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High-Dose Resveratrol Supplementation in Obese Men
An Investigator-Initiated, Randomized, Placebo-Controlled Clinical Trial of Substrate Metabolism, Insulin Sensitivity, and Body Composition

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Management of obesity and its complications is a major health issue (1). Because primary preventive measures often fail and therapeutic options are insufficient, novel treatment modalities are being investigated (2). As such, the polyphenolic compound resveratrol has attracted attention over the past decades (3). It is widely distributed in nature and plays an important role in plants’ defenses against external stressors and infections; it also is a constituent of grapes and wine, albeit in minute amounts (on average, 1.9 mg/L in red wine) (4).

Preclinical trials suggest that resveratrol mimics the metabolic effects of calorie restriction (CR) (5) via activation of silent mating type information regulation 2 homolog 1 (SIRT1) (6–9). By means of NAD+-dependent deacetylase activity, SIRT1 modifies various target proteins, in particular transcription factors critical for energy metabolism, such as nuclear factor-κB, peroxisome proliferator-activated receptor γ coactivator 1-α, forkhead transcription factor class O, and sterol regulatory element-binding protein 1 (10,11).

In experimental animals, these effects translate into improved glucose metabolism (5,7,12–14), reduced inflammation (15–17), cancer prevention (18–21), reversal of nonalcoholic fatty liver disease (7,17,22–24), and prevention of obesity (8,25). In accordance with CR, supplementation with resveratrol promotes longevity in several primitive species (26–28) and protects against diet-induced metabolic abnormalities in rodents (7). Whether these effects of resveratrol apply to human beings remains uncertain, even though it is widely distributed as an over-the-counter nutritional supplement.

Only one trial systematically examining metabolic effects has been performed in humans: 11 obese male participants received a daily dose of 150 mg resveratrol or placebo for 30 days in a double-blind crossover design. Significant albeit moderate improvements in insulin sensitivity, blood pressure, metabolic rate, hepatic steatosis, and pertinent biomarkers were recorded (29). Apart from this single study, only a limited number of human clinical trials of efficacy outcomes have been conducted (30–32). In the present study we tested the impact of high-dose resveratrol administration for 4 weeks on energy and substrate metabolism, insulin sensitivity, ectopic fat disposition, 24-h ambulatory blood pressure, and inflammatory as well as metabolic biomarkers in obese human subjects.

RESEARCH DESIGN AND METHODS

Ethical approval. All participants were given oral and written information before written informed consent was obtained. The protocol was approved by the Regional Committee on Health Research Ethics and the Danish Data Protection Agency, and the study was conducted in agreement with the Declaration of Helsinki II. According to the International Committee of Medical Journal Editors, the protocol was registered at clinicaltrials.gov (NCT01150955) before recruitment was initiated.

Subjects. Twenty-four male volunteers, aged 18–70 years, participated. All participants were obese (BMI >30 kg/m²) but otherwise healthy, were taking no prescriptive medicine, and had no overt endocrine disorders. Eligibility criteria included a body mass index ≥30 kg/m², age 18–70 years, and a waist circumference >102 cm in men. All participants had to be in an energy-neutral or positive energy balance for at least 3 months before recruitment was initiated.

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See accompanying commentary, p. 1022.
ultimately was based on a normal physical examination including routine clinical biochemistry and electrocardiography.

**Study design.** The study was an investigator-initiated randomized, double-blinded, placebo-controlled, parallel-group trial. The subjects were treated for 4 weeks with tablets containing 500 mg trans-resveratrol (Flxume Inc., Stenlose, Denmark) or placebo (Robinson Pharma, Santa Ana, CA) thrice daily. Randomization, blinding, packaging, and labeling was performed by the pharmacy at Aarhus University Hospital. The randomization code was unblinded once all predefined data were recorded.

During the trial period, the subjects were instructed to abstain from using nutritional supplements and food suspected to contain resveratrol in significant amounts. Furthermore, the importance of maintaining their normal way of living was underscored. The compliance rate, defined as the proportion of tablets ingested relative to the intended number, was calculated when participants returned the remaining tablets during the last examination.

