Novel Rat Model of Repetitive Portal Venous Embolization Mimicking Human Non-Cirrhotic Idiopathic Portal Hypertension

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Abstract

Background
Non-cirrhotic idiopathic portal hypertension (NCIPH) is characterized by splenomegaly, anemia and portal hypertension, while liver function is preserved. However, no animal models have been established yet. This study assessed a rat model of NCIPH and characterized the hemodynamics, and compared it to human NCIPH.

Methods
Portal pressure (PP) was measured invasively and coloured microspheres were injected in the ileocecal vein in rats. This procedure was performed weekly for 3 weeks (weekly embolization). Rats without and with single embolization served as controls. After four weeks (one week after last embolization), hemodynamics were investigated, hepatic fibrosis and accumulation of myofibroblasts were analysed. General characteristics, laboratory analyses and liver histology were collected in patients with NCIPH.

Results
Weekly embolization induced a hyperdynamic circulation, with increased PP. The mesenteric flow and hepatic hydroxyproline content was significantly higher in weekly embolized compared to single embolized rats (mesenteric flow +54.1%, hydroxyproline +41.7%). Mesenteric blood flow and shunt volumes increased, whereas splanchnic vascular resistance was decreased in the weekly embolization group. Fibrotic markers αSMA and Desmin were upregulated in weekly embolized rats.

Discussion
This study establishes a model using repetitive embolization via portal veins, comparable with human NCIPH and may serve to test new therapies.
Introduction

Non-cirrhotic idiopathic portal hypertension (NCIPH) is a poorly understood disease of varied etiology, which leads to portal hypertension and its complications. In the absence of cirrhosis, other synonyms are non-cirrhotic portal fibrosis (NCPF), or hepatoportal sclerosis and obliterative venopathy in Eastern countries [1]. Importantly, this condition occurs in the absence of other well characterized causes of portal hypertension such as chronic liver disease and splanchic venous thrombosis [2–5]. The prognosis of this disease is considered as benign with a 5 year survival of almost 100%, but long term outcomes are still unknown [4,6,7]. Since the prevalence of NCIPH in the Western world is low, recently published literature is rare [8]. Various conditions such as chronic infectious diseases, toxin or drug exposure, genetic and immunological disorders have been determined as potential causes of NCIPH [3,6].

Usually the first symptoms of NCIPH are splenomegaly and pancytopenia [1,8]. Gastro-oesophageal varices and bleeding complications remain a major burden for NCIPH-patients. Therefore, prophylaxis and treatment of variceal bleeding are of clinical relevance [4,7].

However, to test new treatment options, experimental models are still missing. One reason for the absence of experimental models is that the underlying pathomechanisms of NCIPH have been poorly understood so far [3,9,10]. In liver samples of patients with NCIPH, vascular lesions in the portal venules have been observed [3]. Micro-thrombotic lesions in small portal venules have been proposed as the initial cause of NCIPH [10].

Therefore, we studied the role of micro-thrombotic lesions as a possible initial step for the processes leading to NCIPH in a new rat model using microsphere embolization and compared the findings with human NCIPH.

Material and Methods

Patients

Five patients with portal hypertension and absence of other causes of hepatic fibrosis or cirrhosis were investigated in the study in the Department of Internal Medicine I, University of Bonn, Germany. Their histology was compared to five non-cirrhotic liver samples without portal hypertension. The use of human liver samples was approved by the Human Ethics Committee of the University of Bonn (Ethikkommission der Universität Bonn, reference number 029/13). It was not possible to obtain written or oral consent, because patients data and pathological specimen were analyzed retrospectively using remnant tissue specimens. Also patients had been lost for follow up and could no longer be contacted. In line with our ethical regulations it is allowed to use retrospective data in anonymous form, when the patients can no longer be contacted. The recruitment period was from 2009 to 2014 and routine laboratory values from baseline and follow up were observed. Biochemical blood analyses were performed using standard tests. A liver biopsy was taken from all patients and evaluated by two senior pathologists.

