulation, and inhibition. *Curr Neuropharmacol* 19(2):114–126. <https://doi.org/10.2174/1570159x18666200429001549>
- Thomas SJ, Endy TP, Rothman AL and Barrett AD (2014). Flaviviruses (Dengue, Yellow Fever, Japanese Encephalitis, West Nile Encephalitis, St. Louis Encephalitis, Tick-Borne Encephalitis, Kyasanur Forest Disease, Alkhurma Hemorrhagic Fever, Zika). In *Mandell, Douglas, and Bennett's Principles and Practice of Infectious Diseases* (Vol. 2, pp. 1881–1903.e6). Elsevier Inc.. <https://doi.org/10.1016/B978-1-4557-4801-3.00155-7>
- Tohidpour A, Morgun AV, Boitsova EB, Malinovskaya NA, Martynova GP, Khilazheva ED, Kopylevich NV, Gertsog GE (2017) Neuroinflammation and Infection: molecular mechanisms associated with dysfunction of neurovascular unit. *Front Cell Infect Microbiol* 20(7):276. <https://doi.org/10.3389/fcimb.2017.00276>. PMID: 28676848;PMCID:PMC5476750
- Toledano M, Davies NWS (2016) Enteroviral postencephalitic parkinsonism with evidence of impaired presynaptic dopaminergic function. *JAMA Neurol* 73(8):1023. <https://doi.org/10.1001/jamaneurol.2016.1043>
- Tseng KY, Wu JS, Chen YH, Airavaara M, Cheng CY (2022) Modulating microglia/macrophage activation by CDNF promotes transplantation of fetal ventral mesencephalic graft survival and function in a hemiparkinsonian rat model. *Biomedicines* 10(6):1446. <https://doi.org/10.3390/biomedicines10061446>. PMID:35740467; PMCID:PMC9221078
- Vera JH, Guo Q, Cole JH, Boasso A, Greathead L, Kelleher P, Rabiner EA, Kalk N, Bishop C, Gunn RN, Matthews PM, Winston A (2016) Neuroinflammation in treated HIV-positive individuals: A TSPO PET study. *Neurology*. 86 (15):1425–1432. <https://doi.org/10.1212/WNL.0000000000002485>. Epub 2016 Feb 24. PMID: 26911637; PMCID: PMC4831035.
- Verbist KC and Nichols KE (2019). "Cytokine Storm Syndromes Associated with Epstein-Barr Virus." *Cytokine Storm Syndrome*, 253–76. [https://doi.org/10.1007/978-3-030-22094-5\\_15](https://doi.org/10.1007/978-3-030-22094-5_15).
- Verghese PB, Castellano JM, Garai K, Wang Y, Jiang H, Shah A, Bu G, Frieden C, Holtzman DM, (2013) ApoE influences amyloid- $\beta$  (A $\beta$ ) clearance despite minimal apoE/A $\beta$  association in physiological conditions. *Proc Natl Acad Sci USA* 110 (19): E1807–16. doi: <https://doi.org/10.1073/pnas.1220484110>. Epub 2013 Apr 25. PMID: 23620513; PMCID: PMC3651443.
- Wahl A, Al-Harathi L (2023) HIV infection of non-classical cells in the brain. *Retrovirology* 20(1):1. <https://doi.org/10.1186/s12977-023-00616-9>. PMID:36639783;PMCID:PMC9840342
- Wang AS, Perez JA (2023) Frequency of Parkinson disease following COVID-19 infection: A two-year retrospective cohort study. *Parkinsonism Relat Disord* 111:105433. <https://doi.org/10.1016/j.parkreldis.2023.105433>. (Epub 2023 Apr 29 PMID: 37141688)
- Wang G, Yin P, Zhang W, Giulietta Fernandez C (2022) Editorial: the relationship of neuroinflammation with aging and neurodegenerative diseases. *Front Aging Neurosci* 9(14):1102613. <https://doi.org/10.3389/fnagi.2022.1102613>. PMID:36570539;PMCID: PMC9782399
- Wang C, Xiong M, Gratuze M, Bao X, Shi Y, Andhey PS, Manis M, Schroeder C, Yin Z, Madore C, Butovsky O, Artyomov M, Ulrich JD, Holtzman DM, (2021) Selective removal of astrocytic APOE4 strongly protects against tau-mediated neurodegeneration and decreases synaptic phagocytosis by microglia. *Neuron*. May 109 (10): 1657–1674.e7. doi: <https://doi.org/10.1016/j.neuron.2021.03.024>. Epub 2021 Apr 7. PMID: 33831349; PMCID: PMC8141024.