**Overall visits and interventions.** Examinations were performed on 3 consecutive days both at baseline and after 4 weeks of treatment with the same equipment and by the same physicians and laboratory technicians on both occasions. On the first day we performed dual-energy X-ray absorptiometry scan and initiated 24-h ambulatory blood pressure recordings. On the second day, we conducted magnetic resonance (MR) spectroscopy and imaging, and on the third day and after an overnight fast the participants underwent a full-day metabolic investigation including a hyperinsulinemic euglycemic clamp with continuous infusion of a primed glucose tracer, indirect calorimetry, repetitive blood sampling, and muscle and adipose tissue biopsies.

When completing the full-day metabolic investigation at baseline, the tablets were provided and the subjects were instructed to initiate tablet consumption in the evening and subsequently three times daily until the overnight fast before the third examination day at week 4. At weeks 2, 4, and 6, potential adverse events were recorded, and fasting blood samples were drawn for safety purposes. Urine from the first void was collected for pharmacokinetic purposes in the morning of the third examination day at week 4.

**Hyperinsulinemic euglycemic clamp.** After an overnight fast (from 10:00 p.m.), the participants presented to the research unit at 7.30 a.m. and were studied in the supine position during thermonutral conditions in the fasting state for 6 h.

For the purpose of infusions, a catheter (Venflon; Viggø, Helsingborg, Sweden) was inserted in an antecubital vein and included a heparin-bulled continuous euglycemic clamp with a continuous infusion of a primed glucose tracer, indirect calorimetry, repetitive blood sampling, and muscle and adipose tissue biopsies.

When completing the full-day metabolic investigation at baseline, the tablets were provided and the subjects were instructed to initiate tablet consumption in the evening and subsequently three times daily until the overnight fast before the third examination day at week 4.

**Muscle and adipose tissue biopsies.** Under sterile conditions and using local anesthesia (lidocaine 10 mg/mL; Amgros, Copenhagen, Denmark) skeletal muscle and adipose tissue biopsies were taken at the end of the basal period and 20 min into the clamp period (140 and 200 min, respectively). The muscle biopsy was obtained from the vastus lateralis muscle using a Bergstrom biopsy needle. The muscle tissue was immediately dissected free from fat and connective tissue and transferred to liquid nitrogen. Subcutaneous abdominal fat was obtained by liposuction 15 cm lateral to the umbilicus, cleaned, and subsequently snap-frozen in liquid nitrogen.

**[3-3H]-glucose tracer.** A primed continuous infusion of [3-3H]-glucose was given during the entire 6-h basal/clamp procedure (bolus 20 μCi followed by 0.19 μCi/min; NEN Life Science Products, Boston, MA). To avoid rapid dilution of [3-3H]-glucose during the hyperinsulinemic euglycemic clamp, [3-3H]-glucose was added to the infused glucose (100 μCi 3-[3H]-glucose/500 μl 20% glucose, specific activity 0.99 Ci/mmol; U.S. Radiopharmaceuticals, Overlea, MD). Specific activity of [3-3H]-glucose was measured (33), and glucose rate of appearance (Ra) and rate of disappearance (Rd) was calculated at 10-min intervals from 150 to 180 and 330 to 360 min using Steele’s non-steady-state equation (34).

**Indirect calorimetry.** The respiratory quotient (RQ) and resting energy expenditure (REE) were estimated by indirect calorimetry (Deltatrac; Datec-Ohmeda, Helsinki, Finland) performed at 90–120 min and 270–300 min in the basal and clamp periods, respectively. Mean values taken during the last 25 min were used for calculations. Glucose and lipid oxidation rates were estimated after correction for protein oxidation, which was calculated on the basis of urea nitrogen excretion (35). Urea nitrogen content was measured by absorption photometry (ROCHE cobas 6000C, Roche Applied Science, Penzberg, Germany).

**Dual-energy X-ray absorptiometry.** Whole-body bone mineral density and body composition were assessed by dual-energy X-ray absorptiometry (Hologic Discovery scanner SN 80027; Hologic Inc., Waltham, MA).