Ultrasound examination. Ultrasound examination was performed in all patients by senior internists applying current standards. Ultrasound included evaluation of liver parenchyma (fibrotic, nodular regenerative hyperplasia (NRH) and atrophic; hyperchoic and hypoechoic; homogeneous and heterogeneous), evaluation of liver vascularization (normal and abnormal; downsized and enlarged; regular limited and irregular limited), the evaluation of biliary ducts (enlarged and downsized) and the portal vein (obstructed, unobstructed).

Sirius red staining and αSMA immunohistochemistry of human biopsies. For the detection of collagen fibers, liver specimen were fixed in 10% formalin, paraffin-embedded and
stained in 0.1% Sirius-red in saturated picric acid (Chroma, Münster, Germany) using standard methods as previously described [11].

For immunohistochemical (IHC) staining of α-smooth muscle actin (αSMA), slides with sections were incubated with a mouse-anti-αSMA (clone 1A4; Sigma–Aldrich, St. Louis, USA) diluted 1:100 in Tris–buffered saline overnight. A secondary biotinylated rabbit-anti-mouse antibody, absorbed with rat serum (Dako, Glostrup, Denmark), was subsequently applied (1:300, 30 min) and complexed with streptavidin-conjugated alkaline phosphatase (1:500, 30 min; Dako). Finally, slides were developed with AEC (3-amino-9-ethylcarbazole) (15 min; Dako) and counterstained with hematoxylin. The amount of staining was evaluated by computational analysis (Histoquant; 3DHistech, Budapest, Hungary). Quantification (% of stained area) of IHC staining is expressed as mean±SEM.

**Human CD105 staining.** For immunohistochemical staining of hepatic CD105, liver specimens were fixed in 10% formalin, paraffin-embedded and stained with goat anti-human endoglin/CD105 (R&D Systems, Inc. Canada) at 4°C overnight. Tissue was stained using anti-goat HRP-DAB and counterstained with hematoxylin. Staining was evaluated by Pannoramic Viewer (Histoquant; 3DHistech, Budapest, Hungary) and quantification (% of stained area) of IHC staining is expressed as mean±SEM.

**Animals**

We used 23 Sprague-Dawley rats for our experiments. Six week-old male Sprague-Dawley rats were housed in a 12:12 h light-dark cycle, with controlled temperature (21°C ± 2°C), and ad libitum standard rat chow and water. The responsible committee for animal studies in North Rhine-Westphalia approved the study (Landesamt für Natur und Umwelt, LANUV 84–02.04.2014.A137).

**Assessment of hepatic fibrosis.** In corresponding segments (200mg) of snap-frozen rat livers, the hepatic hydroxyproline content was determined photometrically as described previously [11,12].

**Hepatic αSMA staining in rat.** Immunohistochemical stainings for hepatic αSMA were prepared using a cryostat. Cryo sections (4–6μm) of snap-frozen liver samples were fixed and incubated with mouse-anti-αSMA antibody (Sigma Aldrich, München, Germany). Thereafter, a biotinylated donkey-anti-rat secondary antibody was used (Abcam, Cambridge, UK). Sections were detected and quantified using computerized image capture device (Histoquant; 3DHistech, Budapest, Hungary). Results are expressed as mean±SEM.

**Protein expression measurement.** Liver pieces of all rats (weekly embolization, single embolization and control) were collected after performing the microsphere technique and snap-frozen. Western blot analyzes were performed with rabbit anti-αSMA antibody (Sigma Aldrich, Germany) and for Desmin with rabbit anti-Desmin antibody (GeneTex Inc., Irvine, CA, USA). Thereafter, the membranes were incubated with corresponding secondary peroxidase-coupled antibodies (Calbiochem, San Diego, USA). The housekeeping gene GADPH was used as loading control. Blots were developed with enhanced chemiluminescence. Intensities of the digitally detected bands were evaluated densitometrically using Chemi-Smart, normalized to control rat livers and represented as mean±SEM.

