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Ibuprofen reduces zebrafish PGE$_2$ levels but steroid hormone levels and reproductive parameters are not affected

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A B S T R A C T

Prostaglandins are important regulators of reproductive function in fish. Analgesics like aspirin and ibuprofen are prostaglandin inhibitors and have been detected in freshwater systems at ng–μg/L levels. We investigated whether ibuprofen would affect prostaglandin and sex steroid hormone levels in adult zebrafish (Danio rerio) and if expression levels of genes involved in steroidogenesis and prostaglandin synthesis were affected. Zebrafish were exposed to moderate concentrations of ibuprofen (21, 201 or 506 μg/L) for 7 days in a semi-static test system. Ibuprofen concentrations were close to nominal levels and decreased by a maximum of 12–13% over 24 h. Prostaglandin E$_2$ (PGE$_2$) levels in whole body homogenates of males and ovaries of females decreased in a monotonic dose–response relationship whereas male 11-ketotestosterone levels and ovarian 17β-estradiol levels remained unchanged. Ibuprofen did not have an influence on vitellogenin levels, female gonadosomatic index or cumulative egg production and no dose–response relationship in ovarian and testicular expression levels of the investigated genes was observed. This study shows that ibuprofen reduces PGE$_2$ levels in male and female zebrafish but has no consistent effects on other investigated reproductive parameters.

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1. Introduction

Human pharmaceuticals are excreted and eliminated into wastewaters either as the parent compound or as metabolites. Removal rates of pharmaceuticals in sewage treatment plants (STPs) are generally high although not 100%. Hence, persistent or highly consumed pharmaceuticals like analgesics have been detected in aquatic systems that receive water from STPs or untreated wastewater from point sources and consequently fish living in receiving waters are exposed to these chemicals (Ankley et al., 2007). Ibuprofen is a commonly used over-the-counter analgesic and has been detected in final effluent from STPs at concentrations up to 25.4 μg/L (Metcalfe et al., 2003; Camacho-Munoz et al., 2010). Little is known about how exposure to analgesics affects reproductive processes of aquatic animals.

Several studies have shown that fish are more sensitive than mammals or invertebrates to environmental estrogens such as 17α-ethinylestradiol (EE2) (Segner et al., 2003; Ankley et al., 2007) and that might also apply for environmental pharmaceuticals. To date, only a few laboratory studies have tested the effect of analgesics on fish. Ibuprofen (1–100 μg/L) caused increased mortality, limb and organ malformations and behavioral disruptions in exposed zebrafish fry (David and Pancharatna, 2009) and egg production in zebrafish was reduced after exposure to indomethacin (Lister and Van Der Kraak, 2008).

Most weak analgesics including ibuprofen are non-steroidal anti-inflammatory drugs (NSAIDs) and they exert their effect by inhibiting the activity of enzymes involved in prostaglandin synthesis (Vane, 1971). Prostaglandins and their fatty acid precursor arachidonic acid are involved in several processes related to vertebrate reproduction including oocyte maturation (Sorbera et al., 2001) and regulation of gonadal steroidogenesis (van der Kraak and Chang, 1990; Wade and Van der Kraak, 1993) and changes in prostaglandin levels could possibly influence other reproduction related processes. Prostaglandins are synthesized continuously through a multi-step pathway initiated by the conversion of arachidonic acid into prostaglandin H$_2$ (PGH$_2$), which serves as substrate for a variety of different enzymes involved in the synthesis of other prostaglandins, prostacyclin and thromboxanes. Arachidonic acid is mobilized from the cell membrane by phospholipases A$_2$ (PLA$_2$) and the conversion of arachidonic acid into PGH$_2$ is catalyzed by the bifunctional enzyme prostaglandin H$_2$ synthase (PGHS) also known as cyclooxygenase (COX). PGHS exists in two isoforms called COX-1 (ptgs1) and COX-2 (ptgs2) and these enzymes are the targets of NSAIDs (FitzGerald and Patrono, 2001). Functional genes for both COX-1 and COX-2 have been found in zebrafish and prostaglandin E$_2$ (PGE$_2$) is the predominant prostaglandin in adult zebrafish (Grosser et al., 2002; Ishikawa et al., 2007). Prostaglandin inhibitors like indomethacin reduced the PGE$_2$ concentration in female zebrafish (Lister and Van Der Kraak, 2008) and both prostaglandins and NSAIDs are known to affect steroidogenesis in vitro (Wade and Van Der Kraak, 1993).
ibuprofen in zebra fi (Didolkar et al., 1980, 1981) and prenatal exposure to prostaglandin inhibitors blocked masculinization of male fetal mice (Gupta and Goldman, 1986). Given the possible cross talk between prostaglandin and steroidogenic pathways, it was interesting to investigate if an inhibition of prostaglandin synthesis would affect the steroidogenic pathway and reproductive parameters in fish.