**Blood pressure.** Noninvasive, 24-h ambulatory blood pressure monitoring was performed based on measurements every 20th minute (Spacelabs, model 90217).

**Western blotting.** Frozen muscles biopsies (~30 mg) were homogenized in ice-cold solubilization buffer (43), and samples were rotated for 60 min at 4°C. Insoluble materials were removed by centrifugation at 16,000g for 20 min at 4°C, and the protein content of the protein-containing supernatant was determined.

Aliquots of protein were resolved by SDS-PAGE, transferred to polyvinylidene fluoride membranes, and incubated with primary antibody. All primary antibodies were from Cell Signaling (Beverly, MA). To determine relative phosphorylation levels, phospho-specific blots were stripped in SDS buffer and re-probed with the primary antibody for the detection. Horseradish peroxidase-conjugated goat anti-rabbit immunoglobulin G (Amersham, GE-Healthcare, Pittsburgh, PA) was used as the secondary antibody. Proteins were visualized by enhanced chemiluminescence (Pierce Supersignal West Dura; Thermo Scientific, Rockford, IL) using a ChemiDoc XRS+ CCD camera (BioRad, Hercules, CA).

**Real-time RT-PCR.** Total RNA was isolated from muscle and adipose tissue using TRIzol (Gibco BRL, Life Technologies, Rosskilde, Denmark); RNA was quantified by measuring absorbance at 260 and 280 nm with a ratio >1.8 using microplate photometer (M.M. POULSEN AND ASSOCIATES).
Table 1: Baseline characteristics

<table>
<thead>
<tr>
<th>Characteristics</th>
<th>Placebo</th>
<th>Resveratrol</th>
<th>P value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age (years)</td>
<td>31.9 ± 2.9</td>
<td>44.7 ± 3.5</td>
<td>0.01</td>
</tr>
<tr>
<td>Weight (kg)</td>
<td>115.9 ± 3.6</td>
<td>107.1 ± 2.7</td>
<td>0.005</td>
</tr>
<tr>
<td>BMI (kg/m²)</td>
<td>35.9 ± 1.2</td>
<td>32.5 ± 0.6</td>
<td>0.053</td>
</tr>
<tr>
<td>Lean mass (kg)</td>
<td>73.4 ± 1.8</td>
<td>69.1 ± 1.6</td>
<td>0.083</td>
</tr>
<tr>
<td>Fat mass (kg)</td>
<td>37.0 ± 2.6</td>
<td>31.7 ± 1.4</td>
<td>0.078</td>
</tr>
<tr>
<td>Fat (%)</td>
<td>32.3 ± 1.4</td>
<td>30.2 ± 0.7</td>
<td>0.37</td>
</tr>
<tr>
<td>Visceral fat volume (cm³)</td>
<td>4285 ± 585</td>
<td>4996 ± 876</td>
<td>0.51</td>
</tr>
<tr>
<td>Abdominal subcutaneous fat volume (cm³)</td>
<td>10737 ± 1086</td>
<td>8103 ± 585</td>
<td>0.051</td>
</tr>
<tr>
<td>Systolic blood pressure (mmHg)</td>
<td>126.1 ± 2.3</td>
<td>124.3 ± 2.9</td>
<td>0.64</td>
</tr>
<tr>
<td>Diastolic blood pressure (mmHg)</td>
<td>74.4 ± 1.6</td>
<td>75.6 ± 2.1</td>
<td>0.61</td>
</tr>
<tr>
<td>Glucose (mmol/L)</td>
<td>5.3 ± 0.1</td>
<td>5.6 ± 0.1</td>
<td>0.18</td>
</tr>
<tr>
<td>HbA1c (%)</td>
<td>5.6 ± 0.1</td>
<td>5.6 ± 0.1</td>
<td>0.55</td>
</tr>
<tr>
<td>Total cholesterol (mmol/L)</td>
<td>5.3 ± 0.4</td>
<td>5.5 ± 0.3</td>
<td>0.65</td>
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<tr>
<td>HDL cholesterol (mmol/L)</td>
<td>0.9 ± 0.1</td>
<td>1.0 ± 0.1</td>
<td>0.84</td>
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<tr>
<td>LDL cholesterol (mmol/L)</td>
<td>3.2 ± 0.2</td>
<td>3.6 ± 0.2</td>
<td>0.22</td>
</tr>
<tr>
<td>Triglycerides (mmol/L)</td>
<td>2.0 ± 0.3</td>
<td>2.0 ± 0.3</td>
<td>0.97</td>
</tr>
</tbody>
</table>