**Portal pressure measurements and induction of portal hypertension in rats.** Rats were anaesthetized by intraperitoneal (i. p.) injection of ketamine/xylazine (78 mg/kg / 12.5 mg/kg). First a laparatomy was performed, a catheter was inserted in an ileocecal vein and pushed to the portal vein to measure the PP. After PP measurement, weekly embolized rats (n = 5) received every week 150,000 coloured microspheres (violet (week 1), blue (week 2), yellow (week 3); 15μm diameter, Triton-Technologies, San Diego, USA) via the catheter in the
ileocecal vein. The single embolized rat group (n = 8) received 150,000 microspheres (blue) once after the first PP measurement. The control group (n = 10) did not receive any microspheres after PP measurements. After weekly PP measurements ileocecal veins were ligated and the linea alba and skin were closed with 4–0 Vicryl and 3–0 Prolene, respectively. All surgeries were performed under aseptic conditions. To prevent pain after assessment of the PP and microsphere injection rats received Carprofen (5mg/kg/d); s. c. for at least 5 days as described previously [13]. The early/human endpoints were defined the following characteristics of the rats: Immobility, isolation from other rats, walking on tiptoe or crookedly, scrubby fur, even breathing, to malnourished, insufficient liquid intake, body weight and the percent to the initial body weight, body temperature, inflamed eyes or interface, swollen abdomen, tremulousness and diarrhea. Stockmen score each characteristic for each rat every day. At a defined scoring point level rats will be euthanized immediately by carbon dioxide intoxication. The direct euthanasia is performed when rats lose 20% of their initial body weight, if the rats is unable to move, if the rat has no liquid intake anymore, if the rat is cachectic or if the feces is fluid. In the experimental group of “weekly embolized rats” 50% of the rats were euthanized prior the experimental endpoint due to weight lost and inflamed interfaces. The same reasons caused the euthanasia of 20% in the “single embolization” group before the experimental endpoint. In the control group all rats reached the experimental endpoint, as expected.

**Portal and systemic hemodynamic assessment.** Hemodynamic studies were performed under ketamine/xylazine anesthesia (78mg/kg / 12.5mg/kg i.p.) as previously described [14–17]. Rats were fasted overnight but allowed free access to water. Median laparotomy was performed; a PE-50 catheter was introduced into a small ileocecal vein and advanced to the portal vein for the measurement of PP. The left femoral artery was cannulated with a PE-50 catheter for measurement of the mean arterial pressure (MAP) and blood withdrawal. Via the right carotid artery, another PE-50 catheter was advanced into the left ventricle under pulse curve control. This catheter was used for microsphere application to investigate the cardiac output. The catheters in the femoral artery and the portal vein were connected to a pressure transducer (ADInstruments Ltd, Oxford, United Kingdom) for blood pressure measurement. The zero point was 1 cm above the operating table. After insertion of all catheters, rats were allowed to stabilize hemodynamically for 30 min [14].

**Microsphere technique.** Hemodynamics were investigated using the coloured microsphere technique as previously described [14–17]. The colored microsphere technique was validated by the more frequently used radioactive microsphere method [18]. It has the advantage of being non-radioactive and using different colors at the same organism. This technique has been shown very accurate in the measurement of hemodynamics in many studies of our group and others [14–16,18–23]. A reference sample was obtained for 1 min at a rate of 0.65mL/min, using a continuous withdrawal pump (Hugo-Sachs-Elektronik, March–Hugsten, Germany). 300,000 systemic (yellow) microspheres were suspended in 0.3mL saline containing 0.05% Tween and injected in the left ventricle 10 sec after the withdrawal pump had been started. Mesenteric portal-systemic shunt volume was estimated after injection of 150,000 white microspheres in 0.3 ml saline containing 0.05% Tween in an ileocecal vein within 30 seconds [24]. At the end of hemodynamic measurements the catheter in the ventricle is used to inject an overdose of ketamine (1ml of 10%) which euthanizes rats immediately without being afflicted with pain. The blood reference probe was digested by addition of 3.8 ml 5.3 M KOH and 0.5 ml Tween 80 and subsequent boiling for 1 hour. The digested tissues and blood samples were vortexed and filtered using Whatman Nucleopore filters (Whatman International Ltd., Madison, UK). The colour of the filtered microspheres was dissolved in 0.2 ml dimethyl-formamide, and the absorption was measured using spectrophotometry. Splanchnic vascular resistance was calculated from the ratio between splanchnic perfusion...
pressure and splanchnic blood flow, without including hepatic arterial flow. Mesenteric shunt flow was measured as the fraction of white microspheres in the lung from total white microspheres injected in an ileocolic vein.