The aim of the present study was to investigate if ibuprofen interfered with steroidogenesis in zebrafish (Danio rerio) by measuring 11-ketotestosterone (11-KT) and 17β-estradiol (E2) levels in males and females, respectively, after a 7-day exposure period. Expression levels of genes involved in both prostaglandin synthesis (ptgs1, ptgs2 and cpla2) and steroidogenesis (star, cyp19a, 11β-hsd2, cyp11β and 17β-hsd3) were investigated in order to reveal possible sites of action of ibuprofen in zebrafish (Table 1). PGE2 levels, vitellogenin concentration, spawning success as measured by the numbers of eggs spawned and female gonadosomatic index (GSI) were also determined.

2. Materials and methods

2.1. Animals and experimental design

Zebrafish (D. rerio) were purchased from DAP International (Etobicoke, ON, Canada) and held at the Hagen Aqualab at the University of Guelph in an A-HAB fish containment unit (Aquatic Habitats, Apopka, FL, USA) with re-circulated well water at 26 °C. The photoperiod was 12 h light:12 h dark. Fish were acclimatized for one week in the test aquaria and fed three times daily: twice with commercial salmon fry pellets (Martin Mills, Elmira, ON, Canada) and once with bloodworms (Oregon Desert Brine Shrimp Co., Lakeview, OR, USA).

The exposure was performed as a 7-day semi-static exposure in 20 L glass aquaria each containing 15 L of water. The water was changed every day and water temperature and oxygen saturation were measured every second day. Each aquarium was aerated and the oxygen level was 91%±1.04 (mean±SEM) of the air saturation when water was renewed. The nominal ibuprofen concentrations and the temperature was 25.5±0.8 °C. Egg trays with artificial pellets from ovary extractions were re-suspended in 30 μL of a 10 times diluted glycogen solution (Roche, 20 mg/mL) and 0.1 g/L (11-KT) and 20, 200 and 500 μg/L and the temperature was set at 350 °C. The nebulizer pressure was set at 35 psi and capillary voltage at 4000 V. Fragmentor voltage was 90 V and collision energy 15 V. The mass-to-charge-ratio (m/z) of precursor ion and quantifier ion was respectively 224.3 and 161.2. The detection limit (signal-to-noise-ratio > 10) for ibuprofen in sample matrix was 11.7 pg on column (0.39 pg/μL).

2.3. Sampling

After seven days of exposure the fish were anesthetized in MS-222 (0.1 g/L), blot dried on KIM™ wipes, and weighed. Individuals used for gene expression had their head and tail separated from the trunk with cuts made right behind the pectoral fins and right behind the dorsal fin, respectively. The head and tail fraction was weighed and immediately put on dry ice and stored at −80 °C until vitellogenin analysis was performed. The ovaries were removed from the trunk and put in RNase-free microcentrifuge tubes, weighed, and frozen in dry ice. The trunks of the males were placed on dry ice with the body cavity opened and intestines removed in order to make the testes easier to locate. The testes were dissected out and placed in RNase-free microcentrifuge tubes. All female fish for gene expression studies were sampled between 10:00 and 11:30 h and all males between 12:00 h and 14:00 h. Male fish for whole body steroid determination were snap frozen in microcentrifuge tubes after the presence of testis had been verified. Ovaries for steroid determination were dissected as described above and GSI was calculated for all female fish by the following equation: GSI = (gonad mass (mg)/[body mass (mg) − gonad mass (mg)])×100.