Twenty-four volunteers were randomly assigned 4 weeks of placebo (n = 12) or resveratrol (n = 12) treatment. Values are shown as means ± SEM.

Table 2: Plasma biochemistry

<table>
<thead>
<tr>
<th></th>
<th>Placebo</th>
<th>Resveratrol</th>
<th>P value*</th>
</tr>
</thead>
<tbody>
<tr>
<td>Glucose (mmol/L)</td>
<td>5.33 ± 0.08</td>
<td>5.33 ± 0.13</td>
<td>1.00</td>
</tr>
<tr>
<td>Insulin (pmol/L)</td>
<td>79.8 ± 10.4</td>
<td>79.9 ± 5.7</td>
<td>0.80</td>
</tr>
<tr>
<td>HOMA-IR index</td>
<td>3.17 ± 0.44</td>
<td>3.19 ± 0.27</td>
<td>0.82</td>
</tr>
<tr>
<td>HbA1c (%)</td>
<td>5.56 ± 0.05</td>
<td>5.42 ± 0.05</td>
<td>0.33</td>
</tr>
<tr>
<td>Cholesterol (mmol/L)</td>
<td>5.27 ± 0.35</td>
<td>5.13 ± 0.21</td>
<td>0.27</td>
</tr>
<tr>
<td>LDL (mg/dL)</td>
<td>0.93 ± 0.06</td>
<td>1.02 ± 0.06</td>
<td>0.13</td>
</tr>
<tr>
<td>Triglycerides (mg/dL)</td>
<td>3.19 ± 0.22</td>
<td>3.03 ± 0.17</td>
<td>0.22</td>
</tr>
<tr>
<td>Cholesteryl (mg/dL)</td>
<td>2.03 ± 0.33</td>
<td>2.18 ± 0.29</td>
<td>0.80</td>
</tr>
<tr>
<td>Leptin (mg/mL)</td>
<td>19.47 ± 3.95</td>
<td>18.00 ± 1.98</td>
<td>0.32</td>
</tr>
<tr>
<td>hsCRP (mg/mL)</td>
<td>2.86 ± 0.39</td>
<td>4.04 ± 1.01</td>
<td>0.03</td>
</tr>
<tr>
<td>IL-6 (pg/mL)</td>
<td>3.21 ± 0.96</td>
<td>3.59 ± 0.72</td>
<td>0.60</td>
</tr>
<tr>
<td>TNF-α (pg/mL)</td>
<td>4.69 ± 0.79</td>
<td>5.00 ± 1.70</td>
<td>0.82</td>
</tr>
<tr>
<td>MCP-1 (pg/mL)</td>
<td>140.1 ± 7.5</td>
<td>134.2 ± 8.4</td>
<td>0.13</td>
</tr>
<tr>
<td>Leukocytes (10⁹/L)</td>
<td>5.98 ± 0.45</td>
<td>6.71 ± 0.58</td>
<td>0.94</td>
</tr>
<tr>
<td>ALT (U/L)</td>
<td>48.5 ± 5.4</td>
<td>54.6 ± 6.7</td>
<td>0.10</td>
</tr>
</tbody>
</table>