**Statistical analysis.** Clinical data were collected and evaluated with SPSS statistical analysis software (IBM SPSS Statistics for Windows, Version 22.0, released 2013. Armonk, NY: IBM Corp.). Data are presented as median, maximum and minimum and mean±SEM. Data of laboratory values at the point of first contact were compared with data at the point of follow up in all groups. In all experimental animals the Wilcoxon test was used for comparison within one group. Unpaired data were compared by the Mann-Whitney test. Data are presented as mean ±SEM. P values <0.05 were considered statistically significant.

**Results**

**Clinical characteristics of NCIPH patients**

All five examined NCIPH patients were between 21 and 72 years old at baseline with a MELD-score between 6 and 16. All patients had oesophageal varices as a sign of portal hypertension.

Doppler-Sonography showed a patent portal vein in all patients. The liver parenchyma was described as hyperechoic and heterogeneous in four patients. Hepatic veins were irregular limited in five patients and of smaller size in three patients.

**Human liver histology**

None of the five liver biopsy specimens of humans had features of cirrhosis. The most common findings were fibrosis in periportal and perisinusoidal areas in two patients (40%). Three patients (60%) showed slight circulatory disturbances of liver perfusion. One patient (20%) had signs of atrophy and four patients (80%) had hyperplasia. Two of the NCIPH patients (40%) showed NPH and signs of abnormal vascularization (Table 1).

**Human hepatic Sirius red stainings**

In the specimens of human NCIPH and controls Sirius red staining was performed to access collagen accumulation. Increased Sirius red staining was observed in NCIPH patients around the portal tract compared to healthy control liver specimens (Fig 1a).

**Table 1. General characteristics of the patients with NCIPH.** General characteristics of the patients have been investigated in all patients and are shown in all patients. Characteristics of the parenchyma and of the liver have been evaluated by senior pathologists in specimens of the liver biopsies.

<table>
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</tr>
<tr>
<td>Biopsy (no/yes)</td>
<td>0 / 5</td>
</tr>
<tr>
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<td>3 / 2</td>
</tr>
<tr>
<td>Abnormal vascularisation (no/yes)</td>
<td>2 / 3</td>
</tr>
<tr>
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<td>4 / 1</td>
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<td>NRH (nodular regenerative hyperplasia) (no/yes)</td>
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Human hepatic αSMA stainings

NCIPH patients showed increased hepatic αSMA staining within portal tracts compared to control liver specimens (Fig 1b).

Human hepatic CD105 staining

The expression of the endothelial marker CD105, also called endoglin, which is involved in angiogenesis and hypoxia, was upregulated in livers of NCIPH patients, shown by immunohistochemically stainings (Fig 1c).
Quantification of human hepatic stainings

Quantifications of all human hepatic stainings were performed using computerized image capture (Histoquant; 3DHistech, Budapest, Hungary). Hepatic stainings of NCIPH patients were compared with control stainings and expressed as % of stained area. Stainings of αSMA, Sirius red and CD105 were significantly increased in NCIPH liver specimens compared to controls (Fig 1d).

Rationale of the animal models to mimic human NCIPH

We investigated three groups of rats: In weekly embolized rats PP was measured every week for three weeks prior microsphere embolization. PP of single embolized rats was measured invasively every week, but rats were only embolized with microspheres after the first PP measurement. In control rats PP was measured every week for three times but no microsphere injection was performed. At the end of experiments the PP and the MAP were measured invasively and the coloured microsphere technique was performed in all groups of rats to investigate hemodynamics (Fig 2a).

Hepatic hydroxyproline content

Collagen amount in livers was investigated by the hydroxyproline content. In weekly embolized rats, significantly higher hepatic hydroxyproline content was measured compared to rats after single embolization or to control rats. The single embolization of microspheres in the ileocecal vein did not increase the hepatic hydroxyproline content compared to control rats (Fig 2b).

Hepatic detection of microspheres and hepatic αSMA stainings

At the end of experiments livers were stained with αSMA to detect activated hepatic stellate cells (HSC) which promote portal hypertension. The microspheres could be detected in hepatic specimens of single and weekly embolized rats (Fig 2c, red circles). Especially signs of atrophy and hyperplasia were observed in the vicinity of microspheres. Increased αSMA stainings and subsequently a higher grade of HSC and myofibroblast activity was detected in weekly embolized rats than in single embolized or in control rats (Fig 2c and 2d).