2.4. RNA extractions

Ovarian tissue from 6 females and testis tissue from 7 to 8 males per aquarium were used for gene expression studies. Gonad tissue was homogenized with a handheld homogenizer and RNA was extracted from the tissue with Trizol® (Invitrogen, Carlsbad, CA, USA) according to manufacturer’s instructions. Chloroform and isopropanol were purchased from Sigma-Aldrich Co. During extraction of testes 3.75 μL of a 10 times diluted glycogen solution (Roche, 20 mg/mL) was added to the aqueous phase to aid in RNA precipitation. The RNA pellets from ovary extractions were re-suspended in 30–50 μL GIBCO™ water (Invitrogen) whereas the volume used for testes was 10 μL.

RNA concentrations were quantified on a NanoDrop-8000 spectrophotometer (Agilent Technologies, Missisauga, ON, Canada) and diluted to 1000 ng/μL (females) or 800 ng/μL (males). RNA quality of each sample was determined by the ratio of absorbance at 260 nm and 280 nm. Further, RNA integrity of a subset of RNA samples was assessed using an Agilent 2100 Bioanalyzer and a RNA 6000 Nano LabChip kit (Agilent Technologies). Before reverse transcription (RT) the samples were quantified again with the NanoDrop-8000 spectrophotometer and DNase treated by mixing each sample with 2 μL DNase I (AMP-D1; Sigma-Aldrich) according to manufacturer’s instructions. Primers were designed according to Ings and Van Der Kraak (2006) and the sequences are shown in Table 2. In order to prevent genomic DNA amplification primers were designed to span an exon–exon boundary in the mRNA sequence.

2.5. Reverse transcription and real-time PCR (RT-PCR)

For the reverse transcription (RT) reaction a total of 2 and 1.6 μg RNA for each sample was used for females and males, respectively. RT water controls were prepared by adding 2 μL of GIBCO™ water instead of sample. The samples were incubated for 5 min at 70 °C with 2 μL (0.1 μg) random primer (Promega, Madison, WI, USA) per tube and immediately transferred to ice. The total reaction volume was...
brought to 25 μL and contained the following final concentrations: 5× RT first strand buffer (50 mM Tris–HCl, 75 mM KCl, 3 mM MgCl2; Invitrogen), DTT (10 mM; Invitrogen), dNTPs (0.5 mM; Roche Molecular Biochemicals, Laval, PQ, Canada), M-MLV reverse transcriptase (200 U; Invitrogen) and Gibco™ water. The reaction was performed at 37 °C for 1 h followed by 5 min at 90 °C and subsequently cooling down to 4 °C.

qPCR was performed using ABI Prism 7000 sequence detection system (Applied Biosystems, Foster City, CA, USA) and each sample was run in duplicate. The following was added to each reaction: 5 μL cDNA template (diluted 5, 10 or 20× depending on gene of interest), 2.5 μL of both forward and reverse primers (0.4 μM) and 10 μL SYBR® Green PCR Master Mix. The qPCR cycling conditions were: 2 min at 50 °C and 10 min at 95 °C followed by 40 cycles of 15 s at 95 °C and 1 min at 60 °C (Lister and Van Der Kraak, 2009). To demonstrate primer specificity and amplification of a single product, a melting curve analysis was applied to all reactions.

### 2.6. Determination of steroid hormones and PGE2

Six females and six males from each aquarium were used for steroid and PGE2 measurements. Extraction of steroids was performed on dissected ovaries and whole body homogenate of males. The ovary tissue was sonicated after addition of 100 μL of a 0.1 M PBS buffer (Na2HPO4, NaH2PO4 and NaCl; pH 7.4) with 1 mM EDTA and 10 μM indomethacin. Whole body male fish were pulverized in a chilled ceramic mortar filled with liquid nitrogen and the homogenate was mixed with 4× homogenate weight (v/v) of PBS buffer and then sonicated.