Circulating parameters before and after 4 weeks' treatment with placebo (n = 12) or resveratrol (n = 12). All data are presented as means ± SEM. HOMA-IR, homeostasis model assessment–insulin resistance; hsCRP, high-sensitivity C-reactive protein; IL-6, interleukin 6; TNF, tumor necrosis factor; MCP-1, monocyte chemotactic protein 1; ALT, alanine aminotransferase. *P values reflect overall comparison of the groups by two-way repeated-measures ANOVA.
Glucose oxidation (mg/kg/min)
- Basal: 1.09 ± 0.11
- Clamp: 1.15 ± 0.12

Protein oxidation (mg/kg/min)
- Basal: 0.62 ± 0.08
- Clamp: 0.82 ± 0.09

Lipid oxidation (mg/kg/min)
- Basal: 0.58 ± 0.05
- Clamp: 0.44 ± 0.04*

Resting energy expenditure (kcal/24 h)
- Basal: 2143 ± 128
- Clamp: 2125 ± 135

Respiratory quotient (%)
- Basal: 0.83 ± 0.01
- Clamp: 0.85 ± 0.01

Insulin (pmol/L)
- Basal: 79.9 ± 10.4
- Clamp: 327.3 ± 16.2*

C-peptide (pmol/L)
- Basal: 1049 ± 68
- Clamp: 790 ± 73*

Glucagon (pg/mL)
- Basal: 71.8 ± 10.9
- Clamp: 30.3 ± 3.2*

Cortisol (ng/mL)
- Basal: 80.2 ± 14.0
- Clamp: 65.8 ± 7.9

Adiponectin (mg/L)
- Basal: 6.09 ± 0.65
- Clamp: 5.94 ± 0.64

FFA (mmol/L)
- Basal: 0.46 ± 0.04
- Clamp: 0.087 ± 0.011*

**RESULTS**

**Baseline and internal validity.** A total of 26 subjects were enrolled, but one participant dropped out because of claustrophobia in relation to MR imaging, and another developed a generalized rash after 1 week and was excluded (Supplementary Fig. 2). Both placebo and resveratrol tablets were generally well tolerated (Supplementary Table 2). The compliance rates, based on pill count, were 89.2 ± 2.9% and 88.9 ± 3.3% in the placebo and resveratrol groups, respectively. By chance the two groups differed slightly in mean age but were comparable on all other baseline parameters (Table 1).

To document systemic absorption of the investigational product, we did a pharmacokinetic pilot study before the clinical trial. In that pilot study, three healthy subjects ingested one resveratrol tablet (from the same batch as the investigational product) of 500 mg followed by plasma samples at 0, 15, 30, 45, 60, 90, 120, 180, 240, and 300 min. The native compound reached traceable amounts at 15 min, and Cmax was reached at 90 min with 300–400 ng/mL, which is consistent with the literature (Supplementary Fig. 3) (44–48) yet distinctly below the concentration usually described in in vitro studies (50 μmol/L ~ 11,400 ng/mL) (8). Furthermore, we compared the absorption profile of our investigational product (trans-resveratrol, Fluxome Inc., Stenlose, Denmark) with that of a similar product (500 mg resVida, DSM Nutritional Products, Ltd., Kaiseraugst, Switzerland). The two products exhibited similar absorption profiles (Supplementary Fig. 3).

The level of urinary resveratrol metabolites was determined at week 4, and no metabolites were detected in the placebo group, whereas measurable amounts were found in the resveratrol group (Supplementary Table 3).
Clinical biochemistry. We recorded no significant effects of resveratrol on HbA1c levels or the HOMA-IR. At baseline, the lipid profile was abnormal and compatible with obesity, but we observed no changes in total cholesterol, HDL, LDL, or triglycerides during the intervention period. Moreover, we did not record significant deviations in any inflammatory biomarkers, leptin, or liver function tests (Table 2).

Substrate metabolism. We did not record statistically significant changes in resting energy expenditure or respiratory quotient or in glucose or lipid oxidation rates when comparing basal and clamp values between the groups (Table 3). As expected, glucose oxidation increased during the clamp at the expense of reduced lipid oxidation. However, these changes did not differ either between or within the two groups (Table 3).