Hepatic αSMA and Desmin protein expressions in NCIPH rats

The increased HSC and myofibroblast activity in weekly embolized rats was confirmed by western blot analysis of hepatic αSMA and hepatic Desmin expression levels, as an additional marker for HSC. Even after a single embolization rats expressed significantly more hepatic αSMA and Desmin than control rats. In weekly embolized rats the hepatic protein expression levels of αSMA and Desmin were significantly higher than in control or single embolized rats (Fig 2e).

Development of portal hypertension in NCIPH rats

Every week the PP was measured in all rat groups. After the embolization of microspheres, the PP increased continuously every week (weekly embolization group). After a single embolization of microspheres the PP did not change (single embolization group). In control rats, which were not embolized with microspheres, the PP did not change (Fig 3a).
Fig 2. Animal models. 

a) Experimental setup of animal models to mimic human NCIPH. The experimental setup of weekly embolized, single embolized and control rats is shown as timeline from start of experiments until the end of experiments after 3 additional weeks. PP measurements are marked as thin arrows and were always performed before embolization, marked by dots. Hemodynamic assessments at the end of experiments are marked as bold arrows.

b) Hepatic hydroxyproline content. The hydroxyproline content was evaluated in rat livers of all groups at the end of experiment. Livers of weekly embolized rats showed increased hydroxyproline content compared to single embolized and control rats. The results are shown as hydroxyproline content in μg/g liver.

c) Hepatic αSMA stainings. After αSMA stainings of

d) Quantification of hepatic αSMA stainings in rats

e) Hepatic αSMA and Desmin protein expressions in NCIPH rats

The hydroxyproline content was evaluated in rat livers of all groups at the end of experiment. Livers of weekly embolized rats showed increased hydroxyproline content compared to single embolized and control rats. The results are shown as hydroxyproline content in μg/g liver. c) Hepatic αSMA stainings. After αSMA stainings of
Portal pressure in NCIPH rats at end of experiments

At the end of experiments, after 3 weeks, the PP was significantly increased in weekly embolized rats compared to single embolized rats (Fig 3b). The mesenteric blood flow was significantly increased after weekly embolization compared to single embolized and control rats (Fig 3c). Also the mesenteric shunt volume was significantly increased after weekly embolization compared to control rats (Fig 3d). Splanchnic vascular resistance was significantly decreased in weekly embolized rats compared to rats with only a single embolization (Fig 3e). This is in line with hemodynamic findings in portal hypertension [16,25]. There were no significant differences in the MAP (weekly embolization: 87.0 ± 13.2; single embolization: 96.2 ± 12.2; control: 91.0 ± 6.9) and in the cardiac output (weekly embolization: 19.0 ± 1.6; single embolization: 29.8 ± 5.3; control: 16.7 ± 4.1), respectively.

Comparison of human and rat data

In summary the features that we find in humans could be reproduced in our new model of NCIPH (Table 2).

Discussion

The present study is the first study to offer an animal model of NCIPH, which compares the findings in rats with the findings in patients with NCIPH.

NCIPH is a rare disorder and causes substantial complications such as gastro-abdominal bleeding [7]. Causes and pathophysiology of NCIPH are still unknown [4]. However, various possible associations, e.g. micro-thrombotic lesions, have been described so far.

A distinctive histopathological feature of NCIPH is obliteration of small portal venules (occlusive venopathy) that may lead to impairment in the intrahepatic portal perfusion resulting parenchymal atrophy, marked by decreased size of cytoplasm and condensed nucleus this was clearly visible in HE stainings. Nodular regenerative hyperplasia was defined by micronodular transformation of the liver parenchyma, with central hyperplasia, an atrophic rim, and no fibrosis. Indeed, in specimens of liver biopsies, a major number of fibrotic processes and abnormal vascularization were shown in our patients, which could be reproduced in our animal models (presence of portal hypertension, fibrosis, activation of myofibroblast, atrophy or hyperplasia).