Ovaries and whole body homogenates were extracted with 400 μL or 4 mL methanol respectively, vortexed and placed at 4 °C for 1 h and then centrifuged at 3000 g for 5 min at 4 °C. The pellet was frozen with dry ice and the upper methanol phase decanted in to a test tube. This procedure was repeated twice with 400 μL (ovaries) or 2 mL (males) methanol and the incubation time reduced to 30 min. The three methanol phases were combined and dried down under a stream of nitrogen or in a TurboVap® (Caliper Life Sciences, Hopkinton, MA, USA). The extract was reconstituted in 300 μL (ovaries) or 4× homogenate weight (males) of a 50 mM acetate buffer (glacial acetic acid and sodium acetate; pH 4.0). The sample extract was passed through 100 mg/ml Amprep C-18 octadecyl mini-columns (Amersham Biosciences, Little Chalfont, England) according to the manufacturer's instructions for non-polar analytes whereas a fraction (250 μL) of all sample extract from the males was passed through 30 mg/ml Strata™-X columns (Phenomenex, Denmark) according to manufacturer's instructions for reversed phase solid phase extraction of neutral compounds. Ethyl acetate (1% methanol) was used to collect the final elute from ovaries and 50:50 methanol/acetonitrile (1% acetic acid) was used to elute the male samples. This fraction was dried down under a stream of nitrogen or in a TurboVap®. Ovary samples were reconstituted in 300 μL EIA buffer whereas males were reconstituted in 1000 μL E2, 11-KT and PGE2 levels were analyzed by enzyme immuno assays (EIA; Cayman Chemical, Ann Arbor, MI, USA) as per manufacturer's instructions. Plate development times for E2, 11-KT and PGE2 were 120, 120 and 90–120 min respectively.

### 2.7. Vitellogenin by ELISA

Homogenization and preparation of the samples for vitellogenin analysis was performed according to Kinnberg et al. (2007). Direct non-competitive sandwich ELISA was used to quantify vitellogenin concentrations in the supernatant of the sample homogenate according to Holbech et al. (2001) with the modifications described in Morthorst et al. (2010).

### 2.8. Data analysis and statistics

Prior to analyses all data sets were screened for homogeneity of variance and normality and if necessary data were log transformed. To make sure the replicates in each exposure group did not differ from each other a one-way ANOVA was performed before values from the replicates were pooled. A Bonferroni–Holm corrected one-way ANOVA was performed to compare the exposure groups with the control. The software package SigmaStat® Statistical Software version 2.0 was used in all statistical analyses and statistical differences were considered significant if p<0.05.

### 3. Results

#### 3.1. Chemical analysis of ibuprofen concentrations in water

The measured concentrations of ibuprofen during the 24-hour period between water renewals are shown in Table 3. The variation between the replicates within a treatment was minimal. Ibuprofen concentrations were constant throughout the experiment and the

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**Table 1**

List of investigated genes and their abbreviations. The pathway in which the genes are active is also listed as well as the tissue they were measured in. (AAs: arachidonic acid).

<table>
<thead>
<tr>
<th>Name</th>
<th>Abbreviation</th>
<th>Pathway</th>
<th>Investigated tissue</th>
</tr>
</thead>
<tbody>
<tr>
<td>Elongation factor-1 α</td>
<td>ef1a</td>
<td>Reference gene</td>
<td>Ovaries and testes</td>
</tr>
<tr>
<td>Cytosolic phospholipase A2</td>
<td>cpla1</td>
<td>AA cascade</td>
<td>Ovaries</td>
</tr>
<tr>
<td>Prostaglandin endoperoxide synthase 1</td>
<td>pgs1</td>
<td>AA cascade</td>
<td>Ovaries</td>
</tr>
<tr>
<td>Prostaglandin endoperoxide synthase 2</td>
<td>pgs2</td>
<td>AA cascade</td>
<td>Ovaries</td>
</tr>
<tr>
<td>Aromatase cyp19a</td>
<td>cyp19a</td>
<td>Steroidogenesis</td>
<td>Ovaries</td>
</tr>
<tr>
<td>Steroidogenic acute regulatory protein</td>
<td>star</td>
<td>Steroidogenesis</td>
<td>Ovaries and testes</td>
</tr>
<tr>
<td>17β-hydroxydehydrogenase type 2</td>
<td>17β-hsd2</td>
<td>Steroidogenesis</td>
<td>Testes</td>
</tr>
<tr>
<td>17β-hydroxydehydrogenase type 3</td>
<td>17β-hsd3</td>
<td>Steroidogenesis</td>
<td>Testes</td>
</tr>
<tr>
<td>Cytochrome P450 family 11 β</td>
<td>cyp11β</td>
<td>Steroidogenesis</td>
<td>Testes</td>
</tr>
</tbody>
</table>

---

**Table 2**

Primer sequences used in real-time PCR assays.