We also failed to detect significant differences in insulin, C-peptide, glucagon, cortisol, adiponectin, or FFAs between the groups. We recorded a significant within-group suppression of C-peptide, glucagon, and FFA levels during the clamp studies both before and after the intervention. No within-group changes in cortisol levels were found during insulin stimulation (Table 3).

Insulin sensitivity. Insulin sensitivity assessed by the hyperinsulinemic euglycemic clamp method was not significantly affected by resveratrol. Insulin levels increased from ~75 pmol/L in the basal period to ~315 pmol/L in the clamp period, with no significant differences between the groups regarding the overall basal and clamp periods (P = 0.66 and P = 0.07, respectively). As depicted in Fig. 1A and B, the GIR increased during the clamp and plateaued.
toward the end of the clamp. GIR did not differ between the groups, and the corresponding M value, defined as the mean GIR during the last 30 min of the clamp, also was comparable between the groups (Fig. 1C). As expected, insulin increased the glucose Rd approximately threefold in the clamp situation compared with the basal condition in all experimental settings, with no significant differences between the two groups (Fig. 1D). Likewise, EGP during both the basal state and the clamp period was comparable between the two groups (Fig. 1E). Finally, the rates of oxidative as well as nonoxidative glucose disposal did not differ between the groups (Fig. 1F).

FIG. 3. Body composition. Absolute changes in essential body composition parameters after 4 weeks of placebo (n = 12) or resveratrol supplementation (n = 12). The box boundaries indicate the 25th–75th percentile range, and error bars represent the 5th–95th percentiles. Solid and dotted lines represent median and mean, respectively. A–D were assessed by whole-body dual-energy X-ray absorptiometry; E and F were quantified by MR imaging based on repetitive axial slices from the proximal border of the left kidney to the femur neck; nine were evaluable in each group. Two-way repeated-measures ANOVA revealed no statistically significant differences between groups.
Body composition. We did not demonstrate any significant differences in total body mass, lean body mass, total body fat mass, or visceral and abdominal subcutaneous fat volumes (Fig. 3).

Liver and skeletal muscle lipid content. Even though considerable hepatic fat infiltration was evident at baseline, no significant changes occurred in either group during the trial. The same was true when analyzing the intramyocellular lipid content (Fig. 4).

Gene expression and protein phosphorylation. Activation of AMP-activated protein kinase (AMPK) in the muscle tissue biopsies by phosphorylation of the Thr172 residue of the catalytic α-subunit was not impacted by resveratrol. Consistent with this, phosphorylation of acetyl-CoA carboxylase, a well-known downstream target of AMPK activity, also was not affected by the intervention. Furthermore, we explored the proposed SIRT1-mediated deacetylase activity potentially induced by resveratrol by assessing the total acetylation status of lysine residues, which also was not affected (Fig. 5 and Supplementary Fig. 4).

Apart from a slightly decreased insulin-stimulated expression of GLUT4 mRNA in muscle in the resveratrol group, gene transcription in muscle and fat did not change in any groups (Fig. 5 and Supplementary Fig. 4).

DISCUSSION

The main finding of our study is that short-term (4 weeks) supplementation with high-dose resveratrol is not associated with detectable physiological effects in obese subjects with modest insulin resistance. We consider this trial important for several reasons. First, the high incidence of obesity and type 2 diabetes calls for novel preventive and therapeutic modalities. Second, substantial evidence from preclinical work suggests that resveratrol stimulates SIRT1 activity and suppresses inflammatory pathways in human adipose tissue in vitro (49,50). Third, the compound is already widely available as an over-the-counter nutritional supplement with an array of alleged salutary effects. Fourth, resveratrol treatment may serve as a model to investigate potential biomarkers for CR and SIRT1 in human models.

Our primary outcome, insulin sensitivity, which was assessed by the hyperinsulenic euglycemic clamp, was not significantly affected by resveratrol. The same was true for HOMA-IR, fasting glucose, and fasting insulin. Our negative findings are in conflict with the work by Timmers et al. (29), who demonstrated statistically significant improvements in HOMA-IR, suggesting a favorable effect on insulin sensitivity. However, the gold standard for the assessment of whole-body insulin sensitivity is the hyperinsulinemic euglycemic clamp technique used in our study. Furthermore, our inclusion of glucose tracer allowed for the determination of glucose turnover in the basal state and EGP during the clamp, all of which also failed to be affected by resveratrol. We also found no effects of resveratrol on either resting energy expenditure or rates of lipid oxidation.