- Wang P, Jin L, Zhang M, Wu Y, Duan Z, Guo Y, Wang C, Guo Y, Chen W, Liao Z, Wang Y, Lai R, Lee LP, Qin J, (2023a) Blood-brain barrier injury and neuroinflammation induced by SARS-CoV-2 in a lung-brain microphysiological system. *Nat Biomed Eng.* <https://doi.org/10.1038/s41551-023-01054-w>. Epub ahead of print. PMID: 37349391.
- Wijarnpreecha K, Chesdachai S, Jaruvongvanich V (2018) Hepatitis C virus infection and risk of Parkinson's disease: a systematic review and meta-analysis. *Eur J Gastroenterol Hepatol* 30(1):9–13. <https://doi.org/10.1097/MEG.0000000000000991>. (PMID: 29049127)
- Wijeyekoon RS, Moore SF, Farrell K, Breen DP, Barker RA, Williams-Gray CH (2020) Cerebrospinal fluid cytokines and neurodegeneration-associated proteins in Parkinson's disease. *Mov Disord* 35(6):1062–1066. <https://doi.org/10.1002/mds.28015>
- Willems HL, Huo X-J, Mao-Ying Q-L, Zijlstra J, Heijnen CJ, Kavelaars A (2012) MicroRNA-124 as a novel treatment for persistent hyperalgesia. *J Neuroinflammation.* <https://doi.org/10.1186/1742-2094-9-143>
- Wojna V (2022) Neuroinflammation and HIV-associated cognitive impairment: can they be eradicated? *Neurology* 99(12):499–500. <https://doi.org/10.1212/WNL.00000000000021095>. (Epub 2022 Aug 2 PMID: 35918171)
- Wolf SA, Boddeke HWGM, Kettenmann H (2017) Microglia in physiology and disease. *Annu Rev Physiol* 79(1):619–643. <https://doi.org/10.1146/annurev-physiol-022516-034406>
- Wu WYY, Kang KH (2015) Hepatitis c virus infection: a risk factor for parkinson's disease. *J Viral Hepatitis* 22:784–791
- Wu Z, Zhang X, Huang Z (2022) SARS-CoV-2 proteins interact with alpha synuclein and induce lewy body-like pathology in vitro. *Int J Mol Sci* 23(6):3394. <https://doi.org/10.3390/ijms23063394>. PMID:35328814;PMCID:PMC8949667
- Wu CT, Lidsky PV, Xiao Y, Cheng R, Lee IT, Nakayama T, Jiang S, He W, Demeter J, Knight MG, Turn RE, Rojas-Hernandez LS, Ye C, Chiem K, Shon J, Martinez-Sobrido L, Bertozzi CR, Nolan GP, Nayak JV, Milla C, Andino R, Jackson PK (2023) SARS-CoV-2 replication in airway epithelia requires motile cilia and microvillar reprogramming. *Cell.* 186 (1):112–130. doi: <https://doi.org/10.1016/j.cell.2022.11.030>. Epub 2022 Dec 2. PMID: 36580912; PMCID: PMC9715480.