<table>
<thead>
<tr>
<th>Gene</th>
<th>Primer sequence</th>
<th>Accession no.</th>
</tr>
</thead>
<tbody>
<tr>
<td>ef1a</td>
<td>Forward GATCACTGTTACTCTTCCAGGTGA</td>
<td>NM_131263</td>
</tr>
<tr>
<td></td>
<td>Reverse GCCGAAAGGCGACAGCCGC</td>
<td></td>
</tr>
<tr>
<td>pgs1</td>
<td>Forward AAGTCTGTTCCAGGCAAGTG</td>
<td>NM_153656.1</td>
</tr>
<tr>
<td></td>
<td>Reverse CAAGAAGATTCCTAAATGTCGCA</td>
<td></td>
</tr>
<tr>
<td>pgs2</td>
<td>Forward GTTAAAGTGGGAAACCTTATACCGAGG</td>
<td>NM_153657.1</td>
</tr>
<tr>
<td></td>
<td>Reverse GGCTACACCTACCATACCAACA</td>
<td></td>
</tr>
<tr>
<td>star</td>
<td>Forward ACCCTTGTTCCTGGCTGATG</td>
<td>NM_131663</td>
</tr>
<tr>
<td></td>
<td>Reverse GGCTCTTTTTCCAGGTTCCTTC</td>
<td></td>
</tr>
<tr>
<td>cyp19a</td>
<td>Forward AGTCCACCGCGACAGCG</td>
<td>NM_131154</td>
</tr>
<tr>
<td></td>
<td>Reverse AGCTCTCCGTCGTCGTCG</td>
<td></td>
</tr>
<tr>
<td>cpla2</td>
<td>Forward TTCTCTGCTGTCGTCG</td>
<td>NM_131295.1</td>
</tr>
<tr>
<td></td>
<td>Reverse TCTCGGTGTCGTCGTCGAAACG</td>
<td></td>
</tr>
<tr>
<td>11β-hsd2</td>
<td>Forward ACCCTACCGCTATACACGGGCAAGGC</td>
<td>NM_212720</td>
</tr>
<tr>
<td></td>
<td>Reverse TTGGGAGAACGCGTACCTGAGCA</td>
<td></td>
</tr>
<tr>
<td>17β-hsd3</td>
<td>Forward ATGTCTCATCCACGCTGAGCA</td>
<td>AY551081</td>
</tr>
<tr>
<td></td>
<td>Reverse CTGACAGATTCCTCCCGAG</td>
<td></td>
</tr>
<tr>
<td>cyp11β</td>
<td>Forward GCAATCACCCGCTACAGGGTACCTC</td>
<td>NM_001080204</td>
</tr>
<tr>
<td></td>
<td>Reverse CACCTCGACAGCGTCC</td>
<td></td>
</tr>
</tbody>
</table>
3.2. Steroid and PGE\textsubscript{2} concentrations

Whole body male and ovarian PGE\textsubscript{2} levels in the control groups were in the range 25–85 and 11 to 45 pg/mg tissue, respectively. PGE\textsubscript{2} levels in both males and females decreased in a monotonic dose–response relationship in response to ibuprofen (Fig. 1A and B). The male PGE\textsubscript{2} levels were significantly different from the control at all exposure concentrations whereas ovarian PGE\textsubscript{2} levels were only significantly different at 506 pg/mg ovary. Whole body 11-KT levels in control males ranged from 0.34 to 5.77 pg/mg tissue and ovarian E\textsubscript{2} were significantly different at 506 pg/mg ovary. No significant difference in PGE\textsubscript{2} levels was observed in any of the treatment groups (Fig. 2A and B).