When assessing blood pressure, we found a trend toward a moderate increase in 24-h blood pressure after resveratrol administration compared with placebo. Again, the differences were insignificant and in opposition to the findings by Timmers et al. (29), who recorded a robust decrease in both systolic and diastolic blood pressures, which were assessed by the mean of conventional clinical blood pressure triplicates. Furthermore, we examined the ectopic fat content in hepatic and muscle tissue by MR spectroscopy and, in contrast to Timmers et al., we recorded no effects of resveratrol. In preclinical settings, the finding of diminished hepatic fat induced by resveratrol is consistent, and the reason why we fail to reproduce the findings by Timmers et al. could relate to baseline differences. Because lipid content is expressed as a relative number relative to the water content, we cannot directly compare the data sets. In accordance with the ectopic fat content, we also failed to detect changes in adipose tissue assessed by MR imaging, which perhaps is less surprising considering the relatively short duration of treatment.

Consistent with the lack of physiological responses to high-dose resveratrol, we also did not find any alterations in gene expression of a panel of pivotal metabolic and

**FIG. 4. Ectopic lipid.** Relative changes in ectopic lipid deposition from baseline after 4 weeks of placebo or resveratrol supplementation, respectively. Results are generated by 1H-MR spectroscopy and raw data processing provides an estimate of the ratio of lipid to water (LWR) within the tissue. The box boundaries indicate the 25th–75th percentile range, and error bars represent the 5th–95th percentiles. Solid and dotted lines represent median and mean, respectively. Two-way repeated-measures ANOVA revealed no statistically significant differences between groups. A: Changes in hepatic lipid content; 11 were evaluable in each group. B: Changes in intramyocellular lipid (IMCL) content; 10 were evaluable in each group.
FIG. 5. Gene expression and protein phosphorylation. Intracellular protein levels and relative mRNA expression in muscle (A–D) and adipose (E and F) tissue biopsies taken before and after 4 weeks’ treatment with placebo (n = 12) or resveratrol (n = 12). Biopsies were taken before and during (30 min after initiation) a hyperinsulinemic euglycemic clamp. Black bars indicate the placebo group and white bars indicate the resveratrol group. Results are presented as group means ± SEM, and overall comparisons of potential treatment effects were performed by two-way repeated-measures ANOVA in the basal and clamp situations, respectively. A: Phosphorylation of the intracellular kinase AMP-activated protein kinase (AMPK) assessed by Western blot analysis in muscle tissue. B: Total acetylation of lysine residues assessed by Western blot analysis in muscle tissue. C: Relative GLUT4 mRNA expression in muscle tissue assessed by RT-PCR. D: Relative peroxisome proliferator-activated receptor γ coactivator 1-α (PGC1α) mRNA expression in muscle tissue assessed by RT-PCR. E: Relative tumor necrosis factor (TNF)-α mRNA expression in subcutaneous adipose tissue assessed by RT-PCR. F: Relative nuclear factor (NF)-κB mRNA expression in subcutaneous adipose tissue assessed by RT-PCR. *In C, we found an overall treatment effect in the clamp situation, and a post hoc test revealed a statistically significant (P < 0.05) decreased expression of GLUT4 in the resveratrol group.
inflammatory biomarkers. Nor were protein phosphorylation levels of AMPK and acetyl-CoA carboxylase or total acetylation status affected. As mentioned, our data disagree with the recently published work by Timmers et al. (29), who recorded significant effects using only one-tenth of our dose in a group of obese men. In comparison with the participants in the study by Timmers et al., ours were slightly younger (age 38.4 ± 2.6 vs. 52.5 ± 2.1 years) and more obese (BMI 34.2 ± 0.7 vs. 31.59 ± 0.7 kg/m²; fat 31.3 ± 0.8 vs. 26.4 ± 0.5%). To what degree age and extent of obesity may impact the effect of resveratrol in human subjects remains to be investigated. We have scrutinized our design to identify potential design-related problems to explain the lack of effect. We have demonstrated that the absorption of our resveratrol formulation is comparable to the absorption of the compound used in the study by Timmers et al. Our study groups were well matched and of a reasonable sample size, the compliance was good, and we consider the duration of treatment sufficient. The difference in age between our two treatment groups was due to two outliers in the resveratrol group, aged 60 and 68 years. We have repeated all our analysis without these two outliers, which did not change the negative outcome (data not shown). Finally, the risk of a type 2 error due to low sample size should also be considered. In the study by Timmers et al., a sample size of 11 was sufficient to demonstrate effects in a cross-over design. Compared with that study, ours holds the potential advantage of a parallel design, which eliminates the risk of untoward seasonal changes and carry-over effects. A limitation of our study is the inability to directly demonstrate the presence of resveratrol and metabolites in plasma. However, we demonstrated the absorption and excretion of the investigational product by means of our pharmacokinetic pilot study and urinary metabolite measurements.