- Yacoubian TA, Fang YD, Gerstenecker A, Amara A, Stover N, Ruffrage L, Collette C, Kennedy R, Zhang Y, Hong H, Qin H, McConathy J, Benveniste EN (2023) Standaert DG brain and systemic inflammation in de novo parkinson's disease. *Mov Disord* 38(5):743–754. <https://doi.org/10.1002/mds.29363>. (Epub 2023 Feb 28 PMID: 36853618)
- Yamazaki Y, Zhao N, Caulfield TR, Liu CC, Bu G (2019) Apolipoprotein E and Alzheimer disease: pathobiology and targeting strategies. *Nat Rev Neurol.* 2019 Sep;15 (9): 501–518. doi: <https://doi.org/10.1038/s41582-019-0228-7>. Epub 2019 Jul 31. PMID: 31367008; PMCID: PMC7055192.
- Yang SH, Lee DK, Shin J, Lee S, Baek S, Kim J, Jung H, Hah JM (2017) Nec-1 alleviates cognitive impairment with reduction of A $\beta$  and tau abnormalities in APP/PS1 mice. *EMBO Mol Med* 9(1):61–77. <https://doi.org/10.15252/emmm.201606566>. PMID: 27861127;PMCID:PMC5210088
- Yao K, Zu H (2020) Microglial polarization: novel therapeutic mechanism against Alzheimer's disease. *Inflammopharmacology* 28(1):95–110. <https://doi.org/10.1007/s10787-019-00613-5>
- Yin F (2022) Lipid metabolism and alzheimer's disease: clinical evidence, mechanistic link and therapeutic promise. *The FEBS Journal*, January. <https://doi.org/10.1111/febs.16344>
- Yin J, Zhao F, Chojnacki JE, Fulp J, Klein WL, Zhang S, Zhu X (2018) NLRP3 Inflammasome Inhibitor Ameliorates Amyloid Pathology in a Mouse Model of Alzheimer's Disease. *Mol Neurobiol.* 55 (3):1977–1987. doi: <https://doi.org/10.1007/s12035-017-0467-9>. Epub 2017 Mar 2. PMID: 28255908; PMCID: PMC5585057.
- Zanza C, Romenskaya T, Manetti AC, Franceschi F, La Russa R, Bertozzi G, Maiese A, Savioli G, Volonnino G (2022) Cytokine storm in COVID-19: immunopathogenesis and therapy. *Medicina (kaunas)* 58(2):144. <https://doi.org/10.3390/medicina58020144>. PMID:35208467;PMCID:PMC8876409
- Zhai H, Lv Y, Xu Y, Wu Y, Zeng W, Wang T, Cao X, Xu Y (2021) Characteristic of Parkinson's disease with severe COVID-19: a study of 10 cases from Wuhan. *J Neural Transm (Vienna)*. 128 (1): 37–48. doi: <https://doi.org/10.1007/s00702-020-02283-y>. Epub 2021 Jan 3. PMID: 33392827; PMCID: PMC7779096.
- Zhang H, Li Y, Yu J, Guo M, Meng J, Liu C, Xie Y, Feng L, Xiao B, Ma C (2013) Rho kinase inhibitor fasudil regulates microglia polarization and function. *Neuroimmunomodulation* 20(6):313–322. <https://doi.org/10.1159/000351221>
- Zhang Q, Lü Y, Bian H, Guo L, Zhu H (2017) Activation of the  $\alpha 7$  nicotinic receptor promotes lipopolysaccharide-induced conversion of M1 microglia to M2. *PubMed* 9(3):971–985
- Zhang Z, Rong L, Li YP (2019b) Flaviviridae Viruses and oxidative stress: implications for viral pathogenesis. *Oxidative Med Cellular Longevity.* <https://doi.org/10.1155/2019/1409582>
- Zhang B, Wei Y-Z, Wang G-Q, Li D-D, Shi J-S, Zhang F (2019a) Targeting MAPK pathways by naringenin modulates microglia M1/M2 polarization in Lipopolysaccharide-stimulated cultures. *Front Cell Neurosci.* <https://doi.org/10.3389/fncel.2018.00531>

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