3.3. Ovarian and testicular gene expressions

A significant decrease in ovarian expression levels of cpla2 was found in the intermediate exposure group (201 µg/L) (Fig. S2). The expression levels of ef1a among the different groups were not significantly different; however, the median expression level in the intermediate group was higher compared to the other groups (Fig. S2).

No effect was observed on the expression levels of the remaining investigated genes in ovaries (ptgs1, ptgs2, star and cypl9a) and testes (cyp11f1, 11β-hsd2, 17β-hsd3, star and ef1a) (Figs. S2 and S3).

3.4. Vitellogenin analysis

Vitellogenin levels in exposed males and females were not significantly different from the control (Fig. 3A and B). Egg production was not affected at any of the exposure concentrations (Fig. 4). GSI of the females was not affected in a significant way but a slight increase in GSI was observed in the high exposure group (Fig. 5).

4. Discussion

Due to high consumption rates and insufficient removal in STPs human and veterinary pharmaceuticals have been detected in aquatic ecosystems and aquatic organisms might therefore be continuously exposed to highly consumed pharmaceuticals like analgesics (Ankley et al., 2007). However, only few experiments have investigated the possible effects of analogics on aquatic vertebrates. Mild analgesics inhibit the enzymes responsible for prostaglandin synthesis, cyclooxygenase-1 and -2 (COX-1 and COX-2). Prostaglandins and their precursor arachidonic acid play key roles in vertebrate reproduction for example oocyte maturation, ovulation and regulation of gonadal steroidogenesis (Van Der Kraak and Chang, 1990; Wade and Van Der Kraak, 1993; Sorbera et al., 2001; Patino et al., 2003). Both steroid- and prostaglandin-derived sex pheromones have been identified in fish (Stacey, 2003), and exposure to prostaglandin F\textsubscript{2α}, (PGF\textsubscript{2α}) increases goldfish testosterone levels (Mennigen et al., 2010) suggesting that alterations in prostaglandin levels could influence fish reproduction.

This experiment clearly demonstrates that PGE\textsubscript{2} levels were significantly reduced in male whole body homogenate at all exposure concentrations (21–506 µg/L) and in ovaries a similar pattern was observed (Fig. 1). Prostaglandins are involved in induction of both male and female sexual behavior in fish (reviewed by Sorensen and Goetz (1993)) suggesting that prostaglandin inhibitors could affect spawning and fertilization. Despite significantly reduced prostaglandin levels the cumulative egg production (Fig. 4), female GSI (Fig. 5) and number of spawning events (data not shown) were not affected. In a

<table>
<thead>
<tr>
<th>Exposure concentration (µg/L)</th>
<th>PGE\textsubscript{2} concentration (pg/mg homogenate)</th>
<th>PGE\textsubscript{2} concentration (pg/mg ovary)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td>0.0±0.0</td>
<td>0.0±0.0</td>
</tr>
<tr>
<td>20</td>
<td>21±0.5</td>
<td>22.7±0.4</td>
</tr>
<tr>
<td>200</td>
<td>201±7.8</td>
<td>215.5±21.1</td>
</tr>
<tr>
<td>500</td>
<td>506±10.9</td>
<td>547.8±6.1</td>
</tr>
</tbody>
</table>