Based on solid preclinical evidence, it is likely that a certain degree of baseline metabolic abnormalities is a prerequisite to benefit from resveratrol treatment, and it could be speculated that our participants were “too healthy.” However, both HOMA and M values at baseline may indicate mild insulin resistance in our cohort: a HOMA value of 2.77 previously has been defined as the threshold for insulin resistance in metabolically healthy subjects (51). Moreover, in a group of lean but slightly younger men (BMI, 24.6 ± 3.2 kg/m²; age 23.0 ± 2.0 years) clamped at an identical insulin dose and with the same procedures as those used in our laboratory, we recorded an M value of 5.8 mg/kg/min after 3 h (52) compared with 2.5 mg/kg/min in the current study, which suggests a ~60% reduction in insulin sensitivity in our participants. Nonetheless, in future studies, and before making any definitive conclusions, the therapeutic potential of resveratrol should be tested in patients with more pronounced morbidity such as type 2 diabetes, nonalcoholic fatty liver disease, and hypertension.

Little is known about the pharmacodynamic properties of resveratrol. Even though the preclinical evidence is quite substantial, basic mechanisms of action are not elucidated or agreed upon. SIRT1 seems to be a pivotal mediator of the metabolic effects of resveratrol, but the up- and downstream mechanisms are not fully understood. One of the main questions pending is whether the effects depend on AMPK and, if so, whether AMPK activation is up- or downstream of SIRT1. Regardless of our inability to demonstrate AMPK activation, resveratrol may have potential targets in addition to AMPK and SIRT1. Emerging evidence suggests certain pathways are affected differently at different dose levels, with subsequent effects on physiological outcomes, both in vitro (53) and in vivo (22). In fact, to our knowledge, the latter work is the first to report an inverse dose-response relationship in vivo; more pronounced effects apparently are observed at lower doses. Even though an inverted or J-shaped dose-response relationship seems to be an unlikely explanation for our results, this remains a possibility. Future studies should focus on dose-response relationships and preferably also compare the impact of treatment duration, including potential acute effects of resveratrol. It also remains to be reported whether alternative small-molecule activators of SIRT1 (STACs) may be more effective compared with resveratrol in clinical settings. Regardless of our findings, the scientific field of sirtuin biology remains an area of great interest.

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M.M.P., N.M., N.J., S.B.P., and J.O.L.J. conceived of and designed the study. P.F.V. and H.S.-J. performed MR scans and processed the raw MR data. B.F.C. performed Western blotting analyses. Y.R. and L.P.C. performed plasma and urine pharmacokinetic analyses. M.M.P. performed all other investigations and analyses and further analyzed data. M.M.P. and J.O.L.J. wrote the manuscript, which was reviewed by all authors. M.M.P. is the guarantor of this work and, as such, had full access to all the data in the study and takes responsibility for the integrity of the data and the accuracy of the data analysis.

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