Repetitive in vivo thrombotic events have been proposed as a potential cause for the development of NCIPH. This occlusion by microspheres might serve as a model in animals. In line with this hypothesis, one of our patients had been treated with antiplatelet medication and showed an important improvement in bilirubin, creatinine and platelet count in follow up.

Indeed, using repetitive weekly embolization, PP increased significantly at each time point of embolization in rats. This experiment confirms that our animal model can reproduce the
Fig 3. Portal and systemic hemodynamic assessment and the NCIPH model. a) Portal pressure. Portal pressure was taken every week in weekly embolized and in control rats. In single embolized rats, the PP was taken at the beginning and the end of experiment. Portal pressures are shown in mmHg. Significant differences are evaluated using paired t-Test within the same group. b) Portal pressure at the end of experiments. At the end of experiments, the portal pressures were measured invasively in all rats. After weekly embolization, the PP was significantly higher compared to rats after a single embolization. The portal pressures are shown in mmHg. The significant difference to single embolized rats is evaluated using the nonparametric Mann-Whitney test. c) Mesenteric blood flow. The mesenteric blood flow was investigated at the end of experiment. The results are shown in ml/min/100g/kg body weight. The mesenteric blood flow...
proposed pathogenesis of NCIPH by micro-thrombotic insults. As expected, portal hyperten-
sion induced by repetitive embolization in the portal vein was combined with increased mesen-
teric blood flow and mesenteric shunt volume, suggesting that the full picture of portal
hypertension with hyperdynamic circulation was developed. Also, liver histology and intrahe-
patic fibrosis showed, that our model correctly reflects HSC activation and discrete perportal
fibrosis accumulation, which has been observed in patients with NCIPH [3], and were dis-
cussed as an important pathogenic stress for the development of NCIPH [10]. Therefore, we
offer a model for the development of NCIPH, which may help to test strategies for the
improvement of portal hypertension and complications of NCIPH.

Histological changes observed in our animal model nicely correspond to changes found in
human livers of patients with NCIPH, as described previously [2,3]. Importantly, in our
patients liver biopsy could confirm the presence of fibrosis accumulation and HSC activation,
which was induced by repeated micro-thrombotic events in the animal model.

The main difference between this model and other models of portal hypertension is that in
the majority of the models liver cirrhosis is induced to elicit portal hypertension, while in this
model liver does not exhibit cirrhosis. Besides immunological disorders, chronic infections,
exposures to medications or toxins and genetic disorders, prothrombotic conditions also lead
to NCIPH. NCIPH patients have a high incidence of portal vein thrombosis. A model of portal
vein thrombosis with consecutive portal hypertension is Partial Portal Vein Ligation (PPVL),
which is not eligible for our study, since portal vein thrombosis is not the reason but the conse-
quence of NCIPH.

Certainly the rat model has its limitations. First, it is a technical challenge to perform inva-
sive PP measurements and embolization in vivo. Another potential limitation of this model is
that patients with NCIPH are heterogeneous concerning their underlying etiology, which com-
prises immunological disorders or HIV. In contrast, NCIPH in the rats was induced by surgery
and iatrogenic embolization. Despite such short comings, the good correspondence of histolog-
ical findings between patients with NCIPH and rats as well as induction of portal hypertension
in the absence of cirrhosis suggests that repeated experimental micro-embolization at least
reproduce key findings of NCIPH. Of note, our results suggest that a single thrombotic event is
not sufficient but repeated portal venous micro-thrombotic events are required to induce

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<td>Presence of portal hypertension</td>
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<td>+</td>
</tr>
<tr>
<td>Fibrosis</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td>Activation of myofibroblast</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td>Atrophia / Hyperplasia</td>
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NCIPH. Thus, our model may allow to study therapeutic interventions of various stages of the pathogenic process ultimately resulting in NCIPH.

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Investigation: SK CH KH RS FU JT.
Methodology: SK CH KH RS FU JT.
Project administration: US JT.
Resources: HPF CPS US JT.
Software: KH HPF RS FU JT.
Supervision: HPF US JT.
Validation: SK CH KH RS FU JT.
Visualization: SK CH KH JT.
Writing – original draft: SK CH US JT.
Writing – review & editing: SK CH KH RS FU HPF CPS US JT.

References