Fig. 1. Male and female PGE\textsubscript{2} concentrations. PGE\textsubscript{2} concentrations in (A) male whole body homogenate and (B) ovaries of adult zebrafish. Data represent mean±SEM. The number of male and female fish in each exposure group is indicated in the bottom of the figure. * indicates significantly different from the control group (p<0.05).
The number of male and female
1980; Kristensen et al., 2011, 2012)
and ovulation.
can be dif-
cult to assess, as the daily egg production is variable and
affected by temperature, age, spawning strategy etc. (Paull et al., 2008).
Indomethacin caused reduced egg production and reduced PGE2 levels
in female zebra
fish after 16 days of exposure (Lister and Van Der Kraak,
2007). The effect of ibuprofen on spawning in the medaka
could be due to a longer exposure period but also zebra
fish egg produc-
tion can be dif-
ficulty mammalian studies have provided con-
clusion testosterone levels in male mice (Martini et al., 2008) and
gen - dependent yolk precursor vitellogenin is a well-known biomarker for exposure to endocrine
disrupters in fish (Tyler et al., 1999; Holbech et al., 2012). Ibuprofen
did not affect male or female vitellogenin levels (Fig. 3), which suggests
that ibuprofen does not affect steroidogenesis through the estrogen
receptor pathway. This is in accordance with results from the yeast
estrogen screen assay (YES assay) in our lab where ibuprofen and other
analgesics failed to induce a response by binding to the human estrogen
receptor (data not published).
Steroids are synthesized through multiple steps and numerous
enzymes are involved in changes in expression levels of involved
enzymes and substrate levels can influence hormone levels. C17, 20-
lyase (also called P450c17 and CYP17) and 11β-hydroxylase (CYP11β)
are involved in androgen synthesis. In an in vitro system with male
carp gonads analgesics impaired the activity of CYP11β and CYP17
Fernandes et al., 2011) but ibuprofen did not influence adult male 11-
KT levels in our study. Enzymes like CYP11β were demonstrated mainly
to be active during early spermatogenesis in sea bass (Fernandes et al.,
2007) and hence exposure to ibuprofen during juvenile life stages
could influence androgen production. Expression levels of the following
genes directly or indirectly involved in androgen synthesis were inves-
tigated in testes: 11β-hsd2, 17β-hsd3, cyp11b and star but expression of
these genes was not affected.

Fig. 2. Male and female 11-KT and E2 concentrations. (A) 11-KT concentrations in male whole body homogenate and (B) E2 concentrations in ovaries. Data represent mean±SEM. The number of male and female fish in each exposure group is indicated in the bottom of the figure.

Fig. 3. Male and female vitellogenin concentrations. Vitellogenin concentrations in (A) males and (B) females. Four outlying male values were removed. Data represent mean±SEM. The number of male or female fish in each exposure group is indicated in the bottom of the figure.
Both cpla2 and ptgs2 are involved in prostaglandin synthesis and ovarian expression levels of those genes fluctuate during the ovulatory cycle of spawning zebrafish whereas expression of pgs1 remains fairly stable (Lister and Van Der Kraak, 2009). In this experiment ovarian expression of cpla2 in the intermediate exposure group (201 μg/L) was significantly different from the control (Fig. S2). Zebrafish ovarian development is asynchronous with follicles in different stages. The proportions of follicles in different stages vary between individuals and could explain the significant difference of cpla2 expression even though sampling occurred at the same time every day. Ibuprofen inhibits the COX enzymes by substrate competition and a response to overcome sampling occurred at the same time every day. Ibuprofen inhibits the COX enzyme catalyzing the conversion of androgens to estrogens. Arachidonic acid and its metabolites are involved in regulation of StAR, the enzyme responsible for transport of cholesterol from the outer to the inner mitochondrial membrane (Wang et al., 2003) and hence changes in prostaglandin levels could also influence cholesterol transport and thereby steroid synthesis. However, ibuprofen did not influence female star and cyp19a or male star gene expression (Fig. S2) and also not 11-KT and E2 levels. Ibuprofen significantly increased both E2 production and aromatase activity by up-regulation of mRNA transcription in H295R cells (Han et al., 2010) and a tendency to decreased testosterone production was also observed, which could be due to increased aromatase activity. Those abovementioned equivocal results suggest that NSAIDs may influence vertebrate species differently and illustrate that further investigations are needed to understand the impact of NSAIDs on aquatic wildlife.

Prostaglandins are involved in many pathways and processes and their synthesis is regulated through different pathways. In the present experiment we observed a significant effect of ibuprofen on PGE2 levels at concentrations also detected in effluent from STPs (Metcalfe et al., 2003; Camacho-Munoz et al., 2010) but no consistent effects on other reproductive endpoints were found. Prostaglandins have been shown to affect several sperm parameters in mammals and a prolonged reduction of prostaglandin levels could lead to more severe effects due to changes in fertilization success. More research is needed in the field of secondary effects of prostaglandin disruption in fish, as wild fish populations are exposed to numerous pharmaceuticals including several NSAIDs in concentrations able to affect endogenous prostaglandin levels.

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Appendix A. Supplementary data

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References


